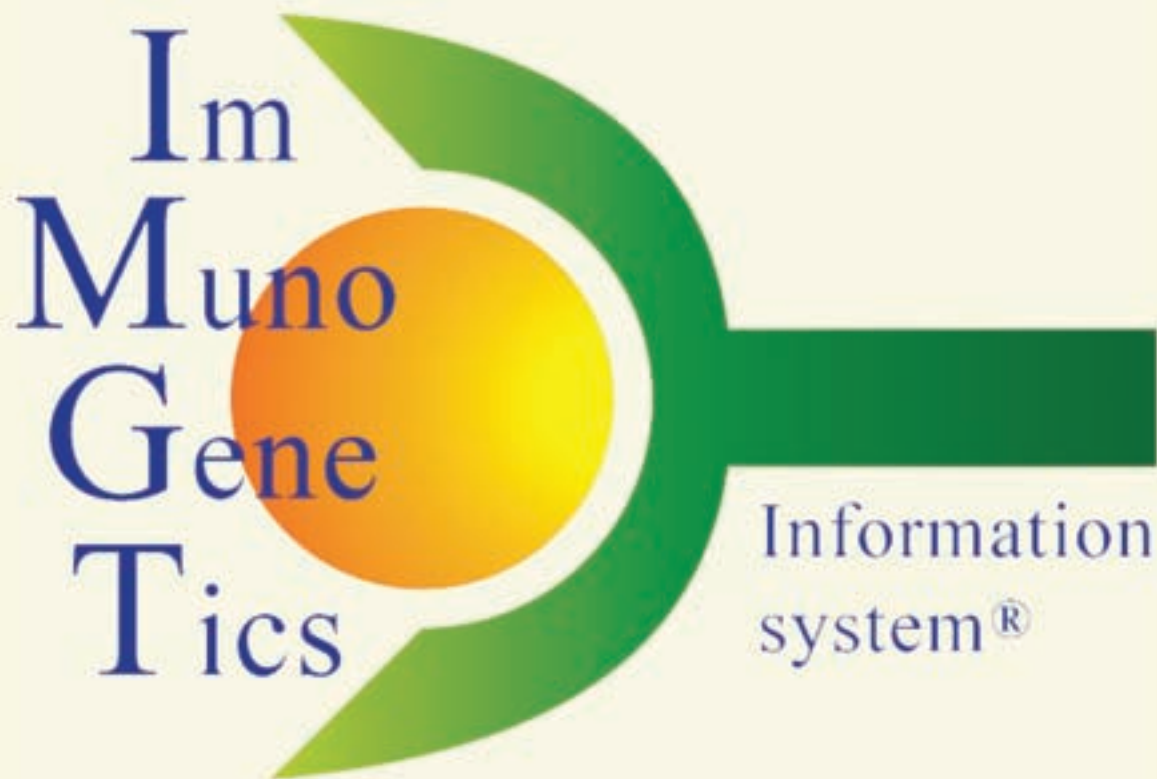
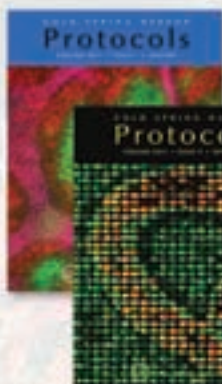


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# IMGT, the International ImMunoGeneTics Information System

Marie-Paule Lefranc

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## INTRODUCTION

IMGT, the international ImMunoGeneTics information system (<http://www.imgt.org>), created in 1989 by the Laboratoire d'ImmunoGénétique Moléculaire LIGM (Université Montpellier 2 and CNRS, Montpellier, France), is the global reference in immunogenetics and immunoinformatics. IMGT-ONTOLOGY concepts, generated from the Formal IMGT-ONTOLOGY or IMGT-Kaleidoscope axioms, have allowed us to set up the IMGT Scientific chart rules and to standardize and manage the complexity of immunogenetics data. IMGT is a high-quality integrated knowledge resource specializing in the immunoglobulins (IG) or antibodies, T cell receptors (TR), and major histocompatibility (MH) of human and other vertebrate species, proteins of the immunoglobulin superfamily (IgSF) and MH superfamily (MhSF), related proteins of the immune systems (RPI) of vertebrates and invertebrates, therapeutic monoclonal antibodies (mAb), and fusion proteins for immune applications (FPIA). IMGT provides a common access to standardized data from genome, proteome, genetics, and three-dimensional (3D) structures. IMGT consists of databases, interactive online tools, and Web resources.

## RELATED INFORMATION

IMGT is freely available at <http://www.imgt.org>, which provides documentation for IMGT databases and tools. IMGT-ONTOLOGY concepts of identification are available at the National Center for Biomedical Ontology (NCBO) BioPortal <http://bioportal.bioontology.org/> and at IMGT <http://www.imgt.org>.

Information is also available in **From IMGT-ONTOLOGY IDENTIFICATION Axiom to IMGT Standardized Keywords: For Immunoglobulins (IG), T Cell Receptors (TR) and Conventional Genes** (Lefranc 2011a), **From IMGT-ONTOLOGY DESCRIPTION Axiom to IMGT Standardized Labels: For Immunoglobulin (IG) and T Cell Receptor (TR) Sequences and Structures** (Lefranc 2011b), **From IMGT-ONTOLOGY CLASSIFICATION Axiom to IMGT Standardized Gene and Allele Nomenclature: For Immunoglobulins (IG) and T Cell Receptors (TR)** (Lefranc 2011c), **IMGT Unique Numbering for the Variable (V), Constant (C), and Groove (G) Domains of IG, TR, MH, IgSF, and MhSF** (Lefranc 2011d), and **IMGT Collier de Perles for the Variable (V), Constant (C), and Groove (G) Domains of IG, TR, MH, IgSF, and MhSF** (Lefranc 2011e).

In addition, protocols are available for **IMGT/V-QUEST: IMGT Standardized Analysis of the Immunoglobulin (IG) and T Cell Receptor (TR) Nucleotide Sequences** (Giudicelli et al. 2011), **IMGT/JunctionAnalysis: IMGT Standardized Analysis of the V-J and V-D-J Junctions of the Rearranged Immunoglobulins (IG) and T Cell Receptors (TR)** (Giudicelli and Lefranc 2011), **IMGT/Collier de Perles: IMGT Standardized Representation of Domains (IG, TR, and IgSF Variable and Constant Domains, MH and MhSF Groove Domains)** (Ehrenmann et al. 2011), **IMGT/DomainGapAlign: IMGT Standardized Analysis of Amino Acid Sequences of Variable, Constant, and Groove Domains (IG, TR, MH, IgSF, MhSF)** (Ehrenmann and Lefranc 2011a), and **IMGT/3Dstructure-DB: Querying the IMGT Database for 3D Structures in Immunology and Immunoinformatics (IG or Antibodies, TR, MH, RPI, and FPIA)** (Ehrenmann and Lefranc 2011b).

## BACKGROUND INFORMATION

The amount of genomics, genetics, 3D, and functional data published in the immunogenetics field is growing exponentially and involves fundamental, clinical, veterinary, and pharmaceutical research.

The number of potential protein forms of the antigen receptors, IG or antibodies, and TR is almost unlimited. The potential repertoire of each individual is estimated to comprise about  $2 \times 10^{12}$  different IG and TR, and the limiting factor is only the number of B and T cells that an organism is genetically programmed to produce. This huge diversity is inherent to the particularly complex and unique molecular synthesis and genetics of the antigen receptor chains (Fig. 1). This includes biological mechanisms such as DNA molecular rearrangements (combinatorial diversity) in multiple loci (three for IG and four for TR in humans) located on different chromosomes (four in humans), nucleotide deletions and insertions at the rearrangement junctions (or N-diversity), and somatic hypermutations in the IG loci (for review, see Lefranc and Lefranc 2001a,b).

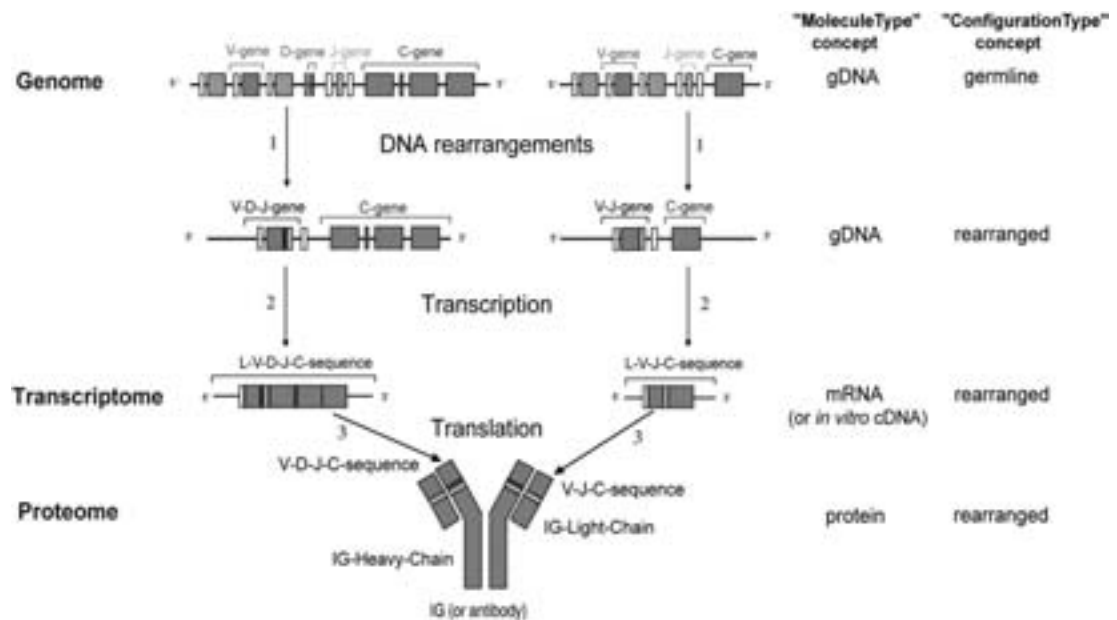
IMGT, the international ImMunoGeneTics information system (<http://www.imgt.org>) (Lefranc et al. 2009), was created in 1989 by Marie-Paule Lefranc, Laboratoire d'ImmunoGénétique Moléculaire LIGM (Université Montpellier 2 and CNRS) at Montpellier, France, in order to standardize and to manage the complexity of immunogenetics data. IMGT has reached that goal through the building of a unique ontology, IMGT-ONTOLOGY (Giudicelli and Lefranc 1999; Lefranc et al. 2004, 2005a, 2008; Duroux et al. 2008), the first ontology in immunogenetics and immunoinformatics.

## IMGT-ONTOLOGY

IMGT-ONTOLOGY (Giudicelli and Lefranc 1999; Lefranc et al. 2004, 2005a; 2008; Duroux et al. 2008), the first ontology for immunogenetics and immunoinformatics, has been built to ensure the accuracy and the consistency of the IMGT data as well as the coherence between the components (databases, tools, and Web resources) of IMGT, the international ImMunoGeneTics information system (<http://www.imgt.org>). IMGT-ONTOLOGY is now acknowledged as the global reference in immunogenetics and immunoinformatics, allowing IMGT to bridge biological and computational spheres in bioinformatics (Lefranc et al. 2008).

## IMGT-ONTOLOGY KNOWLEDGE DOMAIN

An ontology is a formal representation of a knowledge domain (Gruber 1993; Giudicelli and Lefranc 1999). The knowledge domain covered by IMGT-ONTOLOGY is immunogenetics and immunoinformatics.



**FIGURE 1.** Synthesis of an IG or antibody in humans. A human being may potentially synthesize  $10^{12}$  different antibodies (Lefranc and Lefranc 2001a). (1) DNA rearrangements (*is\_rearranged\_into*), (2) Transcription (*is\_transcribed\_into*), (3) Translation (*is\_translated\_into*). The configuration of C-GENE is "undefined" (ConfigurationType) (IMGT Repertoire, <http://www/imgt.org>).



Immunogenetics studies the genetics of immune responses. In addition to the innate immune response present in all living organisms, vertebrates with jaws (or gnathostomata, from fish to humans) have acquired the adaptative immune response, characterized by an extreme diversity of the specific antigen receptors that comprise the IG or antibodies and the TR ( $10^{12}$  different IG and  $10^{12}$  different TR per individual, in humans). Complex molecular mechanisms in B cells (Fig. 1) and in T cells are at the origin of this huge diversity (Lefranc and Lefranc 2001a,b). These mechanisms include, in particular, DNA rearrangements and, for the IG, somatic hypermutations. In addition, there is considerable polymorphism of the MH proteins (human leucocyte antigens [HLA] in humans). These particularities of the adaptive immune system of the vertebrates, and a better knowledge of the innate immune responses found in any species, have made the immune system an excellent model for systems biology.

## IMGT-ONTOLOGY AXIOMS AND CONCEPTS

IMGT-ONTOLOGY manages the immunogenetics knowledge through diverse facets that rely on the seven axioms of the Formal IMGT-ONTOLOGY or IMGT-Kaleidoscope: "IDENTIFICATION," "DESCRIPTION," "CLASSIFICATION," "NUMEROTATION," "LOCALIZATION," "ORIENTATION," and "OBTENTION" (Giudicelli and Lefranc 1999; Lefranc et al. 2004, 2005a, 2008; Duroux et al. 2008). These axioms postulate that any object, any process, and any relation has to be identified, described, classified, numbered, localized, and oriented, and that the way it is obtained can be characterized (Duroux et al. 2008; Lefranc et al. 2008) (Fig. 2).

The concepts generated from these axioms have been essential for establishing the IMGT Scientific chart rules for the genome, proteome, genetics, two-dimensional (2D) and 3D structures (Lefranc 2005, 2008b,c). The IMGT-ONTOLOGY axioms, the concepts generated from them, examples of concepts, and IMGT Scientific chart rules are listed in Table 1.

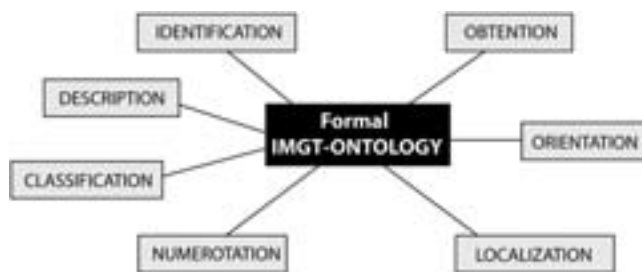
As an example, the NUMEROTATION axiom has generated two major concepts of numerotation:

- the IMGT unique numbering for the variable (V) domain (Lefranc 1997, 1999; Lefranc et al. 2003) and the constant (C) domain (Lefranc et al. 2005c) of the IG, TR, and IgSF, and for the groove (G) domain (Lefranc et al. 2005b) of the MH and MhSF.
- the IMGT Colliers de Perles for V, C, and G domains or graphical 2D representations of the domains that bridge the gap between sequences and 3D structures (Ruiz and Lefranc 2002; Garapati and Lefranc 2007; Kaas and Lefranc 2007; Kaas et al. 2007).

The IMGT Scientific chart rules based on the concepts of identification, description, classification, and numerotation have been essential in providing the standards currently used in immunogenetics and immunoinformatics: standardized keywords, prototypes and standardized labels, official nomenclature of the IG and TR genes and alleles (Lefranc 2000a,b; Lefranc and Lefranc 2001a,b), framework region (FR)-IMGT and complementarity determining region (CDR)-IMGT delimitations, contact analysis and paratope/epitope characterization, etc. (see Table 2).

## IMGT DATABASES, TOOLS, AND WEB RESOURCES

Based on the IMGT-ONTOLOGY concepts, IMGT is a high-quality integrated knowledge resource, specializing in the IG, TR, and MH of human and other vertebrates, proteins of the immunoglobulin



**FIGURE 2.** The seven axioms of the Formal IMGT-ONTOLOGY or IMGT-Kaleidoscope. (Modified from Duroux et al. 2008 and reprinted with permission from Elsevier © 2008.)

**Table 1. Axioms of the formal IMGT-ONTOLOGY (or IMGT-Kaleidoscope), concepts generated from them, examples of concepts, and IMGT Scientific chart rules**

Formal IMGT-ONTOLOGY axioms	Concepts generated from the axioms	Examples of IMGT-ONTOLOGY concepts	IMGT Scientific chart rules <sup>a</sup>
IDENTIFICATION axiom	Concepts of identification	ChainType <sup>b</sup> ConfigurationType DomainType FunctionalityType FunctionType GeneType LocationType Molecule_EntityType MoleculeType MoleculeUnit ReceptorType <sup>b</sup> SpecificityType StructureType TaxonRank <sup>b</sup>	Standardized keywords
DESCRIPTION axiom	Concepts of description	Core Molecule_EntityPrototype Recombination signal (RS)	Prototypes Standardized labels
CLASSIFICATION axiom	Concepts of classification	Group Subgroup Gene Allele	Gene and allele nomenclature
NUMEROTATION axiom	Concepts of numerotation	IMGT_unique_numbering (IMGT unique numbering) for V, C and G IMGT_Collier_de_Perles (IMGT Collier de Perles) for V, C and G domains	FR-IMGT and CDR-IMGT delimitations Contact analysis Paratope/epitope characterization Conserved positions
LOCALIZATION axiom	Concepts of localization	Locus Chromosome Cell compartment Organ part Organ	Locus representation Chromosomal localization Standardized keywords
ORIENTATION axiom	Concepts of orientation	DNA_strand_orientation Genomic_orientation <sup>b</sup>	IMGT Index
OBTENTION axiom	Concepts of obtention	Origin Methodology	Standardized keywords

<sup>a</sup>The corresponding controlled vocabulary and rules are available in the IMGT Scientific Chart at <http://www.imgt.org>.

<sup>b</sup>Indicates a high concept, that is, a concept with different levels of granularity.

superfamily (IgSF) and MH superfamily (MhSF), RPI of vertebrates and invertebrates, therapeutic mAbs and FPIA (Lefranc et al. 2009). IMGT provides a common access to standardized data from genome, proteome, genetics, and 3D structures (Lefranc 2005, 2008b,c). The IMGT information system manages immunogenetics knowledge according to three main IMGT biological approaches: genomic, genetic, and structural (Fig. 3). For each approach, IMGT provides databases, interactive online tools, and Web resources (Table 2).

### Genomic Approach

The IMGT genomic approach is gene-centered and mainly oriented toward the study of the genes within their loci and on the chromosomes.

IMGT/GENE-DB (Giudicelli et al. 2005), the IMGT genome database, is the official repository of all of the IG and TR genes and alleles approved by the World Health Organization (WHO)/International Union

**Table 2. IMGT databases, tools, and Web resources for genomic, genetic, and structural approaches**

Approaches	Databases	Tools	Web resources <sup>a</sup>
Genomic	IMGT/GENE-DB	IMGT/GeneView IMGT/LocusView IMGT/CloneSearch IMGT/GeneSearch IMGT/GenelInfo IMGT/LIGMotif IMGT/GeneFrequency	IMGT Repertoire “Locus and genes” section: Chromosomal localizations Locus representations Locus description, etc. Gene tables Potential germline repertoires Lists of genes Correspondence between nomenclatures, etc.
Genetic	IMGT/LIGM-DB IMGT/MHC-DB IMGT/PRIMER-DB IMGT/ 2Dstructure-DB	IMGT/V-QUEST IMGT/JunctionAnalysis IMGT/HighV-QUEST IMGT/BlastSearch IMGT/Allele-Align IMGT/PhyloGene IMGT/DomainDisplay IMGT/DomainGapAlign	IMGT Repertoire “Proteins and alleles” section: Alignments of alleles Tables of alleles Allotypes Isotypes Protein displays, etc.
Structural	IMGT/ 3Dstructure-DB	IMGT/Collier de Perles IMGT/ DomainSuperimpose IMGT/StructuralQuery	IMGT Repertoire “2D and 3D structures” section: IMGT Colliers de Perles FR-IMGT and CDR-IMGT lengths IMGT classes for amino acid characteristics IMGT Colliers de Perles reference profiles 3D representations, etc.

<sup>a</sup>Only Web resources examples from the IMGT Repertoire sections are shown.

of Immunological Societies (IUIS) Nomenclature Subcommittee for IG and TR (Lefranc 2007, 2008a). Reciprocal links exist between IMGT/GENE-DB and the Human Genome Nomenclature Committee (HGNC) database and Gene at the National Center for Biotechnology Information (NCBI).

The IMGT genomics tools manage the locus organization and gene location and provide the display of physical maps for the human and mouse IG, TR, and MH loci. They allow one to view genes in a locus (IMGT/GeneView, IMGT/LocusView), to search for clones (IMGT/CloneSearch), to search for genes in a locus based on IMGT gene names, functionality, or localization on the chromosome (IMGT/GeneSearch, IMGT/GenelInfo) (Baum et al. 2004, 2006), to identify and describe IG and TR genes in large genomic sequences (IMGT/LIGMotif) (Lane et al. 2010), or to provide a graphical representation of the numbers of available sequences containing rearranged IG and TR genes (IMGT/GeneFrequency).

The IMGT genomics resources include chromosomal localizations, locus representations, locus description, gene positions, gene exon/intron organization, gene exon/intron splicing sites, gene tables, potential germline repertoires, lists of genes, and correspondence between nomenclatures (IMGT Repertoire “Locus and genes” section).

## Genetic Approach

The IMGT genetic approach refers to the study of the genes in relation to their sequence polymorphisms and mutations, their expression, their specificity, and their evolution.

The IMGT sequences databases include:

1. IMGT/LIGM-DB (Giudicelli et al. 2006), created in 1989 and on the Web since July 1995. It is the first and the largest IMGT database, containing IG and TR nucleotide sequences and the translation of annotated sequences from human and other vertebrate species (more than 150,000 sequences from 261 species in December 2010).
2. IMGT/MH-DB, hosted at EBI, for the MH sequences of human (HLA) and other vertebrates.





**FIGURE 3.** IMGT (<http://www.imgt.org>) comprises databases (shown as cylinders), tools (shown as rectangles), and Web resources (data not shown, see Table 2 for examples). Interactions between databases and tools in the genomic, genetic, and structural approaches are represented with continuous, dotted, and broken lines. (Modified from Lefranc et al. 2009 and reprinted with permission from Oxford University Press © 2009.)

3. IMGT/PRIMER-DB that contains oligonucleotide sequences used as primers for combinatorial library constructions, single-chain Fragment variable (scFv), phage display, or microarray technologies.
4. IMGT/2Dstructure-DB that contains amino acid sequences of Kabat IG (those only known at the amino acid level) and of therapeutic mAbs and FPIA of the WHO/International Nonproprietary Name (INN) programme (Lefranc 2011f).

The IMGT sequence analysis tools include:

1. IMGT/V-QUEST (Giudicelli et al. 2004; Brochet et al. 2008) for the identification of the variable (V), diversity (D), and joining (J) genes and the description of their mutations in rearranged IG and TR sequences. IMGT/V-QUEST has been approved by the European Research Initiative on chronic lymphocytic leukaemia CLL (ERIC) for the analysis of the IGHV gene mutational status in CLL (Ghia et al. 2007; Giudicelli and Lefranc 2009).
2. IMGT/JunctionAnalysis (Yousfi Monod et al. 2004) for a comprehensive analysis of the V-J and V-D-J junctions that confer the antigen receptor specificity (Bleakley et al. 2006, 2008).
3. IMGT/HighV-QUEST, a high-throughput version of IMGT/V-QUEST, for the analysis of the IG and TR sequences, generated from Next Generation Sequencing (NGS) or deep sequencing, by batches of up to 150,000 sequences (Alamyar et al. 2010).

4. IMGT/BlastSearch for a sequence search against the following databases: IMGT/LIGM-DB, IMGT/GENE-DB reference sequences, IMGT/3Dstructure-DB sequences, IMGT/3Dstructure-DB domain sequences, or IMGT/DomainDisplay reference sequences.
5. IMGT/Allele-Align for the detection of allelic polymorphisms.
6. IMGT/Phylogene (Elemento and Lefranc 2003) for IG and TR gene evolution analyses.
7. IMGT/DomainDisplay (Ehrenmann et al. 2010a,b) for the display of amino acid sequences from the IMGT domain directory.

The IMGT genetics resources include alignments of alleles, tables of alleles, allotypes (Jefferis and Lefranc 2009), isotypes, and protein displays (IMGT Repertoire “Proteins and alleles” section).

### Structural Approach

The IMGT structural approach refers to the study of the 2D and 3D structures and to the antigen- or ligand-binding characteristics in relationship to the protein functions, polymorphisms, and evolution.

IMGT/3Dstructure-DB (Kaas et al. 2004; Ehrenmann et al. 2010b), the IMGT 3D structure database, comprises IG, TR, MH, IgSF, MhSF, RPI, mAb, and FPIA with known 3D structures. The IMGT/3Dstructure-DB cards provide chain details with IMGT annotations (receptor, chain and domain description, IMGT gene and allele names), domain delimitations and IMGT Collier de Perles, contact analysis, downloadable renumbered IMGT/3Dstructure-DB flat files, visualization tools (Jmol and QuickPDB), and external links. IMGT Residue@Position cards provide detailed information on the inter- and intradomain contacts at each residue position, based on the IMGT unique numbering. IMGT/3Dstructure-DB entries containing IG/antigen (IG/Ag) or TR/peptide/MH (TR/pMH) complexes can be queried for standardized analysis of the interactions between paratope/epitope (Kaas and Lefranc 2005; Kaas et al. 2008; Lefranc 2009; Ehrenmann et al. 2010a,b).

The IMGT structural tools include:

1. IMGT/DomainGapAlign (Ehrenmann et al. 2010b) for the analysis of amino acid sequences per domain, including those of the IG and TR V domains.
2. IMGT/Collier de Perles to make your own IMGT Collier de Perles.
3. IMGT/DomainSuperimpose to superimpose two domain 3D structures from IMGT/3Dstructure-DB.
4. IMGT/StructuralQuery (Kaas et al. 2004) to retrieve the IMGT/3Dstructure-DB entries containing a V domain, based on specific structural characteristics of the intramolecular interactions: e.g.,  $\phi$  and  $\psi$  angles, distance in angstrom between amino acids, CDR-IMGT lengths.

The IMGT structural resources include IMGT Colliers de Perles (2D representations on one layer or two layers), FR-IMGT and CDR-IMGT lengths, IMGT classes for amino acid characteristics, and profiles based on the 11 IMGT amino acid physicochemical classes (Pommié et al. 2004) (IMGT Repertoire “2D and 3D structures” section).

### Other IMGT Databases and Web Resources

IMGT/CLL-DB (Brochet 2008) contains IG sequences of CLL patients, analyzed by IMGT/V-QUEST. IMGT/mAb-DB (Poirion et al. 2010) provides information on therapeutic mAbs and FPIA. In addition to the IMGT Scientific chart and IMGT Repertoire, IMGT Web resources comprise IMGT Index, IMGT Bloc-notes, IMGT Education (e.g., IMGT Lexique, Aide-Mémoire, Tutorials), The IMGT Medical page, The IMGT Veterinary page, The IMGT Biotechnology page, The IMGT Immunoinformatics page, and IMGT other accesses.

### IMGT USERS

Since July 1995 IMGT has been available on the Web at the IMGT Home page <http://www.imgt.org>. The knowledge provided by the IMGT information system is of much value to clinicians and biological scientists in general. IMGT databases, tools, and Web resources are extensively queried and used by scientists from both academic and research laboratories from pharmaceutical companies, who are equally

distributed between the United States, Europe, and the remaining world. IMGT is used in very diverse domains: (1) basic research, (2) medical research (vaccine design, repertoire analysis of the IG antibody sites and of the TR recognition sites in normal and pathological situations such as autoimmune diseases, infectious diseases, AIDS, leukemias, lymphomas, and myelomas), (3) veterinary research (immune repertoires in domestic and wildlife species, animal models), (4) genomics (study of the genome diversity and evolution of the adaptive immune response), (5) structural biology (evolution of the domains of the IgSF and MhSF proteins), (6) biotechnology related to antibody engineering and antibody humanization (construction and analysis of scFv, phage displays, combinatorial libraries, chimeric, humanized, and human antibodies), (7) diagnostics (clonalities, detection, and follow-up of minimal residual diseases), and (8) therapeutic approaches (grafts, immunotherapy, and vaccinology).

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# From IMGT-ONTOLOGY IDENTIFICATION Axiom to IMGT Standardized Keywords: For Immunoglobulins (IG), T Cell Receptors (TR), and Conventional Genes

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## INTRODUCTION

IMGT standardized keywords have been defined for the identification of immunoglobulins (IG) and T cell receptors (TR) and conventional genes. These keywords are based on the concepts of identification, generated from the IDENTIFICATION axiom of the Formal IMGT-ONTOLOGY or IMGT-Kaleidoscope that postulates that any molecule, cell, tissue, organ, organism, or population, any process and any relation, has to be identified. Most of the IMGT standardized keywords used for the identification of sequences and structures are names of IMGT-ONTOLOGY leafconcepts (a leafconcept is a concept that corresponds to the finest level of granularity). IMGT standardized keywords have been essential for the data entries into the IMGT databases and tools, and particularly into IMGT/LIGM-DB, the first and the largest IMGT database that contains IG and TR nucleotide sequences and the translation of annotated sequences. IMGT standardized keywords for IG, TR, and conventional genes are defined in the context of the IMGT-ONTOLOGY concepts of identification.

## RELATED INFORMATION

IMGT standardized keywords for sequences of IG, TR, and conventional genes that are used in IMGT/LIGM-DB (Giudicelli et al. 2006) are available at the database query page (IMGT Home page <http://www.imgt.org>). IMGT standardized keywords for amino acid sequences and structures are available in the IMGT Scientific chart at <http://www.imgt.org/textes/IMGTScientificChart/SequenceDescription/IMGT3Dkeywords.html>. IMGT-ONTOLOGY concepts of identification are available at the National Center for Biomedical Ontology (NCBO) BioPortal <http://bioportal.bioontology.org/> and at IMGT <http://www.imgt.org>.

A detailed description of IMGT is provided in **IMGT, The International ImMunoGeneTics Information System** (Lefranc 2011a). Information is also available on **From IMGT-ONTOLOGY DESCRIPTION Axiom to IMGT Standardized Labels: For Immunoglobulin (IG) and T Cell Receptor (TR) Sequences and Structures** (Lefranc 2011b), **From IMGT-ONTOLOGY CLASSIFICATION Axiom to IMGT Standardized Gene and Allele Nomenclature: For Immunoglobulins (IG) and T Cell Receptors (TR)** (Lefranc 2011c), **IMGT Unique Numbering for the Variable (V), Constant (C), and Groove (G) Domains of IG, TR, MH, IgSF, and MhSF** (Lefranc 2011d), and **IMGT Collier de Perles for the Variable (V), Constant (C), and Groove (G) Domains of IG, TR, MH, IgSF, and MhSF** (Lefranc 2011e).

In addition, protocols are available for **IMGT/V-QUEST: IMGT Standardized Analysis of the Immunoglobulin (IG) and T Cell Receptor (TR) Nucleotide Sequences** (Giudicelli et al. 2011), **IMGT/JunctionAnalysis: IMGT Standardized Analysis of the V-J and V-D-J Junctions of the Rearranged Immunoglobulins (IG) and T Cell Receptors (TR)** (Giudicelli and Lefranc 2011), **IMGT/Collier de Perles: IMGT Standardized Representation of Domains (IG, TR, and IgSF Variable and Constant Domains, MH and MhSF Groove Domains)** (Ehrenmann et al. 2011), **IMGT/DomainGapAlign: IMGT Standardized Analysis of Amino Acid Sequences of Variable, Constant, and Groove Domains (IG, TR, MH, IgSF, MhSF)** (Ehrenmann and Lefranc 2011a), and

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## BACKGROUND INFORMATION

The IDENTIFICATION axiom is an axiom of the Formal IMGT-ONTOLOGY or IMGT-Kaleidoscope (Giudicelli and Lefranc 1999; Lefranc et al. 2004, 2005a, 2008; Duroux et al. 2008). The IDENTIFICATION axiom postulates that any molecule, cell, tissue, organ, organism, or population, any process and any relation, has to be identified. The IDENTIFICATION axiom has generated the concepts of identification of IMGT-ONTOLOGY, the global reference in immunogenetics and immunoinformatics, built by IMGT, the international ImMunoGeneTics information system (<http://www.imgt.org>; Lefranc et al. 2009). The IDENTIFICATION axiom is one of seven axioms of the Formal IMGT-ONTOLOGY, the others being "CLASSIFICATION axiom," "DESCRIPTION axiom," "NUMEROTATION axiom," "LOCALIZATION axiom," "ORIENTATION axiom," and "OBTENTION axiom." Keywords used in the IMGT databases and tools mostly derive from the concepts of identification generated from the IDENTIFICATION axiom, and among them, most particularly the following: "MoleculeType," "GeneType," "ConfigurationType," "FunctionalityType," "Molecule\_EntityType," "StructureType," "LocationType," "ChainType," "DomainType," and "ReceptorType."

## CONCEPTS OF IDENTIFICATION

### "MoleculeType"

"MoleculeType" is a concept of identification that allows one to identify the type of molecule, based on the type of the constitutive elements and on the concepts of obtention.

The "MoleculeType" concept comprises four major leafconcepts (keywords in the IMGT databases and tools):

1. "**gDNA**" identifies genomic DNA, a polynucleotide sequence made of a, t, c, g (representing the bases adenine [a], thymine [t], cytosine [c], and guanine [g], attached to the 2-deoxyribose/phosphate backbone), obtained from a genome, or by extension, synthetic DNA having the characteristics of genomic DNA.
2. "**mRNA**" identifies messenger RNA, a polynucleotide sequence made of a, u, c, g (representing the bases adenine [a], uracil [u], cytosine [c], and guanine [g], attached to the ribose/phosphate backbone), and obtained by transcription of gDNA.
3. "**cDNA**" identifies complementary DNA, a polynucleotide sequence made of a, t, c, g (representing the bases adenine [a], thymine [t], cytosine [c], and guanine [g], attached to the 2-deoxyribose/phosphate backbone), and obtained in vitro by reverse transcription of mRNA.
4. "**protein**" identifies a sequence made of amino acids, obtained by translation of mRNA, or by in vitro synthesis.

### "GeneType"

"GeneType" is a concept of identification that allows one to identify the type of gene.

The "GeneType" concept comprises six leafconcepts (keywords in the IMGT databases and tools):

- Two leafconcepts "**conventional-with-leader**" and "**conventional-without-leader**" identify "conventional" genes, that is any (coding or noncoding) gene other than the IG or TR genes, with or without leader L region (or signal peptide), respectively. A conventional gene has by definition the characteristics of a classical gene (initiation codon and stop codon in the same gene unit).
- Four leafconcepts identify the IG and TR genes of the vertebrate adaptive immune response and are specific to immunogenetics. Three of these leafconcepts, "**variable**" (V), "**diversity**" (D), and "**joining**" (J), identify the IG and TR genes that rearrange at the DNA level in the B and T cells and code the V, D, and J regions, respectively, of the IG and TR variable domains; the fourth leafconcept, "**constant**" (C), identifies the IG and TR genes that code the C region of the IG and TR chains (Lefranc and Lefranc 2001a,b). IG and TR genes have special characteristics compared to the conventional

genes: They have a recombination signal (RS) in 3' (for the V genes), in 5' and 3' (for the D genes), and in 5' (for the J genes) and require DNA rearrangements (V-J or V-D-J) during the biosynthesis of the IG and TR chains (Lefranc and Lefranc 2001a,b).

### “ConfigurationType”

“**ConfigurationType**” is a concept of identification that allows one to identify the type of configuration of a gene, and by extension, the type of configuration of the Molecule\_EntityType leafconcepts that contain it.

The “ConfigurationType” concept comprises four leafconcepts (keywords in the IMGT databases and tools):

1. “**undefined**” identifies, whatever the molecule type, the configuration of the conventional genes and that of the IG and TR C genes (Lefranc and Lefranc 2001a,b), and by extension, the configuration of the Molecule\_EntityType leafconcepts that only contain genes in undefined configuration.
2. “**germline**” identifies, whatever the molecule type, the configuration of the IG and TR V, D, and J genes **before** DNA rearrangements (Lefranc and Lefranc 2001a,b), and by extension, the configuration of the Molecule\_EntityType leafconcepts that contain germline genes (with or without C genes in undefined configuration).
3. “**rearranged**” identifies, whatever the molecule type, the configuration of the IG and TR V, D, and J genes **after** DNA rearrangements, and by extension, the configuration of the Molecule\_EntityType leafconcepts that contain rearranged genes with, if present, completely rearranged D genes (with or without C genes in undefined configuration).
4. “**partially-rearranged**” identifies, whatever the molecule type, the configuration of partially rearranged IG or TR D genes, and by extension, the configuration of Molecule\_EntityType leafconcepts that contain at least one partially rearranged D gene (with another D or with rearranged V or J genes [with or without C genes in undefined configuration]).

### “FunctionalityType”

“**FunctionalityType**” is a concept of identification that allows one to identify, whatever the molecule type (gDNA, cDNA, mRNA, or protein), the type of functionality of a Molecule\_EntityType leafconcept (Giudicelli and Lefranc 1999; Lefranc et al. 2004, 2005a, 2008; Duroux et al. 2008). The “FunctionalityType” concept comprises five leafconcepts (keywords in the IMGT databases and tools), divided into two categories, according to the configuration type of the Molecule\_EntityType leafconcept:

Three leafconcepts, “**functional**”, “**ORF**” (open reading frame), and “**pseudogene**,” identify the functionality of Molecule\_EntityType leafconcepts in **undefined** configuration (conventional genes and IG and TR C genes), or in **germline** configuration (IG and TR V, D, and J genes **before** DNA rearrangements).

Two leafconcepts, “**productive**” and “**unproductive**”, identify the functionality of Molecule\_EntityType leafconcepts in **rearranged** or **partially-rearranged** configuration (IG and TR entities **after** DNA rearrangements, and by extension, fusion entities resulting from translocations, and hybrid entities obtained by biotechnology molecular engineering).

Thus:

1. “**functional**” identifies, whatever the molecule type, the functionality of Molecule\_EntityType leafconcepts in **undefined** or **germline** configuration, whose coding region has an ORF without stop codon, and for which there is no described defect in the splicing sites, recombination signals, and/or regulatory elements.
2. “**ORF**” identifies, whatever the molecule type, the functionality of Molecule\_EntityType leafconcepts in **undefined** or **germline** configuration, whose coding region has an ORF:
  - but alterations have been described in the splicing sites, recombination signals, and/or regulatory elements.
  - and/or changes of conserved amino acids have been suggested by the authors to lead to incorrect folding.
  - and/or the entity is an orphan.

3. **“Pseudogene”** identifies, whatever the molecule type, the functionality of Molecule\_EntityType leaf-concepts in **undefined** or **germline** configuration, whose coding region has stop codon(s) and/or frameshift mutation(s), and/or for which a mutation affects the initiation codon (for a conventional or a V gene).
4. **“Productive”** identifies, whatever the molecule type, the functionality of Molecule\_EntityType leaf-concepts in **rearranged** or **partially-rearranged** configuration, whose coding region has an ORF without stop codon, whose junction is in-frame (for IG and TR), and for which there is no described defect in the initiation codon, splicing sites, and/or regulatory elements.
5. **“Unproductive”** identifies, whatever the molecule type, the functionality of Molecule\_EntityType leafconcepts in **rearranged** or **partially-rearranged** configuration, whose coding region has stop codon(s) and/or frameshift mutation(s), and/or whose junction is out-of-frame (for IG and TR), and/or for which a mutation affects the initiation codon, and/or for which there are defects in the splicing sites and/or in the regulatory element(s), and/or there are unusual features (e.g., translocation, gene fusion), and/or changes of conserved amino acids demonstrated as leading to incorrect folding.

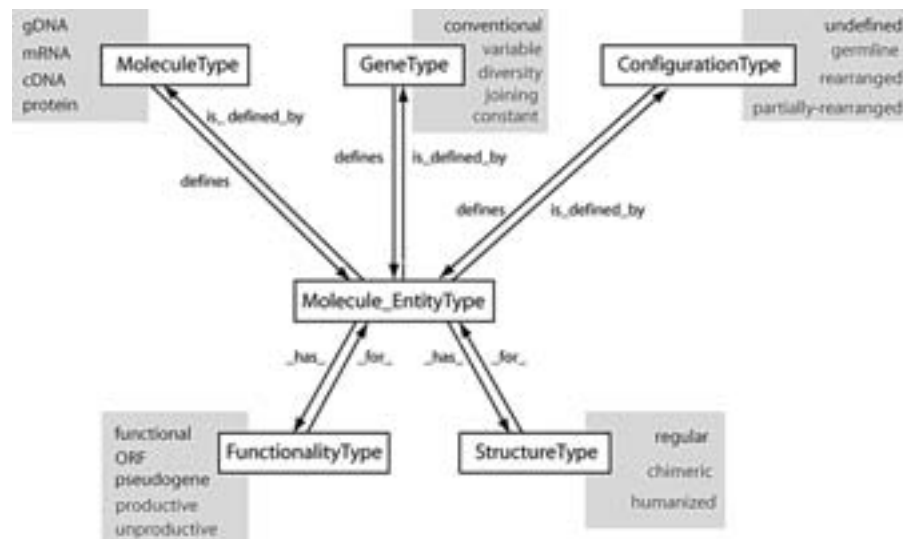
### “Molecule\_EntityType”

“Molecule\_EntityType” is a major concept of identification that allows one to identify the molecule entity type. The “Molecule\_EntityType” concept is defined by the “MoleculeType,” “GeneType,” and “ConfigurationType” concepts and has properties identified in the “FunctionalityType” and “StructureType” concepts (Fig. 1).

The “Molecule\_EntityType” concept comprises 38 leafconcepts (keywords in the IMGT databases and tools) that, based on the “MoleculeType” (“gDNA”, “mRNA”, “cDNA,” or “protein”), identify the molecule entities of the four major “MoleculeUnit” leafconcepts that are genes (10), transcripts (11), cDNA sequences (11), and chains (6) (as indicated by the suffix).

The 10 leafconcepts that are the most classical representatives of IG and TR data identification in the IMGT information system are shown as examples (Table 1). They are also illustrated in Immunoglobulin synthesis (Fig. 2).

“V-gene” identifies, for gDNA, molecule entities with a germline V gene. V-gene is in germline configuration.



**FIGURE 1.** The “Molecule\_EntityType” concept. The “Molecule\_EntityType” concept is defined by the “MoleculeType,” “GeneType,” and “ConfigurationType” concepts of identification and has properties identified in the “FunctionalityType” and “StructureType” concepts (IDENTIFICATION axiom). Arrows indicate reciprocal relations “is\_defined\_by” and “defines,” “\_has\_” and “\_for\_.” Leafconcepts are general (in blue) or specific of the IG and TR (in red) The “Molecule\_EntityType” concept has 38 leafconcepts (or keywords in the IMGT databases and tools). Only a few examples of the “StructureType” leafconcepts (or keywords in the IMGT databases and tools) are shown.

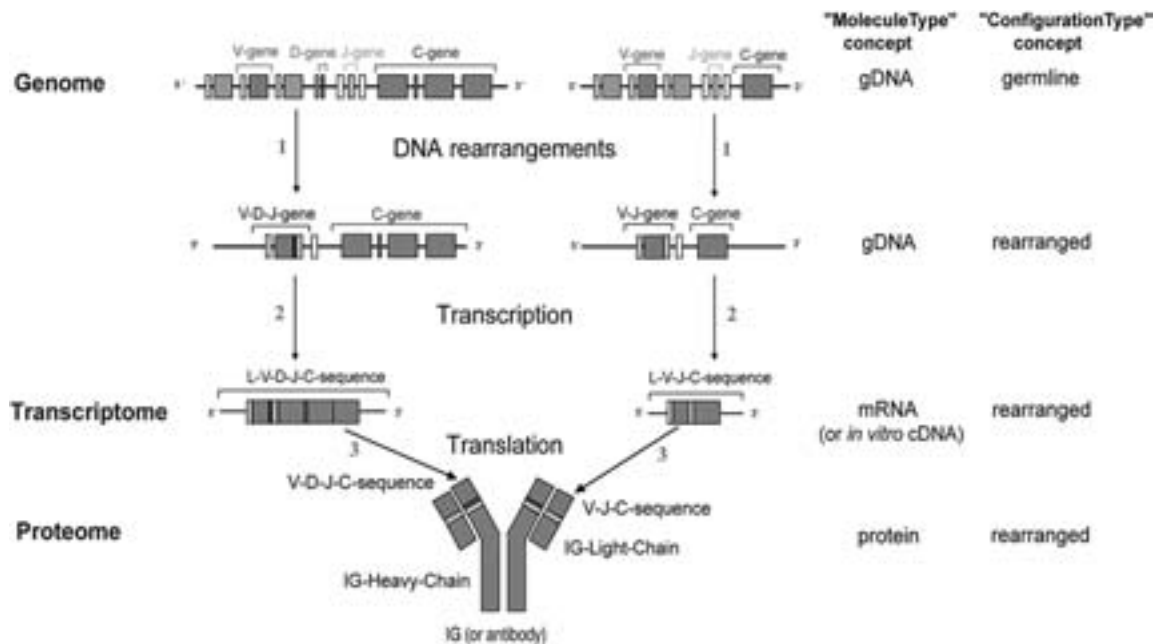


**Table 1. Molecule\_EntityType concept**

Concepts	Leafconcepts	Molecule_EntityType concept																			
		V-gene	D-gene	J-gene	C-gene	V-J-gene	V-D-J-gene	L-V-J-C-sequence	L-V-D-J-C-sequence	V-J-C-chain	V-D-J-C-chain										
MoleculeType	gDNA cDNA	x	x	x	x	x	x														
GeneType	protein variable diversity joining constant undefined	x																			
ConfigurationType	germline rearranged																				
FunctionalityType	functional ORF pseudogene productive unproductive	x	x	x	x	x	x	x	x	x	x	x	x	x	x	x	x	x	x	x	x

Ten leafconcepts (keywords in the IMGT databases and tools) are shown as examples. An “x” indicates the leafconcepts (keywords in the IMGT databases and tools) that define them (“MoleculeType,” “GeneType,” “ConfigurationType”) and their possible functionality (“FunctionalityType”).

Abbreviations: gDNA, genomic DNA; cDNA, complementary DNA; ORF, open reading frame.



**FIGURE 2.** Synthesis of an IG or antibody in humans. A human being may potentially synthesize  $10^{12}$  different antibodies (Lefranc and Lefranc 2001a). 1, DNA rearrangements (is\_rearranged\_into); 2, Transcription (is\_transcribed\_into); 3, Translation (is\_translated\_into). The configuration of C-GENE is "undefined" (ConfigurationType) (IMGT Repertoire, <http://www.imgt.org>).

**"D-gene"** identifies, for gDNA, molecule entities with a germline D gene. D-gene is in germline configuration.

**"J-gene"** identifies, for gDNA, molecule entities with a germline J gene. J-gene is in germline configuration.

**"C-gene"** identifies, for gDNA, molecule entities with a C gene in undefined configuration. C-gene is in undefined configuration.

**"V-J-gene"** identifies, for gDNA, molecule entities with a rearranged V gene and a rearranged J gene. V-J-gene is in rearranged configuration.

**"V-D-J-gene"** identifies, for gDNA, molecule entities with a rearranged V gene, at least one rearranged D gene, and a rearranged J gene. V-D-J-gene is in rearranged configuration.

**"L-V-J-C-sequence"** identifies, for cDNA, molecule entities with a leader L region, a rearranged V region, a rearranged J region, and a C region in undefined configuration. L-V-J-C-sequence is in rearranged configuration.

**"L-V-D-J-C-sequence"** identifies, for cDNA, molecule entities with a leader L region, a rearranged V region, at least one rearranged D region, a rearranged J region, and a C region in undefined configuration. L-V-D-J-C-sequence is in rearranged configuration.

**"V-J-C-chain"** identifies, for protein, molecule entities without a leader L region (mature form) and with a rearranged V region, a rearranged J region, and a C region in undefined configuration. V-J-C-chain is in rearranged configuration.

**"V-D-J-C-chain"** identifies, for protein, molecule entities without a leader L region (mature form) and with a rearranged V region, at least one rearranged D region, a rearranged J region, and a C region in undefined configuration. V-D-J-C-chain is in rearranged configuration.

The 28 other leafconcepts (keywords in the IMGT databases and tools) identify sterile transcripts, partially rearranged entities, partial sequences, or other molecule units, and are not detailed here.

### "StructureType"

**"StructureType"** is a concept of identification that allows one to identify, whatever the molecule type (gDNA, cDNA, mRNA, protein), the type of structure of Molecule\_EntityType leafconcepts.

The “StructureType” concept comprises 17 leafconcepts (keywords in the IMGT databases and tools) that identify structures with a classical organization (“regular”), from those which have been modified either naturally in vivo (“processed”, “unprocessed”, “partially-processed”, “sterile-transcript”, “unspliced”, “partially-spliced”, “spliced”, “membrane”, “secreted”, “membrane-and-secreted”, “mature-form”, “immature-form”), or artificially in vitro (“engineered”, “humanized”, “chimeric”, “fusion”) (“chimeric” and “fusion” can also be obtained in vivo) (Giudicelli and Lefranc 1999; Lefranc et al. 2004, 2005a, 2008; Duroux et al. 2008).

### “LocationType”

“LocationType” is a concept of identification that allows one to identify, whatever the molecule type (gDNA, cDNA, mRNA, protein), the Molecule\_EntityType leafconcepts, based on their location.

The “LocationType” concept comprises, for instance, the following leafconcepts (keywords in the IMGT databases and tools):

“**orphon**” identifies, whatever the molecule type, a gene that is found in vivo in a different locus from the main locus (either on the same chromosome or on another chromosome).

“**transgene**” identifies, whatever the molecule type, a gene that is artificially introduced into a multicellular organism (e.g., mouse, plant).

“**translocated**” identifies, whatever the molecule type, a gene that results from a translocation (in vivo).

“**transposed**” identifies, whatever the molecule type, a transgene or a retrotransposon that is permanently inserted in a chromosome.

### “ChainType”

“ChainType” is a concept of identification that allows one to identify the type of chain. It is one of the most important concepts of identification for the standardization of genome, transcriptome, and proteome data in systems biology. Indeed, being able to identify a type of chain means that it is possible to identify the transcript and the encoding gene(s).

The “ChainType” concept is defined by the “Molecule\_EntityType” and the “DomainType” concepts of identification and by concepts of classification (**From IMGT-ONTOLOGY CLASSIFICATION Axiom to IMGT Standardized Gene and Allele Nomenclature: For Immunoglobulins (IG) and T Cell Receptors (TR)** [Lefranc 2011c]). The “ChainType” concept is a “highconcept” as it corresponds to a hierarchy of concepts which allow one to identify the chain type at different levels of granularity (up to five for “ChainType”).

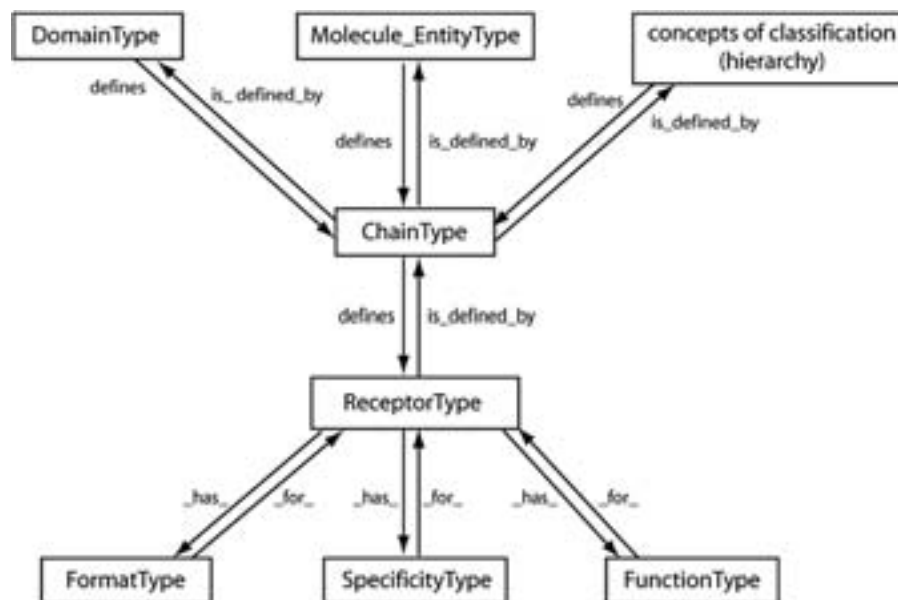
The finest level of granularity, the “GeneLevelChainType” concept, identifies the type of chain by reference to the gene(s) which code(s) the chain. It represents the main concept for a very precise identification because it establishes a relationship with “Gene” (concept of classification generated from the CLASSIFICATION axiom) (the reciprocal relations are: “is\_coded\_by” and “codes”).

Thus:

- A *Homo sapiens* IG-Heavy-Gamma-1-Chain is defined by a leafconcept of the “Molecule\_EntityType” (V-D-J-C-chain), “DomainType” and numbers (1 V domain, 3 C domains), and the gene encoding the constant region (*Homo sapiens*IGHG1) (Fig. 3).
- Similarly, a *Homo sapiens* IG-Kappa chain is defined by a leafconcept of the “Molecule\_EntityType” (V-J-C-chain), “DomainType” and numbers (1 V domain, 1 C domain), and the gene encoding the constant region (*Homo sapiens*IGKC).

The number of instances of the leafconcepts of the “GeneLevelChainType” concept (and therefore of the keywords in the IMGT databases and tools) depends on the number of functional genes and ORF (“FunctionalityType”), per haploid genome, in a given species (in the case of the IG and TR genes, it is the number of functional and ORF C genes which is taken into account). If only the functional genes are considered, the instances of this leafconcept correspond to the isotypes.

As the “ChainType” concept can be used by extrapolation for any of the four “MoleculeUnit” (gene, transcript, cDNA sequence, chain), in order to facilitate query and/or data extraction from the IMGT/LIGM-DB database (Giudicelli et al. 2006), the suffix -Chain (used in “ChainType” leafconcepts) is not specified in the “ChainType” keywords.



**FIGURE 3.** The “ReceptorType” concept. The “ReceptorType” concept is defined by the “ChainType” concept of identification and has properties identified in the “FormatType,” “SpecificityType,” and “FunctionType” concepts (IDENTIFICATION axiom). The “ChainType” concept is itself defined by the “Molecule\_EntityType” and “DomainType” concepts and by concepts of classification organized in a hierarchy (see CLASSIFICATION axiom). Arrows indicate reciprocal relations “is\_defined\_by” and “defines,” “\_has\_” and “\_for\_.” The “ChainType” and “ReceptorType” concepts have different levels of granularity (up to five) and are highconcepts.

### “DomainType”

“**DomainType**” is a concept of identification that allows one to identify the type of domain.

A domain is a chain subunit characterized by its three-dimensional (3D) structure, and by extension its amino acid sequence and the nucleotide sequence which encodes it.

In IMGT-ONTOLOGY, the “DomainType” concept has three major leafconcepts that were first described in the proteins of the adaptive immune response, IG or antibodies, TR and MH, and then extended to the proteins of the IgSF and MhSF (Lefranc 1997, 1999; Lefranc et al. 2003, 2005b,c). These leafconcepts (keywords in the IMGT databases and tools) are:

the “**V domain**” (Variable [V] domain) that includes the variable domains of the IG and TR and the V-like domains of other IgSF proteins; (Lefranc 1999; Lefranc et al. 2003)

the “**C domain**” (Constant [C] domain) that includes the constant domains of the IG and TR and the C-like domains of other IgSF proteins; (Lefranc et al. 2005c)

the “**G domain**” (Groove [G] domain) that includes the groove domains of the MH and the G-like domains of other MhSF proteins (Lefranc et al. 2005b).

Standardized description of the V, C, and G domains is based on the IMGT unique numbering (Lefranc 1997, 1999; Lefranc et al. 2003, 2005b,c). Graphical two-dimensional (2D) representations or IMGT Colliers de Perles (Ruiz and Lefranc 2002; Kaas and Lefranc 2007; Kaas et al. 2007) can be obtained using the IMGT/DomainGapAlign and IMGT/Collier de Perles tools (Ehrenmann et al. 2010a,b; <http://www.imgt.org>).

### “ReceptorType”

“**ReceptorType**” is a concept that allows one to identify the type of receptor, based on the ChainType leafconcept(s) that identify the associated chains. “ReceptorType” is a “highconcept” as it corresponds to a hierarchy of concepts (depending itself on the “ChainType” hierarchy) which allows one to identify the receptor type at different levels of granularity (up to five for “ReceptorType”). The

“ReceptorType” concept is defined by the “ChainType” concept of identification and has properties identified in the “FormatType,” “SpecificityType,” and “FunctionType” concepts (Fig. 3; Duroux et al. 2008; Lefranc et al. 2008).

Thus, “IG” identifies an IG-Heavy-Chain\_IG-Light-Chain dimer, whereas at a different granularity (“GeneLevelChainType”), the same receptor will be identified as an IG-Heavy-Gamma-1-Chain\_Light-Kappa-Chain dimer.

IG and TR are the specific antigen receptors of the adaptive immune response (and therefore major leafconcepts of the “ReceptorType” concept) characterized by their function of binding and recognition of the antigen (“FunctionType”). The “SpecificityType” concept allows an amino acid characterization of this binding.

The “SpecificityType” concept has two major leafconcepts (keywords in the IMGT databases and tools):

“paratope” identifies the amino acids of the antigen receptor V domains that recognize and bind the antigen (Ag).

“epitope” identifies the antigen part recognized and bound by the antigen receptor (IG or TR).

The “FormatType” concept has many leafconcepts (keywords in the IMGT databases and tools) that identify the different types of format of the receptors, and particularly those of the antibodies (Fab, F(ab')<sub>2</sub>, Fc, scFv, Fv...) and of the fusion proteins for immune applications (FPIA) (Lefranc 2011f).

The corresponding keywords are extensively used in the identification of the IMGT/2Dstructure-DB and IMGT/3Dstructure-DB entries (Lefranc 2008, 2009; Lefranc et al. 2008; Ehrenmann et al. 2010a,b) and of the IMGT/mAb-DB therapeutic mAbs and FPIA (Poiron et al. 2010). Paratope/epitope are used in IMGT/3Dstructure-DB (<http://www.imgt.org>) for analysis of interactions in IG/Ag and TR/peptide/major histocompatibility (TR/pMH) complexes (Kaas and Lefranc 2005; Kaas et al. 2008).

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# From IMGT-ONTOLOGY DESCRIPTION Axiom to IMGT Standardized Labels: For Immunoglobulin (IG) and T Cell Receptor (TR) Sequences and Structures

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## INTRODUCTION

More than 560 IMGT standardized labels have been defined for the description of immunoglobulin (IG) and T cell receptor (TR) sequences and structures. These labels, as detailed here, are based on the concepts of description (generated from the DESCRIPTION axiom) of IMGT-ONTOLOGY, the global reference in immunogenetics and immunoinformatics, built by IMGT, the international ImMunoGeneTics information system. IMGT labels for nucleotide sequences correspond to the "Molecule\_Entity-Prototype" concept with its leafconcepts (a leafconcept is the finest level of granularity), its associated concepts of description, and its prototypes or graphical representations. These labels have been essential for the IG and TR description in the IMGT sequence and gene databases (IMGT/LIGM-DB, IMGT/GENE-DB) and tools (IMGT/V-QUEST, IMGT/JunctionAnalysis, IMGT/HighV-QUEST, IMGT/LIGMotif). IMGT labels for amino acid sequences and structures correspond to the "RECEPTOR," "CHAIN," and "DOMAIN" concepts. These labels have been crucial for the IG and TR description in the IMGT two-dimensional (2D) and three-dimensional (3D) databases (IMGT/2Dstructure-DB, IMGT/3Dstructure-DB) and tools (IMGT/DomainDisplay, IMGT/DomainGapAlign). As all leafconcepts of description are directly translated into IMGT standardized labels for use in the IMGT databases and tools, IMGT-ONTOLOGY concepts of description contribute to data coherence and consistency and facilitate the interoperability between the different components of IMGT, bridging the description of sequences and structures.

## RELATED INFORMATION

IMGT standardized labels for nucleotide sequences of IG and TR are available at the IMGT/LIGM-DB (Giudicelli et al. 2006) database query page (IMGT Home page <http://www.imgt.org>). IMGT standardized labels for amino acid sequences and structures (IMGT/2Dstructure-DB, IMGT/3Dstructure-DB [Kaas et al. 2004; Ehrenmann et al. 2010b]) are available in the IMGT Scientific chart at: <http://www.imgt.org/textes/IMGTScientificChart/SequenceDescription/IMGT3Dkeywords.html>. These labels describe the amino acid sequences and the 2D and 3D structure organization (in receptors, chains, and domains) of IG, TR, major histocompatibility (MH), related proteins of the immune system (RPI), and fusion proteins for immune applications (FPIA) (Lefranc 2011f). The definition of these labels is available in the IMGT Scientific chart at: <http://www.imgt.org/textes/IMGTScientificChart/SequenceDescription/IMGT3Dlabeldef.html>.

A detailed description of IMGT is provided in **IMGT, The International ImMunoGeneTics Information System** (Lefranc 2011a). Information is also available on **From IMGT-ONTOLOGY IDENTIFICATION Axiom to IMGT Standardized Keywords: For Immunoglobulins (IG), T Cell Receptors (TR), and Conventional Genes** (Lefranc 2011b), **From IMGT-ONTOLOGY CLASSIFICATION Axiom to IMGT Standardized Gene and Allele Nomenclature: For Immunoglobulins (IG) and T Cell Receptors (TR)** (Lefranc 2011c), **IMGT Unique Numbering for the Variable (V), Constant (C), and Groove**

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(G) Domains of IG, TR, MH, IgSF, and MhSF (Lefranc 2011d), and **IMGT Collier de Perles for the Variable (V), Constant (C), and Groove (G) Domains of IG, TR, MH, IgSF, and MhSF** (Lefranc 2011e).

In addition, protocols are available for **IMGT/V-QUEST: IMGT Standardized Analysis of the Immunoglobulin (IG) and T Cell Receptor (TR) Nucleotide Sequences** (Giudicelli et al. 2011), **IMGT/JunctionAnalysis: IMGT Standardized Analysis of the V-J and V-D-J Junctions of the Rearranged Immunoglobulins (IG) and T Cell Receptors (TR)** (Giudicelli and Lefranc 2011), **IMGT/Collier de Perles: IMGT Standardized Representation of Domains (IG, TR, and IgSF Variable and Constant Domains, MH and MhSF Groove Domains)** (Ehrenmann et al. 2011), **IMGT/DomainGapAlign: IMGT Standardized Analysis of Amino Acid Sequences of Variable, Constant, and Groove Domains (IG, TR, MH, IgSF, MhSF)** (Ehrenmann and Lefranc 2011a), and **IMGT/3Dstructure-DB: Querying the IMGT Database for 3D Structures in Immunology and Immunoinformatics (IG or Antibodies, TR, MH, RPI, and FPIA)** (Ehrenmann and Lefranc 2011b).

## BACKGROUND INFORMATION

The DESCRIPTION axiom is an axiom of the Formal IMGT-ONTOLOGY or IMGT-Kaleidoscope (Giudicelli and Lefranc 1999; Lefranc et al. 2004, 2005a, 2008; Duroux et al. 2008). The DESCRIPTION axiom postulates that any molecule, cell, tissue, organ, organism, or population, any process and any relation, has to be described. The DESCRIPTION axiom has generated the concepts of description of IMGT-ONTOLOGY, the global reference in immunogenetics and immunoinformatics, built by IMGT (<http://www.imgt.org>; Lefranc et al. 2009). The DESCRIPTION axiom is one of seven axioms of the Formal IMGT-ONTOLOGY, the others being "IDENTIFICATION axiom," "CLASSIFICATION axiom," "NUMEROTATION axiom," "LOCALIZATION axiom," "ORIENTATION axiom," and "OBTENTION axiom." The leafconcepts (finest level of granularity of the concepts) of description are directly translated into IMGT standardized labels and used in the IMGT databases and tools. IMGT standardized labels have been essential for the sequence description in IMGT/LIGM-DB (Giudicelli et al. 2006), the first and the largest IMGT database that contains IG and TR nucleotide sequences and the translation of annotated sequences, IMGT/GENE-DB (Giudicelli et al. 2005), the IMGT gene database, IMGT/2Dstructure-DB, the IMGT amino acid sequence and 2D structure database (Ehrenmann et al. 2010b), and IMGT/3Dstructure-DB, the IMGT 3D structure database (Kaas et al. 2004; Ehrenmann et al. 2010b). IMGT standardized labels have been crucial in the IMGT tools for the analysis of nucleotide sequences (IMGT/V-QUEST [Giudicelli et al. 2004; Brochet et al. 2008; Giudicelli and Lefranc 2009], IMGT/JunctionAnalysis [Yousfi Monod et al. 2004; Bleakley et al. 2006], IMGT/HighV-QUEST [Alamyar et al. 2010]), of amino acid sequences (IMGT/DomainGapAlign [Ehrenmann et al. 2010b]), or of large genomic sequences (IMGT/LIGMotif [Lane et al. 2010]).

## MOLECULE ENTITY PROTOTYPE

### "Molecule\_EntityPrototype" Concept

The "Molecule\_EntityPrototype" is a concept, generated from the DESCRIPTION axiom, that provides the description for the "Molecule\_EntityType" concept (IDENTIFICATION axiom). There are as many leafconcepts in the "Molecule\_EntityPrototype" as there are leafconcepts in the "Molecule\_EntityType" (**From IMGT-ONTOLOGY IDENTIFICATION Axiom to IMGT Standardized Keywords: For Immunoglobulins (IG), T Cell Receptors (TR), and Conventional Genes** [Lefranc 2011b]). Thus the "Molecule\_EntityPrototype" comprises 38 leafconcepts that describe the organization of each entity with its constitutive motifs and relations. Each "Molecule\_EntityPrototype" leafconcept is linked to a "Molecule\_EntityType" leafconcept by the reciprocal relations "describes" and "is\_described\_by." For example, a "V-gene" is described by "V-GENE," a "V-D-J-gene" by "V-D-J-GENE," and a "L-V-D-J-sequence" by "L-V-D-J-SEQUENCE." Leafconcepts of description (labels in the IMGT databases and tools) are written in capital letters.

The 38 "Molecule\_EntityPrototype" leafconcepts, as indicated by the suffix, describe the four major "MoleculeUnit" leafconcepts that are genes (10), transcripts (11), cDNA sequences (11), and chains (6). The 10 leafconcepts that are the most classical representatives of IG and TR data description in the IMGT information system are shown as examples (Table 1).

**Table 1. IMGT-ONTOLOGY “Molecule\_EntityProtoType” concept**

Labels	Molecule_EntityProtoType leafconcepts (labels in the IMGT databases and tools)										
	V-gene	D-gene	J-gene	C-gene	V-J-gene	V-D-J-gene	L-V-J-C-sequence	L-V-D-J-C-sequence	V-J-C-chain	V-D-J-C-chain	V-D-J-C-chain
1st-CYS	x			x	x	x	x	x	x	x	x
2nd-CYS	x			x	x						
3'D-HEPTAMER		x									
3'D-NONAMER		x									
3'D-RS		x									
3'D-SPACER		x									
3'UTR	x		x	x	x						
5'D-HEPTAMER		x									
5'D-NONAMER		x									
5'D-RS		x									
5'D-SPACER		x									
5'UTR	x		x	x	x						
ACCEPTOR-SPLICE	x			x	x						
C-GENE-UNIT				x							
C-REGION				x							
CDR1-IMGT	x				x	x	x	x	x	x	x
CDR2-IMGT	x				x	x	x	x	x	x	x
CDR3-IMGT	x				x	x	x	x	x	x	x
CONSERVED-TRP	x			x	x						
D-GENE-UNIT		x									
D-REGION		x			x						
DONOR-SPLICE	x		x	x	x						
FR1-IMGT	x				x	x	x	x	x	x	x
FR2-IMGT	x				x	x	x	x	x	x	x
FR3-IMGT	x				x	x	x	x	x	x	x
FR4-IMGT	x				x	x	x	x	x	x	x
INIT-CODON	x				x						
J-GENE-UNIT			x								
J-HEPTAMER			x								
J-NONAMER			x								
J-PHE			x		x	x	x	x	x	x	x
J-REGION			x		x						
J-RS			x								
J-SPACER			x								
J-TRP			x		x	x	x	x	x	x	x
JUNCTION					x	x	x	x	x	x	x
L-INTRON-L	x				x						

(continued)

**Table 1. Continued**

Labels	Molecule_EntityProtoType leafconcepts (labels in the IMGT databases and tools)										
	V-gene	D-gene	J-gene	C-gene	V-J-gene	V-D-J-gene	L-V-J-C-sequence	L-V-D-J-C-sequence	V-J-C-chain	V-D-J-C-chain	
L-PART1	x				x	x					
L-PART2	x				x	x					
L-REGION							x				
L-V-D-J-C-REGION								x			
L-V-GENE-UNIT	x										
L-V-J-C-REGION							x				
(N-D)-REGION						x				x	
N-REGION					x	x			x		
V-D-J-C-REGION								x		x	
V-D-J-EXON						x					
V-D-J-REGION						x				x	
V-EXON	x										
V-HEPTAMER	x										
V-INTRON	x					x					
V-J-C-REGION					x			x			
V-J-EXON					x				x		
V-J-REGION					x						
V-NONAMER	x										
V-REGION	x				x				x		
V-RS	x										
V-SPACER	x										

The 10 most representative leafconcepts for IG and TR are shown with the most important labels (indicated by an “x”) that describe them in the IMGT databases and tools. The labels for a detailed description of the C-GENE exons or of the C-REGION in these leafconcepts are not shown.



## Associated Concepts of Description

The labels used for the description of the "Molecule\_EntityPrototype" (Table 1) correspond to the leafconcepts of the following concepts:

### "GeneUnit"

The "GeneUnit" concept has four leafconcepts (labels in the IMGT databases and tools): "L-V-GENE-UNIT," "D-GENE-UNIT," "J-GENE-UNIT," and "C-GENE-UNIT" (Lane et al. 2010). In contrast to "V-GENE," "D-GENE," "J-GENE," and "C-GENE," the "GeneUnit" labels are precisely delimited in 5' and 3', respectively, by the 5' end and 3' end of their constitutive labels: "L-PART1" and "V-RS" for V, "5'D-RS" and "3'D-RS" for D, "J-RS" and "J-REGION" for J, "ACCEPTOR-SPLICE" and "STOP-CODON" for C.

### "RecombinationSignal"

The "RecombinationSignal" (RS) concept is specific to the IG and TR V, D, and J genes which rearrange at the DNA level in B cells (for the IG) and T cells (for the TR) (Lefranc and Lefranc 2001a,b) and represents the major difference from "conventional" genes. The "RecombinationSignal" labels describe the RS that are localized in 3' of the V genes ("V-RS"), in 5' of the J genes ("J-RS"), and on both sides of the D genes ("5'D-RS" and "3'D-RS") (Lefranc and Lefranc 2001a,b). They consist of conserved heptamers (-HEPTAMER) and nonamers (-NONAMER) separated by less conserved spacers (-SPACER) of  $12 \pm 1$  or  $23 \pm 1$  base pairs (bp) which vary between loci and species (IMGT Repertoire, <http://www.imgt.org/>).

### "Core"

The "Core" concept allows one to describe the coding region of genes and contains five leafconcepts which are "REGION" (for "conventional" gene), "V-REGION," "D-REGION," "J-REGION," and "C-REGION" (for "variable" [V], "diversity" [D], "joining" [J], "constant" [C] genes). These important labels have permitted the definition and standardized description of the IG and TR alleles (concepts of classification), now approved at the international level (Lefranc and Lefranc 2001a,b) (**From IMGT-ONTOLOGY CLASSIFICATION Axiom to IMGT Standardized Gene and Allele Nomenclature: For Immunoglobulins (IG) and T Cell Receptors (TR)** [Lefranc 2011c]). Moreover, they have allowed one to link sequences and structures of the IG and TR by their composite labels that describe rearranged sequences. Thus, a V domain is encoded by a "V-J-REGION" or a "V-D-J-REGION," and IG and TR chains are encoded by a "V-J-C-REGION" or a "V-D-J-C-REGION."

### "FR-IMGT and CDR-IMGT"

The "FR-IMGT" and "CDR-IMGT" concepts allow one to describe the framework regions and complementarity determining regions, respectively, based on the IMGT unique numbering (Lefranc 1997, 1999; Lefranc et al. 2003) (**IMGT Unique Numbering for the Variable (V), Constant (C), and Groove (G) Domains of IG, TR, MH, IgSF, and MhSF** [Lefranc 2011d]). "FR-IMGT" has four leafconcepts (labels in the IMGT databases and tools): "FR1-IMGT," "FR2-IMGT," "FR3-IMGT," and, for rearranged sequences, "FR4-IMGT." "CDR-IMGT" has three leafconcepts (labels in the IMGT databases and tools): "CDR1-IMGT," "CDR2-IMGT," and "CDR3-IMGT" (the composition of "CDR3-IMGT" is different depending on whether the sequence is germline or rearranged) (Lefranc and Lefranc 2001a,b).

### "Junction"

The "JUNCTION" concept allows one to describe the junction resulting from the V-J or V-D-J rearrangement. "JUNCTION" leafconcepts (labels in the IMGT databases and tools) comprise: "3'V-REGION," "5'J-REGION," "N-REGION," "(N-D)-REGION," and "P-REGION" (Yousfi Monod et al. 2004; Bleakley et al. 2006).

### "Leader"

The "leader" concept allows one to describe the leader (L) or signal peptide. "Leader" has three leafconcepts (labels in the IMGT databases and tools): "L-PART1," "L-PART2," and "L-REGION."

In the IG and TR V genes, “L-PART1” (first exon of “L-V-GENE-UNIT”) and L-PART2 (first codons of V-EXON, the second exon of “L-V-GENE-UNIT”) are separated by the “V-INTRON” and their splicing frame is of type 1 (sf1) (IMGT Aide-mémoire, <http://www.imgt.org>). In cDNA and chains (spliced L-PART1 and L-PART2), the leader is described as “L-REGION.” The L-REGION is absent from mature chains as a result of proteolytic cleavage in the endoplasmic reticulum (ER). The “L-PART1,” “L-PART2,” and “L-REGION” labels are also used for the description of “conventional-with-leader” genes (**From IMGT-ONTOLOGY IDENTIFICATION Axiom to IMGT Standardized Keywords: For Immunoglobulins (IG), T Cell Receptors (TR), and Conventional Genes** [Lefranc 2011b]).

### “Conserved\_AA\_codons”

The “Conserved\_AA\_codon” concept allows one to describe conserved amino acids (AA) and/or codons. It includes for example, for the V-DOMAIN, five leafconcepts (labels in the IMGT databases and tools): “1st-CYS” (position 23), “CONSERVED-TRP” (position 41), “2nd-CYS” (position 104), “J-TRP,” and “J-PHE” (position 118) (positions in parentheses are according to the IMGT unique numbering [Lefranc 1997, 1999; Lefranc et al. 2003]) (**IMGT Unique Numbering for the Variable (V), Constant (C), and Groove (G) Domains of IG, TR, MH, IgSF, and MhSF** [Lefranc 2011d]). The “2nd-CYS” 104 and “J-TRP” or “J-PHE” 118 (that belong to the FR3-IMGT and FR4-IMGT, respectively) are the anchors of the rearranged CDR3-IMGT. They delimit the “JUNCTION” that spans 104 to 118 and includes them at its ends (Yousfi Monod et al. 2004; Bleakley et al. 2006).

Conserved amino acids “J-TRP” and “J-PHE” are part of a conserved motif “[W,F]-G-X-G” in the “FR4-IMGT” of the “V-DOMAIN” of IG and TR (where W, tryptophan “J-TRP”; F, phenylalanine “J-PHE”; G, glycine; and X, any amino acid). Codons of conserved amino acids are “tgg” for “CONSERVED-TRP” and “J-TRP,” “tgc,” and “tgt” for “1st-CYS” and “2nd-CYS,” and “ttt” and “ttc” for “J-PHE.” These conserved codons and amino acids and their mutations are identified and described by IMGT/V-QUEST (Giudicelli et al. 2004; Brochet et al. 2008; Giudicelli and Lefranc 2009) and IMGT/HighV-QUEST (Alamyar et al. 2010) for nucleotide sequences.

The “1st-CYS” (position 23), “CONSERVED-TRP” (position 41), and “2nd-CYS” (position 104) are also conserved in the “C-DOMAIN” of IG and TR (Lefranc et al. 2005c). Conserved amino acids for V, C, or G domains are highlighted in IMGT/DomainDisplay (Ehrenmann et al. 2010b; <http://www.imgt.org>).

### Prototype or Graphical Representation

A prototype is a graphical representation of a “Molecule\_EntityPrototype” leafconcept. Two prototypes, those of “V-GENE” and “V-D-J-GENE,” are shown in Figure 1 as examples of a germline entity and of a rearranged entity, respectively. Twenty-six labels for “V-GENE” and 29 labels for “V-D-J-GENE” (19 of them being shared by the two prototypes), on a total of 277 different labels for sequences in IMGT/LIGM-DB, are necessary and sufficient for a complete description of these prototypes. Definitions of the IMGT/LIGM-DB labels used for the two “V-GENE” and “V-D-J-GENE” prototypes are provided, as examples, in Table 2.

The organization of a prototype is based on the relations that order two labels. A set of 12 relations is necessary and sufficient to describe the relations between labels in a prototype (Table 3; Giudicelli and Lefranc 1999; Lefranc et al; 2004; Duroux et al. 2008; Lane et al. 2010). The reciprocal relations “is\_in\_5\_prime\_of” and “is\_in\_3\_prime\_of” describe the relative position of labels on a 5’–3’ DNA strand when there is no intersection between labels.

## GENE CLUSTER

### “GeneCluster” Concept

The “GeneCluster” concept allows one to describe gene clusters, which are genomic sequences containing several genes. The genes in a cluster can be described either by the same prototype (e.g., a “V-CLUSTER” only contains genes described by the “V-GENE” prototype) or by different prototypes (e.g., a “V-D-J-CLUSTER” contains at least one “V-GENE,” one “D-GENE,” and one “J-GENE”). Examples of IMGT-ONTOLOGY “GeneCluster” leafconcepts (labels in the IMGT databases and tools) are shown in Table 4. They are particularly useful for the annotation of large scale genomic IG and TR loci (Lane et al. 2010) and several of them are also used by Sequence Ontology (SO) (Table 4; Eilbeck et al. 2005).



**Table 2. List of the 39 labels used for the description of the “Molecule\_EntityPrototype” “V-GENE” and “V-D-J-GENE” and their definitions (from IMGT/LIGM-DB)**

Label name	Definition
1st-CYS	codon (3 nt) for conserved cysteine at position 23 in B-STRAND (FR1-IMGT)
2nd-CYS	codon (3 nt) for conserved cysteine at position 104 in F-STRAND (FR3-IMGT)
3'UTR	3' untranslated sequence, EMBL feature Key signification
3'V-REGION	region from 2nd-CYS to the 3' end of the V-REGION (for germline and rearranged)
5'UTR	5' untranslated sequence, EMBL feature Key signification
5'J-REGION	region from the 5' end of the J-REGION to the J-PHE or J-TRP (for germline and rearranged)
ACCEPTOR-SPLICE	splicing site in 5' of coding region (nagnn), with splicing occurring after g
CDR1-IMGT	first complementarity determining region according to the IMGT unique numbering
CDR2-IMGT	second complementarity determining region according to the IMGT unique numbering
CDR3-IMGT	third complementarity determining region according to the IMGT unique numbering
CONSERVED-TRP	codon (3 nt) for conserved tryptophan at position 41 in C-STRAND (FR2-IMGT)
D-REGION	coding region of D-GENE (plus 1 or 2 nt after the 5'D-HEPTAMER and/or before the 3'D-HEPTAMER, if present), or corresponding region in cDNA
DONOR-SPLICE	splicing site in 3' of coding region (ngt), with splicing occurring before g
FR1-IMGT	first framework according to the IMGT unique numbering
FR2-IMGT	second framework according to the IMGT unique numbering
FR3-IMGT	third framework according to the IMGT unique numbering
FR4-IMGT	fourth framework according to the IMGT unique numbering
INIT-CODON	initiation codon ATG
J-REGION	coding region of J-GENE (plus 1 or 2 nt after J-HEPTAMER, if present) or corresponding region in cDNA
J-TRP	codon (3 nt) for conserved tryptophan at position 118 in G-STRAND (FR4-IMGT) (in J-REGION of IG heavy chain)
JUNCTION	coding region encompassing the V-J or V-D-J junction from 2nd CYS to the J-PHE or J-TRP of the J-REGION
L-INTRON-L	sequence including L-PART1, V-INTRON, and L-PART2, in genomic DNA, or corresponding sequence in unspliced cDNA
L-PART1	exon encoding the first part of the leader peptide of a V-, V-D-, V-D-J-, or V-J-GENE or corresponding region in unspliced cDNA
L-PART2	5' region of V-EXON encoding the second part of leader peptide of a V-, V-D-, V-D-J-, or V-J-GENE or corresponding region in unspliced cDNA
L-V-GENE-UNIT	germline genomic DNA including L-PART1, V-INTRON, V-EXON, and V-RS
N-REGION	coding region encompassing an N diversity sequence, in a JUNCTION
(N-D)-REGION	coding region encompassing N-REGION and D-REGION, and if present P-REGION, in a JUNCTION
P-REGION	region encompassing a palindromic (P) sequence at the untrimmed end of a V-REGION, D-REGION, or J-REGION, in a JUNCTION
V-D-J-EXON	rearranged genomic DNA including L-PART2, V-, any D- and N-REGION, and J-REGION
V-D-J-GENE	rearranged genomic DNA including L-PART1, V-INTRON, and V-D-J-EXON, with the 5'UTR and 3'UTR
V-D-J-REGION	coding region including V-, any D- and N-REGION, and J-REGION, in rearranged genomic DNA, or corresponding region in cDNA
V-EXON	germline genomic DNA including L-PART2 and V-REGION
V-GENE	germline genomic DNA including L-PART1, V-INTRON, and V-EXON, with the 5'UTR and 3'UTR
V-HEPTAMER	7 nt recombination site, like CACAGTC, part of V-RS
V-INTRON	noncoding sequence between L-PART1 and V-EXON, in genomic DNA, or corresponding sequence in unspliced cDNA
V-NONAMER	9 nt recombination site, like ACAAAAACC, part of V-RS
V-REGION	coding region of V-GENE without the leader peptide (plus 1 or 2 nt before the V-HEPTAMER, if present), or corresponding region in cDNA
V-RS	recombination signal including V-HEPTAMER, V-SPACER, and V-NONAMER in 3' of V-REGION of a V-GENE or V-SEQUENCE
V-SPACER	12 or 23 nt spacer between the V-HEPTAMER and the V-NONAMER of a V-RS

A description of IMGT/LIGM-DB can be found in Giudicelli et al. (2006). In the list, 20 labels are shared by the two prototypes, seven are specific for “V-J-GENE,” and 12 are specific for “V-D-J-GENE.”

**Table 3. IMGT-ONTOLOGY relations between labels used for the description of prototypes**

Relation	Reciprocal relation
"adjacent_at_its_5_prime_to"	"adjacent_at_its_3_prime_to"
"included_with_same_5_prime_in"	"includes_with_same_5_prime"
"included_with_same_3_prime_in"	"includes_with_same_3_prime"
"overlaps_at_its_5_prime_with"	"overlaps_at_its_3_prime_with"
"included_in"	"includes"
"is_in_5_prime_of"	"is_in_3_prime_of"

Twelve relations are necessary and sufficient for the description of prototypes (Giudicelli and Lefranc 1999; Lefranc et al. 2004; Duroux et al. 2008; Lane et al. 2010).

leafconcepts (labels in the IMGT databases and tools) have allowed us to bridge the gap between sequence and structure description.

### "DOMAIN" Concept

The "DOMAIN" concept allows one to describe a subunit of a chain characterized by its 3D structure (Kaas et al. 2007). The IMGT-ONTOLOGY "DOMAIN" concept includes six main leafconcepts (labels in IMGT/3Dstructure-DB and in IMGT/DomainGapAlign [Ehrenmann et al. 2010b]):

- the "V-DOMAIN" (V for variable)
- the "C-DOMAIN" (C for constant)
- the "G-DOMAIN" (G for groove)
- the "V-LIKE-DOMAIN"
- the "C-LIKE-DOMAIN"
- the "G-LIKE-DOMAIN"

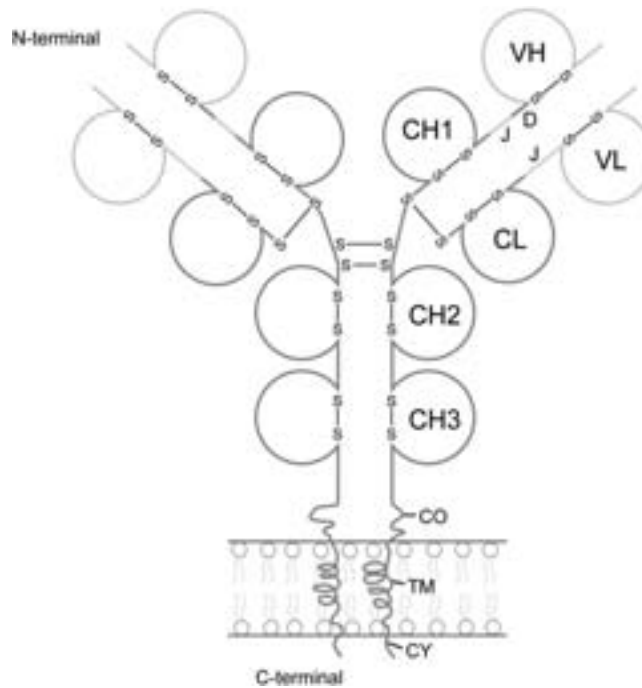
"V-DOMAIN" and "C-DOMAIN" are found in IG and TR chains and "V-LIKE-DOMAIN" and "C-LIKE-DOMAIN" in chains other than IG or TR (Lefranc 1997, 1999; Lefranc et al. 2003, 2005c). "G-DOMAIN" is found in MH chains and "G-LIKE-DOMAIN" in chains other than MH (Lefranc et al.

**Table 4. Examples of IMGT-ONTOLOGY "GeneCluster" leafconcepts**

IMGT-ONTOLOGY "GeneCluster" leafconcepts (labels in the IMGT databases and tools)	Sequence ontology	"Molecule_EntityPrototype" leafconcepts (labels and prototypes in the IMGT databases and tools)	
		Minimum number of different prototypes	Name of the different prototypes
V-CLUSTER	SO:0000526	1	V-GENE
J-CLUSTER	SO:0000513	1	J-GENE
D-CLUSTER	SO:0000559	1	D-GENE
D-J-CLUSTER	SO:0000560	2	D-GENE J-GENE
V-D-CLUSTER		2	V-GENE D-GENE
V-J-CLUSTER	SO:0000534	2	V-GENE J-GENE
V-D-J-CLUSTER	SO:0000532	3	V-GENE D-GENE J-GENE

These examples are those used in IMGT/LIGMotif (Lane et al. 2010). Six of them are also used by SO (Eilbeck et al. 2005). Relations with the "Molecule\_EntityPrototype" leafconcepts comprise the minimum number of different prototypes and the name of these different prototypes.

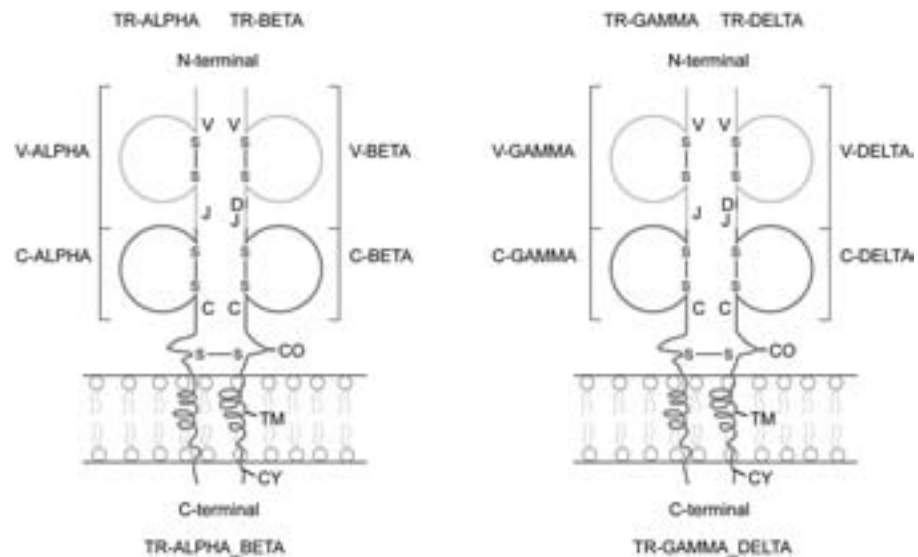




**FIGURE 2.** Schematic representation of an Ig. The Ig chain and domain labels (IMGT/3Dstructure-DB) and the corresponding sequence labels (IMGT/LIGM-DB) are shown in Tables 5 and 6. A membrane Ig (mIg) is shown. CO, CONNECTING-REGION; TM, TRANSMEMBRANE-REGION; CY, CYTOPLASMIC-REGION. The example is *Homo sapiens* IG-GAMMA1\_KAPPA (IgG1 $\kappa$ ) (Lefranc and Lefranc 2001a).

2005b) (IMGT Collier de Perles for the Variable (V), Constant (C), and Groove (G) Domains of Ig, TR, MH, IgSF, and MhSF [Lefranc 2011e]).

Correspondence between domain labels used for the description of amino acid sequences and structures (e.g., in IMGT/3Dstructure-DB) and sequence labels used for the description of nucleotide



**FIGURE 3.** Schematic representation of TR. The TR chain and domain labels (IMGT/3Dstructure-DB) and the corresponding sequence labels (IMGT/LIGM-DB) are shown in Tables 5 and 7. CO, CONNECTING-REGION; TM, TRANSMEMBRANE-REGION; CY, CYTOPLASMIC-REGION. The examples are *Homo sapiens* TR-ALPHA\_BETA and TR-GAMMA1\_DELTA (TR- $\alpha\beta$  and TR- $\gamma1\delta$ ) (Lefranc and Lefranc 2001b).

**Table 5. *Homo sapiens* “CHAIN” labels and relations with the sequence labels and the Molecule\_EntityPrototype leafconcepts**

CHAIN labels (IMGT/3Dstructure-DB)			Sequence labels (IMGT/LIGM-DB)	Molecule_EntityPrototype leafconcepts (labels in the IMGT databases and tools)		
IG-CHAIN	L-CHAIN	L-KAPPA	V-J-C-REGION	V-J-C-CHAIN		
		L-LAMBDA-1				
		L-LAMBDA-2				
		L-LAMBDA-3				
		L-LAMBDA-6				
		L-LAMBDA-7				
		H-CHAIN			V-D-J-C-REGION	V-D-J-C-CHAIN
	H-MU					
	H-DELTA					
	H-GAMMA-1					
	H-GAMMA-2					
	H-GAMMA-3					
	H-GAMMA-4					
	TR-CHAIN		H-ALPHA-1	V-J-C-REGION	V-J-C-CHAIN	
		H-ALPHA-2				
		H-EPSILON				
		TR-ALPHA	V-D-J-C-REGION			V-D-J-C-CHAIN
		TR-BETA-1	V-J-C-REGION			V-J-C-CHAIN
		TR-BETA-2	V-D-J-C-REGION			V-D-J-C-CHAIN
	TR-GAMMA-1	V-J-C-REGION	V-J-C-CHAIN			
	TR-GAMMA-2	V-D-J-C-REGION	V-D-J-C-CHAIN			
	TR-DELTA	V-D-J-C-REGION	V-D-J-C-CHAIN			

sequences (e.g., in IMGT/LIGM-DB) are shown for the IG domains (Table 6; Fig. 2) and the TR domains (Table 7; Fig. 3) from *Homo sapiens*.

## IMGT LABELS: A PARADIGM FOR IMMUNOGENETICS AND ANTIBODY ENGINEERING

With >560 IMGT standardized labels for the nucleotide and amino acid sequences, 2D and 3D structures, IMGT databases and tools provide a detailed and accurate description of IG and TR sequences and structures that is crucial in basic and clinical research (Lefranc 2005, 2008a,b; Ghia et al. 2007; Giudicelli and Lefranc 2009) and in biotechnology related to antibody engineering and humanization (Lefranc 2009; Ehrenmann et al. 2010a). IMGT/mAb-DB (Poiron et al. 2010) allows one to query for therapeutic antibodies for which sequences and 3D structures are available. Novel IMGT chain labels have been defined for the description of new antibody formats and FPIA (Ehrenmann et al. 2010b). Antibodies

**Table 6. Correspondence between domain and sequence labels for *Homo sapiens* IG domains**

Domain labels (IMGT/3Dstructure-DB)		Sequence labels (IMGT/LIGM-DB)			
V-DOMAIN	VL	V-KAPPA	V-J-REGION		
		V-LAMBDA	V-J-REGION		
C-DOMAIN	VH		V-D-J-REGION		
	CL	C-KAPPA	C-REGION		
		C-LAMBDA			
	CH	CH1	CH1	C-REGION <sup>a</sup>	
		CH2	CH2		
CH3		CH3			
	CH4 <sup>b</sup>	CH4 <sup>b</sup>			

<sup>a</sup>The H-CHAIN “C-REGION” also includes the “HINGE-REGION” for the “H-ALPHA,” “H-DELTA,” and “H-GAMMA” chains and, for membrane IG (mIG), the “CONNECTING-REGION” (CO), “TRANSMEMBRANE-REGION” (TM), and “CYTOPLASMIC-REGION” (CY) (Fig. 2); for secreted IG (sIG), the “C-REGION” includes “CH-S” instead of CO, TM, and CY.

<sup>b</sup>For “H-MU” and “H-EPSILON.”

**Table 7. Correspondence between domain and sequence labels for *Homo sapiens* TR domains**

Domain labels (IMGT/3Dstructure-DB)		Sequence labels (IMGT/LIGM-DB)
V-DOMAIN	V-ALPHA	V-J-REGION
	V-BETA	V-D-J-REGION
	V-GAMMA	V-J-REGION
	V-DELTA	V-D-J-REGION
C-DOMAIN	C-ALPHA	C-REGION <sup>a</sup>
	C-BETA	C-REGION <sup>a</sup>
	C-GAMMA	C-REGION <sup>a</sup>
	C-DELTA	C-REGION <sup>a</sup>

<sup>a</sup>The TR-CHAIN “C-REGION” also includes the CONNECTING-REGION (CO), TRANSMEMBRANE-REGION (TM), and CYTOPLASMIC-REGION (CY) (Fig. 3).

and FPIA represent a large number of the pharmaceutical substances submitted to the World Health Organization/International Nonproprietary Names (WHO/INN) Program (General policies for monoclonal antibodies World Health Organization. INN Working Document 09.251. Update 18/12/2009. <http://www.who.int/medicines/services/inn/en/>). The INN definition of antibodies is based on the IMGT-ONTOLOGY concepts of classification (nomenclature), description (labels), and numerotation (IMGT unique numbering) and this information is obtained with the IMGT/DomainGapAlign tool (Lefranc 2011f). Since 2008, amino acid sequences of monoclonal antibodies (mAb, suffix -mab) and of fusion proteins for immune applications (FPIA, suffix -cept) from WHO/INN have been entered in IMGT. These therapeutic applications emphasize the importance of the IMGT-ONTOLOGY concepts in bridging the gap between antibody sequences, 2D and 3D structures.

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# From IMGT-ONTOLOGY CLASSIFICATION Axiom to IMGT Standardized Gene and Allele Nomenclature: For Immunoglobulins (IG) and T Cell Receptors (TR)

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## INTRODUCTION

Since the creation of IMGT, the international ImMunoGeneTics information system in 1989, at New Haven during the 10th Human Genome Mapping Workshop (HGM10), the standardized classification and nomenclature of the immunoglobulins (IG) and T cell receptors (TR) of human and other vertebrate species have been under the responsibility of the IMGT Nomenclature Committee (IMGT-NC). In 1995, following the first demonstration online of the nucleotide database IMGT/LIGM-DB at the 9th International Congress of Immunology in San Francisco, IMGT-NC has become the World Health Organization-International Union of Immunological Societies (WHO-IUIS)/IMGT Nomenclature Subcommittee for IG and TR. As described here, IMGT gene and allele names are based on the concepts of classification of "Group," "Subgroup," "Gene," and "Allele," generated from the IMGT-ONTOLOGY CLASSIFICATION axiom. The IMGT gene nomenclature for IG and TR genes was approved at the international level by the Human Genome Organisation (HUGO) Nomenclature Committee (HGNC) in 1999 and by the WHO-IUIS. The IMGT IG and TR gene names are the official reference for the vertebrate genome projects and, as such, have been entered in IMGT/GENE-DB, the IMGT gene database, in Entrez Gene (National Center for Biotechnology Information [NCBI]), in Ensembl (European Bioinformatics Institute [EBI]), and in the Vega Genome Browser (Wellcome Trust Sanger Institute).

## RELATED INFORMATION

IMGT/GENE-DB, the IMGT gene database, can be found at <http://www.imgt.org>. Following approval by the WHO-IUIS/IMGT Nomenclature Subcommittee for IG and TR, "IMGT Locus in Focus" publications have provided overviews on the standardized IMGT nomenclature of the IG and TR genes and alleles, with functionality and polymorphism, according to the IMGT Scientific chart rules. "IMGT Locus in Focus" publications (available as pdf from IMGT Index>IMGT Locus in Focus <http://www.imgt.org/textes/IMGTindex/imgtfocus.html>) comprise:

- *Homo sapiens* IGHV, IGHD, and IGHJ (Pallarès et al. 1999; Ruiz et al. 1999), IGKV and IGKJ (Barbié and Lefranc 1998), IGLV and IGLJ (Pallarès et al. 1998), TRAV and TRAJ (Scaviner and Lefranc 2000a,b), TRBV, TRBD, and TRBJ (Folch and Lefranc 2000a,b); nomenclature of the *Homo sapiens* IGH (Lefranc 2001a), IGK (Lefranc 2001b), IGL (Lefranc 2001c); protein displays of the human IG (Scaviner et al. 1999) and TR (Folch et al. 2000) variable and joining regions.
- *Mus musculus* IGKV, IGKJ, and IGKC (Martinez and Lefranc 1998; Martinez-Jean et al. 2001), TRAV and TRDV (Bosc and Lefranc 2003), TRBV, TRBD, and TRBJ (Bosc and Lefranc 2000), TRDV, TRDD, and TRDJ (Bosc et al. 2001).
- Teleostei IGH and IGI (Artero and Lefranc 2000a,b).

Abbreviations are explained below in IMGT-ONTOLOGY CONCEPTS OF CLASSIFICATION.

A detailed description of IMGT is provided in **IMGT, The International ImMunoGeneTics Information System** (Lefranc 2011a). Information is also available on **From IMGT-ONTOLOGY IDENTIFICATION Axiom to IMGT Standardized Keywords: For Immunoglobulins (IG), T Cell Receptors**

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(TR), and Conventional Genes (Lefranc 2011b), From IMGT-ONTOLOGY DESCRIPTION Axiom to IMGT Standardized Labels: For Immunoglobulin (IG) and T Cell Receptor (TR) Sequences and Structures (Lefranc 2011c), IMGT Unique Numbering for the Variable (V), Constant (C), and Groove (G) Domains of IG, TR, MH, IgSF, and MhSF (Lefranc 2011d), and IMGT Collier de Perles for the Variable (V), Constant (C), and Groove (G) Domains of IG, TR, MH, IgSF, and MhSF (Lefranc 2011e).

In addition, protocols are available for IMGT/V-QUEST: IMGT Standardized Analysis of the Immunoglobulin (IG) and T Cell Receptor (TR) Nucleotide Sequences (Giudicelli et al. 2011), IMGT/JunctionAnalysis: IMGT Standardized Analysis of the V-J and V-D-J Junctions of the Rearranged Immunoglobulins (IG) and T Cell Receptors (TR) (Giudicelli and Lefranc 2011), IMGT/Collier de Perles: IMGT Standardized Representation of Domains (IG, TR, and IgSF Variable and Constant Domains, MH and MhSF Groove Domains) (Ehrenmann et al. 2011), IMGT/DomainGapAlign: IMGT Standardized Analysis of Amino Acid Sequences of Variable, Constant, and Groove Domains (IG, TR, MH, IgSF, MhSF) (Ehrenmann and Lefranc 2011a), and IMGT/3Dstructure-DB: Querying the IMGT Database for 3D Structures in Immunology and Immunoinformatics (IG or Antibodies, TR, MH, RPI, and FPIA) (Ehrenmann and Lefranc 2011b).

## BACKGROUND INFORMATION

The CLASSIFICATION axiom is an axiom of the Formal IMGT-ONTOLOGY or IMGT-Kaleidoscope (Giudicelli and Lefranc 1999; Lefranc et al. 2004, 2005, 2008; Duroux et al. 2008). The CLASSIFICATION axiom postulates that any molecule, cell, tissue, organ, organism, or population, any process and any relation, has to be **classified**. The CLASSIFICATION axiom has generated the concepts of classification of IMGT-ONTOLOGY, the global reference in immunogenetics and immunoinformatics, built by IMGT (<http://www.imgt.org>) (Lefranc et al. 2009). The CLASSIFICATION axiom is one of seven axioms of the Formal IMGT-ONTOLOGY, the others being "IDENTIFICATION axiom," "DESCRIPTION axiom," "NUMEROTATION axiom," "LOCALIZATION axiom," "ORIENTATION axiom," and "OBTENTION axiom." The IMGT gene and allele nomenclature for IG (Lefranc 2000a; Lefranc and Lefranc 2001a) and TR (Lefranc 2000b; Lefranc and Lefranc 2001b) derives from the concepts of classification generated from the CLASSIFICATION axiom, and among them, the concepts of "Group," "Subgroup," "Gene," and "Allele."

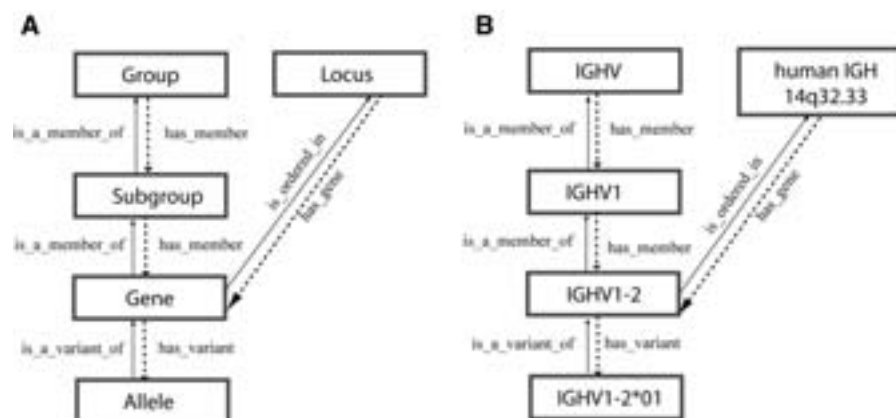
## IMGT-ONTOLOGY CONCEPTS OF CLASSIFICATION

The IMGT-ONTOLOGY CLASSIFICATION axiom has generated the concepts of classification which have been necessary to propose a standardized nomenclature for the IG and TR genes. These concepts take into account the highly polymorphic multigenic loci and families to which the IG and TR belong, their rearrangements, and their allelic polymorphisms. These concepts are used whatever the antigen receptor (IG or TR), whatever the locus (for mammals, e.g., immunoglobulin heavy IGH, immunoglobulin kappa IGK, immunoglobulin lambda IGL, T cell receptor alpha TRA, T cell receptor beta TRB, T cell receptor gamma TRG, and T cell receptor delta TRD), whatever the gene configuration (germline, undefined, or rearranged), and whatever the species, from fish to human. Among the concepts of classification, the "Group," "Subgroup," "Gene," and "Allele" concepts are essential (Fig. 1):

"**Group**" is a concept of classification that allows one to classify a set of genes which belong to the same multigene family, within the same species or between different species. For the IG and TR, the set of genes is identified by the "GeneType" concept: V (for "variable"), D (for "diversity"), J (for "joining"), or C (for "constant"), e.g., "**IGHV**" (Fig. 1). In *Homo sapiens*, the "Group" concept has 24 leafconcepts (a leafconcept is a concept that corresponds to the finest level of granularity), of which 10 are for IG ("IGHV," "IGHD," "IGHJ," "IGHC," "IGKV," "IGKJ," "IGKC," "IGLV," "IGLJ," "IGLC") and 14 are for TR ("TRAV," "TRAJ," "TRAC," "TRBV," "TRBD," "TRBJ," "TRBC," "TRGV," "TRGJ," "TRGC," "TRDV," "TRDD," "TRDJ," "TRDC").

"**Subgroup**" is a concept of classification that allows one to classify a subset of genes which belong to the same group, and which, in a given species, share at least 75% of identity at the nucleotide sequence level (and in the germline configuration for the V, D, and J genes), for instance "***Homo sapiens* IGHV1**" (Fig. 1).

"**Gene**" is a concept of classification that allows one to classify a unit of DNA sequence that can be potentially transcribed and/or translated (this definition includes the regulatory elements in 5' and 3',



**FIGURE 1.** Concepts of classification for gene and allele nomenclature (generated from the IMGT-ONTOLOGY CLASSIFICATION axiom). (A) Hierarchy of the concepts of classification and their relations. (B) Examples of leafconcepts for each concept of classification. They are associated with a “TaxonRank” level, and more precisely for the “Gene” and “Allele” concepts with a leafconcept of “Species” (here, *Homo sapiens*). The “Locus” concept is a concept of localization (LOCALIZATION axiom).

and the introns, if present). The leafconcepts of the “Gene” concept are gene names. In IMGT-ONTOLOGY, a gene name is composed of the name of the species (leafconcept of the “Species” concept) and of the international HGNC/IMGT gene symbol, e.g., “*Homo sapiens* IGHV1-2” (human immunoglobulin heavy variable 1–2) (Fig. 1). By extension, orphans and pseudogenes are also instances of the “Gene” concept. For example, in December 2010, the “Gene” concept had 685 *Homo sapiens* IG and TR gene names (leafconcepts) (of which 442 were for IG and 243 were for TR).

“Allele” is a concept of classification that allows one to classify a polymorphic variant of a gene. The leafconcepts of the “Allele” concept are allele names. In IMGT-ONTOLOGY, an allele name identified by mutations at the nucleotide level is composed of the IMGT gene name with an asterisk (\*) and a two-figure number (for instance, “*Homo sapiens* IGHV1-2\*01”) (Fig. 1). The alleles are classified by reference to allele \*01. Full description of mutations and allele name designations are currently recorded for the core sequences (V-REGION, D-REGION, J-REGION, C-REGION). For example, in December 2010, the “Allele” concept had 1254 *Homo sapiens* IG and TR allele names (leafconcepts) (of which 832 were for IG and 422 were for TR).

## IMGT SCIENTIFIC CHART RULES FOR NOMENCLATURE

IMGT nomenclature for IG and TR gene names of human and other vertebrates, based on the IMGT-ONTOLOGY concepts of classification, follows as closely as possible the rules of the HUGO Gene Nomenclature Committee (<http://www.genenames.org/>). This was applied as early as 1988 for the human IGL and IGH loci (Bensmana et al. 1988; Ghanem et al. 1988) and 1989 for all the genes of the human TRG locus (Lefranc 1989a,b; Lefranc and Rabbitts 1990): **Gene names in capital letters, no greek letters, no commas, no dots** (hyphens are accepted, as for the human leucocyte antigens [HLA] and major histocompatibility [MH] genes). The three first letters indicate the locus: IGH, IGK, IGL, TRA, TRB, TRG, TRD. The fourth letter is V, D, J, or C (except for IGH, see below). The following number(s) and/or letter(s) allow, if necessary, unambiguous identification of the gene, a single number or letter being used whenever it is possible (IGKC, TRGV9, TRGVA, TRAJ4...).

- For the IGH locus, the constant genes are designated by the letter (and eventually number) corresponding to the encoded isotype (IGHM, IGHD, IGHG3,...), instead of using the letter C.
- For the more complex situation where genes have not yet been localized and have to be named accordingly to their subgroup (for V) or cluster (for D and J), a temporary designation is used in which the Arabic number (for the subgroup or cluster) is followed by the letter S (for subgroup or sequential, respectively), in turn followed by the number of the gene in the subgroup or cluster.

- Orphan genes are designated by the IMGT four letter gene designation, followed by a number for the subgroup (if known), followed by a slash, OR (for orphan), the chromosome number (if known), a dash, and a specific gene number and/or letter.
- Examples include IGKV2/OR2-1, IGHD1/OR15-1a, IGKV1/OR-1. In the IG and TR HGNC symbols, slashes and parentheses are omitted, and capital letters replace the lower-case letters found in some provisional IMGT gene names. Otherwise, the gene symbols and all the full names (including slashes and parentheses) are identical in IMGT and HGNC nomenclatures.

Genes and alleles are recorded in IMGT/GENE-DB (Giudicelli et al. 2005) and displayed in Alignments of alleles (IMGT Repertoire, <http://www.imgt.org>).

## INTEROPERABILITY BETWEEN IMGT, HGNC, AND NCBI GENE

Since the creation of IMGT, in 1989, at New Haven during HGM10, the standardized classification and nomenclature of the IG and TR of human and other vertebrate species have been under the responsibility of the IMGT-NC. The WHO-IUIS/IMGT Nomenclature Subcommittee for IG and TR has received official delegation from HGNC for the IG and TR genes and alleles (Wain et al. 2002). The IMGT gene nomenclature for IG and TR genes (Lefranc 2000a,b; Lefranc and Lefranc 2001a,b) was approved by the HGNC in 1999 and by the WHO-IUIS (Lefranc 2007, 2008). IMGT IG and TR gene names have been entered in IMGT/GENE-DB (Giudicelli et al. 2005), the IMGT gene database, in the Human Genome Database (GDB), in LocusLink at the NCBI in 1999–2000, in Entrez Gene (NCBI) (Maglott et al. 2007) when this gene database superseded LocusLink, in MapViewer (NCBI), in Ensembl at the EBI in 2006, and in the Vega Genome Browser at the Wellcome Trust Sanger Institute.

The IMGT nomenclature is acknowledged as the official standard for IG and TR by the scientific community (Glusman et al. 2001; Lefranc 2005, 2007, 2008, 2011f) and the IMGT IG and TR gene names are the official reference for the vertebrate genome projects. Indeed the antigen receptors (IG and TR) that characterize the adaptive immune response of the vertebrates with jaws (gnathostomes) have specific characteristics compared to the conventional genes. It is through the IMGT-ONTOLOGY concepts that the IG and TR genes are now managed in the genome databases. The first unified presentation of the locus “maps” of the seven human loci (IGH, IGK, IGL, TRA, TRB, TRG, and TRD) (Lefranc 2000c,d) has become the standard for the IG and TR locus representation in other vertebrate species (Bosc and Lefranc 2003; Lefranc and Lefranc 2004) (IMGT Repertoire, <http://www.imgt.org>).

To facilitate interoperability between the databases, IMGT works in close collaboration with HGNC (Wain et al. 2004; Bruford et al. 2008), NCBI Gene (Maglott et al. 2007), the Mouse Genomic Nomenclature Committee (MGNC) (to which IMGT provided all the mouse IG and TR genes in 2002), the Nomenclature Committees of newly sequenced genomes, the national and international societies (Immunology, Immunogenetics, Genetics, Antibody), and editors and publishers for recommendations to authors.

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# IMGT Unique Numbering for the Variable (V), Constant (C), and Groove (G) Domains of IG, TR, MH, IgSF, and MhSF

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## INTRODUCTION

The “IMGT unique numbering” (or “IMGT\_unique\_numbering”) concept is a major concept of numerotation (generated from the NUMEROTATION axiom) of IMGT-ONTOLOGY, the global reference in immunogenetics and immunoinformatics, built by IMGT, the international ImMunoGeneTics information system. The “IMGT unique numbering” concept, described here, allows one to number domain types that are characteristic of protein superfamilies, whatever the species, the molecule type or the chain type. Three leafconcepts (a leafconcept is a concept that corresponds to the finest level of granularity) have been defined, respectively, for the variable (V) domain and constant (C) domain of the immunoglobulin superfamily (IgSF) and for the groove (G) domain of the major histocompatibility (MH) superfamily (MhSF). The IMGT unique numbering concept has been a great breakthrough in immunogenetics and systems biology in allowing, for the first time, the bridging of the gap between amino acid (and nucleotide) sequences of any V, C, or G domain and their two-dimensional (2D) and three-dimensional (3D) structures. The IMGT unique numbering concept has been fundamental in the creation of the IMGT Collier de Perles and in the standardization of the description of mutations, amino acid changes, polymorphisms, and contact analysis in the IMGT databases, tools, and web resources.

## RELATED INFORMATION

A detailed description of IMGT is provided in **IMGT, the International ImMunoGeneTics Information System** (Lefranc 2011a). The NUMEROTATION axiom is an axiom of the Formal IMGT-ONTOLOGY or IMGT-Kaleidoscope (Giudicelli and Lefranc 1999; Lefranc et al. 2004, 2005a, 2008; Duroux et al. 2008). The NUMEROTATION axiom postulates that any molecule, cell, tissue, organ, organism, or population, any process and any relation, has to be numbered. The NUMEROTATION axiom has generated the concepts of numerotation of IMGT-ONTOLOGY, the global reference in immunogenetics and immunoinformatics, built by IMGT (<http://www.imgt.org>; Lefranc et al. 2009). The NUMEROTATION axiom is one of seven axioms of the Formal IMGT-ONTOLOGY, the others being “IDENTIFICATION axiom” (From IMGT-ONTOLOGY IDENTIFICATION Axiom to IMGT Standardized Keywords: **For Immunoglobulins (IG), T Cell Receptors (TR), and Conventional Genes** [Lefranc 2011b]), “DESCRIPTION axiom” (From IMGT-ONTOLOGY DESCRIPTION Axiom to IMGT Standardized Labels: **For Immunoglobulin (IG) and T Cell Receptor (TR) Sequences and Structures** [Lefranc 2011c]), “CLASSIFICATION axiom” (From IMGT-ONTOLOGY CLASSIFICATION Axiom to IMGT Standardized Gene and Allele Nomenclature: **For Immunoglobulins (IG) and T Cell Receptors (TR)** [Lefranc 2011d]), “LOCALIZATION axiom,” “ORIENTATION axiom,” and “OBTENTION axiom.” The IMGT unique numbering (Lefranc 1997, 1999; Lefranc et al. 2003, 2005b,c) is one of the major concepts of numerotation, generated from the NUMEROTATION axiom, the other being the IMGT Collier de Perles (**IMGT Collier de Perles for the Variable (V), Constant (C), and Groove (G) Domains of IG, TR, MH, IgSF, and MhSF** [Lefranc 2011e]).

Protocols are available for **IMGT/V-QUEST: IMGT Standardized Analysis of the Immunoglobulin (IG) and T Cell Receptor (TR) Nucleotide Sequences** (Giudicelli et al. 2011), **IMGT/JunctionAnalysis: IMGT Standardized Analysis of the V-J and V-D-J Junctions of the Rearranged Immunoglobulins**

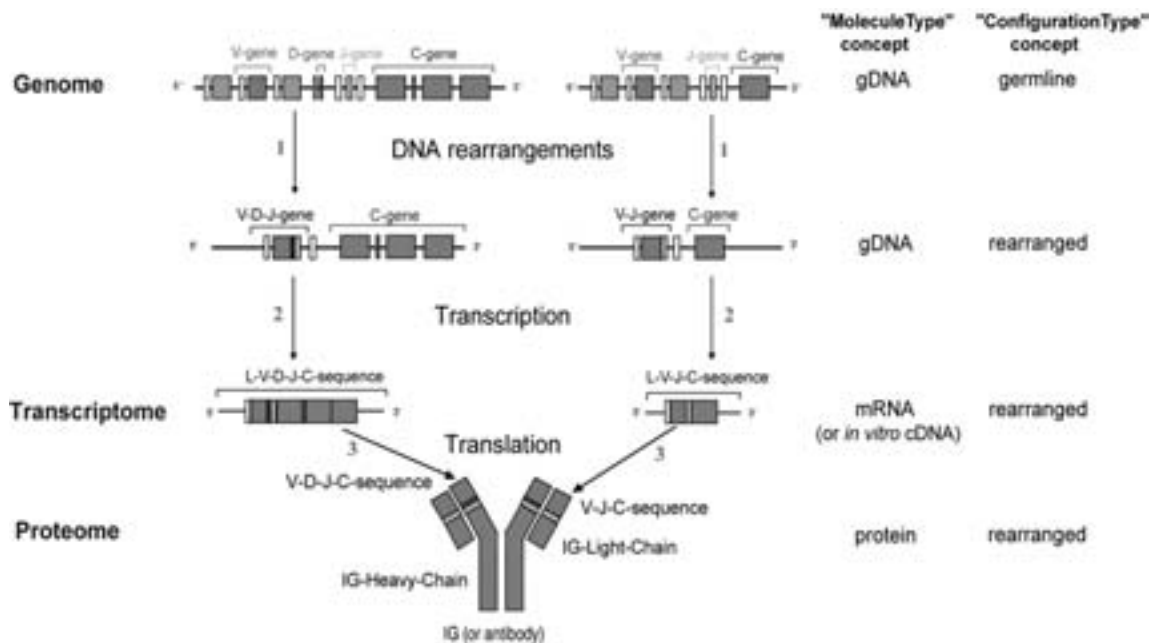
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(IG) and T Cell Receptors (TR) (Giudicelli and Lefranc 2011), IMGT/Collier de Perles: IMGT Standardized Representation of Domains (IG, TR, and IgSF Variable and Constant Domains, MH and MhSF Groove Domains) (Ehrenmann et al. 2011), IMGT/DomainGapAlign: IMGT Standardized Analysis of Amino Acid Sequences of Variable, Constant, and Groove Domains (IG, TR, MH, IgSF, MhSF) (Ehrenmann and Lefranc 2011a), and IMGT/3Dstructure-DB: Querying the IMGT Database for 3D Structures in Immunology and Immunoinformatics (IG or Antibodies, TR, MH, RPI, and FPIA) (Ehrenmann and Lefranc 2011b).

## BACKGROUND INFORMATION

The immunoglobulin (IG) and T cell receptor (TR) variable domains form a huge repertoire of  $2 \times 10^{12}$  antigen receptors per individual and, owing to the particularities of their synthesis that involve DNA rearrangements (Fig. 1), there was a need for a systematic and coherent numbering of the amino acids and codons, whatever the molecule, configuration, or chain type. It was therefore a great breakthrough when the “IMGT unique numbering” and its representation, the IMGT Collier de Perles (IMGT Collier de Perles for the Variable (V), Constant (C), and Groove (G) Domains of IG, TR, MH, IgSF, and MhSF [Lefranc 2011e]), were defined for the first time in 1997 for the V domain (Lefranc 1997, 1999). This IMGT unique numbering was created by taking into account the high conservation of the structure of the V domain and by integrating knowledge acquired by the analysis of multiple sources: alignment of >5000 sequences, literature data on the framework (FR) and complementarity determining regions (CDR), structural data from X-ray diffraction studies, and characterization of the CDR hypervariable loops (Lefranc 1997, 1999). The standardized delimitation of the FR-IMGT and CDR-IMGT, a new and crucial feature of this numbering, was defined based on the longest CDR1-IMGT and CDR2-IMGT found in the IMGT multiple alignments of the germline IG and TR genes and, for the rearranged CDR3-IMGT, on statistical analysis of the IG and TR rearrangements (Lefranc 1997, 1999; Lefranc et al. 2003). The IMGT unique numbering, originally defined for the numerotation of the IG and TR V-DOMAIN (Lefranc 1997), was rapidly extended to the V-LIKE-DOMAIN of the IgSF other than IG and TR (Lefranc 1999; Lefranc et al. 2003), then to the C domain (C-DOMAIN of IG and TR and C-LIKE-DOMAIN of IgSF other than IG and TR) (Lefranc et al. 2005a). Based on the same concepts, and despite a very different structure of the G domain, the IMGT unique numbering for G domain



**FIGURE 1.** Synthesis of an IG or antibody in humans. A human being may potentially synthesize  $10^{12}$  different antibodies (Lefranc and Lefranc 2001a). 1, DNA rearrangements (is\_rearranged\_into); 2, Transcription (is\_transcribed\_into); 3, Translation (is\_translated\_into). The configuration of C-GENE is “undefined” (ConfigurationType) (IMGT Repertoire, <http://www.imgt.org>).

was successfully set up for the G-DOMAIN of MH and G-LIKE-DOMAIN of MhSF other than MH (Lefranc et al. 2005b).

## IMGT UNIQUE NUMBERING FOR V DOMAIN

### V Domain Strands and Loops

The IMGT unique numbering for V domain numbers the V-DOMAIN of IG or antibodies and TR and the V-LIKE-DOMAIN of the IgSF other than IG and TR. The V domain strands and loops and their IMGT positions and lengths, based on the IMGT unique numbering for V domain (V-DOMAIN and V-LIKE-DOMAIN) (Lefranc et al. 2003), are shown in Table 1.

The V domain (V-DOMAIN of IG and TR and V-LIKE-DOMAIN of IgSF other than IG and TR) is composed of the A-STRAND of 15 (or 14 if gap at position 10) amino acids (positions 1 to 15), the B-STRAND of 11 amino acids (positions 16 to 26) with the first conserved cysteine (1st-CYS) at position 23, the BC-LOOP (positions 27 to 38; the longest BC loops have 12 amino acids), the C-STRAND of eight amino acids (positions 39 to 46) with the tryptophan (CONSERVED-TRP) at position 41, the C'-STRAND of nine amino acids (positions 47 to 55), the C'C''-LOOP (positions 56 to 65; the longest C'C'' loops have 10 amino acids), the C''-STRAND of nine (or eight if gap at position 73) amino acids (positions 66 to 74), the D-STRAND of 10 (or eight if gaps at positions 81 and 82) amino acids (positions 75 to 84), the E-STRAND of 12 amino acids (positions 85 to 96) with a conserved hydrophobic amino acid at position 89, the F-STRAND of eight amino acids (positions 97 to 104) with the second conserved cysteine (2nd-CYS) at position 104, the FG-LOOP (positions 105 to 117; these positions correspond to a FG loop of 13 amino acids), and the G-STRAND of 11 (or 10) amino acids (positions 118 to 128) (Table 1; Fig. 2). In the IG and TR V-DOMAIN, the G-STRAND is the C-terminal part of the J-REGION, with J-PHE or J-TRP 118 and the canonical motif F/W-G-X-G at positions 118–121 (Table 1; Lefranc et al. 2003).

In the IG and TR V-DOMAIN, the structurally conserved antiparallel beta strands are also designated as framework regions (FR-IMGT), whereas the loops are designated as complementarity determining

**Table 1. V domain strands and loops, IMGT positions and lengths, based on the IMGT unique numbering for V domain (V-DOMAIN and V-LIKE-DOMAIN)**

V domain strands and loops <sup>a</sup>	IMGT positions	Lengths <sup>b</sup>	Characteristic Residue@Position <sup>c</sup>	V-DOMAIN FR-IMGT and CDR-IMGT
A-STRAND	1–15	15 (14 if gap at 10)		FR1-IMGT
B-STRAND	16–26	11	1st-CYS 23	
BC-LOOP	27–38	12 (or less)		CDR1-IMGT
C-STRAND	39–46	8	CONSERVED-TRP 41	FR2-IMGT
C'-STRAND	47–55	9		
C'C''-LOOP	56–65	10 (or less)		CDR2-IMGT
C''-STRAND	66–74	9 (or 8 if gap at 73)		FR3-IMGT
D-STRAND	75–84	10 (or 8 if gaps at 81, 82)		
E-STRAND	85–96	12	hydrophobic 89	
F-STRAND	97–104	8	2nd-CYS 104	
FG-LOOP	105–117	13 (or less, or more)		CDR3-IMGT
G-STRAND	118–128	11 (or 10)	V-DOMAIN J-PHE 118 or J-TRP 118 <sup>d</sup>	FR4-IMGT

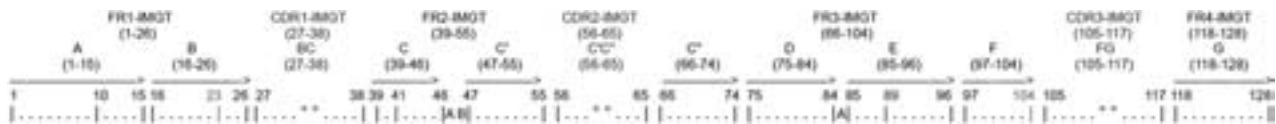
From Lefranc et al. (2003).

<sup>a</sup>IMGT labels (concepts of description) are written in capital letters (**From IMGT-ONTOLOGY DESCRIPTION Axiom to IMGT Standardized Labels: For Immunoglobulin (IG) and T Cell Receptor (TR) Sequences and Structures** [Lefranc 2011c]).

<sup>b</sup>In number of amino acids (or codons).

<sup>c</sup>Residue@Position is an IMGT concept of numerotation that numbers the position of a given residue (or that of a conserved property amino acid class), based on the IMGT unique numbering.

<sup>d</sup>In the IG and TR V-DOMAIN, the G-STRAND (or FR4-IMGT) is the C-terminal part of the J-REGION, with J-PHE or J-TRP 118 and the canonical motif F/W-G-X-G at positions 118–121.



**FIGURE 2.** “IMGT Protein display for V domain” header. The header comprises seven lines. Line 1: Framework regions FR-IMGT (e.g. FR1-IMGT) and complementarity determining regions CDR-IMGT (e.g. CDR1-IMGT). Line 2: start and end positions of FR-IMGT and CDR-IMGT, e.g. (1–26). Line 3: name of strands (A, B...) and loops (AB, BC...). Line 4: start and end positions of strands, e.g. (1–15). Line 5: arrows (for the strands). Line 6: positions according to the IMGT unique numbering for V-DOMAIN and V-LIKE-DOMAIN (Lefranc 1999; Lefranc et al. 2003). Line 7: Pipes for the start and end positions of strands and loops and highlighted positions, dots for the other positions, plus if needed, letters or numbers for additional positions (for example AB or 123...). Optionally small circles above the line (instead of dots) may indicate the positions at the top of the BC, C'C'', and FG loops considering loop lengths of [12.10.13] (these positions are 32, 33 for BC, 60, 61 for C'C'', and 111, 112 for FG).

regions (CDR-IMGT) (Lefranc et al. 2003). Strands A and B correspond to the FR1-IMGT (positions 1 to 26), strands C and C' to the FR2-IMGT (positions 39 to 55), strands C', D, E, and F to the FR3-IMGT (positions 66 to 104), and strand G to the FR4-IMGT (positions 118 to 128). The BC, C'C'', and FG loops correspond to the CDR1-IMGT, CDR2-IMGT, and CDR3-IMGT, respectively (Table 1; Fig. 2; Lefranc et al. 2003).

The loop length (number of amino acids (or codons), that is the number of occupied positions) is a crucial and original concept of IMGT-ONTOLOGY. The lengths of the BC (CDR1-IMGT), C'C'' (CDR2-IMGT), and FG (CDR3-IMGT) loops characterize the V-DOMAIN and V-LIKE-DOMAIN. Thus, the length of the three loops BC, C'C'', and FG is shown, in number of amino acids (or codons), in brackets and separated by dots. For example [9.6.9] means that the BC, C'C'', and FG loops (or CDR1-IMGT, CDR2-IMGT, and CDR3-IMGT for V-DOMAIN) have a length of 9, 6, and 9 amino acids (or codons), respectively.

### IMGT Gaps and Additional Positions

Dots in IMGT Protein displays (Fig. 2; Scaviner et al. 1999; Folch et al. 2000; Lefranc and Lefranc 2001a,b) and hatched circles or squares in IMGT Colliers de Perles for V domain (**IMGT Collier de Perles for the Variable (V), Constant (C), and Groove (G) Domains of IG, TR, MH, IgSF, and MhSF** [Lefranc 2011e]) correspond to missing positions according to the IMGT unique numbering for V domain (Lefranc et al. 2003).

For BC, C'C'', or FG loops shorter than 12, 10, 13 amino acids, respectively, gaps are created at the apex (missing positions, hatched in IMGT Collier de Perles, or not shown in structural data representations). The gaps are placed at the apex of the loop with an equal number of amino acids (or codons) on both sides if the loop length is an even number, or with one more amino acid (or codon) in the left part if it is an odd number. For example, for FG loops <13 amino acids, gaps are created from the apex of the loop, in the following order: 111, 112, 110, 113, 109, etc. For FG loops >13 amino acids, additional positions are created, between positions 111 and 112 at the top of the FG loop, in the following order: 112.1, 111.1, 112.2, 111.2, 112.3, etc. (Lefranc et al. 2003).

## IMGT UNIQUE NUMBERING FOR C DOMAIN

### C Domain Strands, Loops, and Turns

The IMGT unique numbering for C domain numbers the C-DOMAIN of IG or antibodies and TR and the C-LIKE-DOMAIN of the IgSF other than IG and TR. The C domain strands, turns, and loops and their IMGT positions and lengths, based on the IMGT unique numbering for C domain (C-DOMAIN and C-LIKE-DOMAIN) (Lefranc et al. 2005a), are shown in Table 2.

The C domain (C-DOMAIN of IG and TR and C-LIKE-DOMAIN of IgSF other than IG and TR) is composed of the A-STRAND of 15 (or 14 if gap at 10) amino acids (positions 1 to 15), the AB-TURN (additional positions 15.1, 15.2, and 15.3; the longest AB-TURN have three amino acids), the B-STRAND of 11 amino acids (positions 16 to 26) with the 1st-CYS at position 23, the BC-LOOP (positions 27 to 31, 34 to 38), the C-STRAND of seven amino acids (positions 39 to 45) with the CONSERVED-TRP at position 41, the CD-STRAND of one to nine amino acids (additional positions 45.1 to 45.9), the D-STRAND of eight (or seven if gap at 82) amino acids (positions 77 to 84), the DE-TURN (additional

**Table 2. C domain strands, turns, and loops, IMGT positions and lengths, based on the IMGT unique numbering for C domain (C-DOMAIN and C-LIKE-DOMAIN)**

C domain strands, turns and loops <sup>a</sup>	IMGT positions	Lengths <sup>b</sup>	Characteristic Residue@Position <sup>c</sup>
A-STRAND	1–15	15 (14 if gap at 10)	
AB-TURN	15.1–15.3	0–3	
B-STRAND	16–26	11	1st-CYS 23
BC-LOOP	27–31 34–38	10 (or less)	
C-STRAND	39–45	7	CONSERVED-TRP 41
CD-STRAND	45.1–45.9	0–9	
D-STRAND	77–84	8 (or 7 if gap at 82)	
DE-TURN	84.1–84.7 85.1–85.7	0–14	
E-STRAND	85–96	12	hydrophobic 89
EF-TURN	96.1–96.2	0–2	
F-STRAND	97–104	8	2nd-CYS 104
FG-LOOP	105–117	13 (or less, or more)	
G-STRAND	118–128	11 (or less)	

From Lefranc et al. (2005a).

<sup>a</sup>IMGT labels (concepts of description) are written in capital letters (**From IMGT-ONTOLOGY DESCRIPTION Axiom to IMGT Standardized Labels: For Immunoglobulin (IG) and T Cell Receptor (TR) Sequences and Structures** [Lefranc 2011c]).

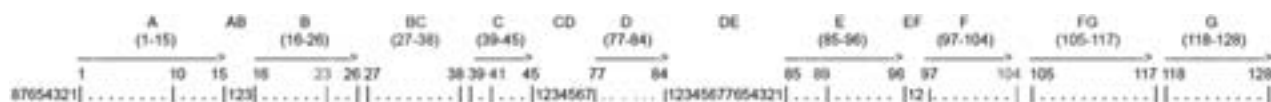
<sup>b</sup>In number of amino acids (or codons).

<sup>c</sup>Residue@Position is a IMGT concept of numerotation that numbers the position of a given residue (or that of a conserved property amino acid class), based on the IMGT unique numbering.

positions 84.1 to 84.7 and from the apex 85.7 to 85.1, corresponding to 14 amino acids), the E-STRAND of 12 amino acids (positions 85 to 96) with a conserved hydrophobic amino acid at position 89, the EF-TURN (additional positions 96.1 and 96.2, corresponding to two amino acids), the F-STRAND of eight amino acids (positions 97 to 104) with the 2nd-CYS at position 104, the FG-LOOP (positions 105 to 117, these positions corresponding to an FG loop of 13 amino acids), and the G-STRAND of 11 (or less) amino acids (positions 118 to 128) (Table 2; Fig. 3; Lefranc et al. 2005a).

### C Domain and V Domain Comparison

The A-STRAND and B-STRAND of the C domain are similar to those of the V domain (Lefranc et al. 2005a). The longest BC-LOOP of the C domain have 10 amino acids (missing positions 32 and 33), instead of 12 amino acids in the V domain. The C-STRAND and the D-STRAND of the C domain are shorter by one position (46) and two positions (75, 76), respectively, compared to those of the V domain. The transversal CD-STRAND is a characteristic of the C domain (a V domain has instead two anti-parallel beta strands C'-STRAND and C''-STRAND linked by the C'C''-LOOP). The E-STRAND, F-STRAND, and G-STRAND of the C domain are similar to those of the V domain (Lefranc et al. 2005a) (IMGT Scientific chart rules, <http://www.imgt.org>).



**FIGURE 3.** “IMGT Protein display for C domain” header. The header comprises five lines. Line 1: name of strands (A, B...), turns, and loops (AB, BC...). Line 2: start and end positions of strands, e.g. (1-15). Line 3: arrows (for the strands). Line 4: positions according to the IMGT unique numbering for C-DOMAIN and C-LIKE-DOMAIN (Lefranc et al. 2005b). Missing positions (32 and 33 in the BC loop and 46 to 76) in the C domain by comparison with the V domain are not shown. Line 5: Pipes for the start and end positions of strands and loops and high-lighted positions, dots for the other positions, plus if needed, numbers for additional positions (for example: 123...) (Lefranc et al. 2005a).

## IMGT Gaps and Additional Positions

Dots in IMGT Protein displays (Fig. 3) and hatched circles or squares in IMGT Colliers de Perles for C domain (**IMGT Collier de Perles for the Variable (V), Constant (C), and Groove (G) Domains of IG, TR, MH, IgSF, and MhSF** [Lefranc 2011e]) correspond to missing positions according to the IMGT unique numbering for C domain (Lefranc et al. 2005a).

The longest BC-LOOP of the C domain have 10 amino acids (missing positions 32 and 33, that are a feature of the C domain are not shown in the IMGT Colliers de Perles and IMGT Protein displays for C domain). For BC loops <10 amino acids, gaps are created from the apex in the following order: 34, 31, 35, 30, 36, etc. The FG-LOOP of the C domain is similar to that of the V domain. Gaps for FG loops <13 amino acids and additional positions for FG loops >13 amino acids are created following the same rules as those of the V domain.

Additional positions in the C domain define the AB-TURN, DE-TURN, and EF-TURN (Table 2). For AB-TURN shorter than three amino acids, gaps are created (hatched in IMGT Colliers de Perles, or not shown in structural data representations) in a decreasing ordinal manner. For DE-TURN <14 amino acids, gaps are created in the following order: 85.7, 84.7, 85.6, 84.6, 85.5, etc. For EF-TURN shorter than two amino acids, gaps are created in the following order: 96.2, 96.1 (Lefranc et al. 2005a).

## IMGT UNIQUE NUMBERING FOR G DOMAIN

### G Domain Strands, Turns, and Helix

The IMGT unique numbering for G domain numbers the G-DOMAIN of MH and the G-LIKE-DOMAIN of MhSF other than MH. The G domain strands, turns, and helix and their IMGT positions and lengths, based on the IMGT unique numbering for G domain (G-DOMAIN and G-LIKE-DOMAIN) (Lefranc et al. 2005b), are shown in Table 3.

The G domain (G-DOMAIN of MH and G-LIKE-DOMAIN of MhSF other than MH) comprises a sheet of four antiparallel beta strands linked by turns and a helix that sits on the beta strands, its axis forming an angle of ~40 degrees with the strands. Two G domains are needed to form the MhSF groove made of a "floor" and two "walls." Each G domain contributes by its strands and turns to half of the groove floor and by its helix to one wall of the groove (Lefranc et al. 2005b).

**Table 3. G domain strands, turns, and helix, IMGT positions and lengths, based on the IMGT unique numbering for G domain (G-DOMAIN and G-LIKE-DOMAIN)**

G domain strands, turns, and helix <sup>a</sup>	IMGT positions	Lengths <sup>b</sup>	Characteristic Residue@Position <sup>c</sup> and additional positions <sup>d</sup>
A-STRAND	1–14	14	7A, CYS-11
AB-TURN	15–17	3 (or 2 or 0)	
B-STRAND	18–28	11 (or 10 <sup>e</sup> )	
BC-TURN	29–30	2	
C-STRAND	31–38	8	
CD-TURN	39–41	3 (or 1 <sup>f</sup> )	
D-STRAND	42–49	8	49.1 to 49.5
HELIX	50–92	43 (or less or more)	54A, 61A, 61B, 72A, CYS-74, 92A

From Lefranc et al. (2005b).

<sup>a</sup>IMGT labels (concepts of description) are written in capital letters (**From IMGT-ONTOLOGY DESCRIPTION Axiom to IMGT Standardized Labels: For Immunoglobulin (IG) and T Cell Receptor (TR) Sequences and Structures** [Lefranc 2011c]).

<sup>b</sup>In number of amino acids (or codons).

<sup>c</sup>Residue@Position is a IMGT concept of numerotation that numbers the position of a given residue (or that of a conserved property amino acid class), based on the IMGT unique numbering.

<sup>d</sup>For details on the characteristic Residue@Position and additional positions, see text (Lefranc et al. 2005b).

<sup>e</sup>Or 9 in some G-BETA (Lefranc et al. 2005b).

<sup>f</sup>Or 0 in some G-ALPHA2-LIKE (Lefranc et al. 2005b).



The G domain is composed of the A-STRAND of 14 amino acids (positions 1 to 14), the AB-TURN of three or less amino acids (positions 15 to 17), the B-STRAND of 11 or less amino acids (positions 18 to 28), the BC-TURN of two amino acids (positions 29 and 30), the C-STRAND of eight amino acids (positions 31 to 38), the CD-TURN of three or less amino acids (positions 39 to 41), the D-STRAND of eight amino acids (positions 42 to 49), and a helix of 43 (or less or more) amino acids (positions 50 to 92) (Table 3; Fig. 4; Lefranc et al. 2005b).

The helix is split into two parts separated by a kink, positions 58 of G-ALPHA1, 61 of G-ALPHA2, 63 of G-ALPHA, and 62 of G-BETA being the “highest” points on the groove floor (Lefranc et al. 2005b).

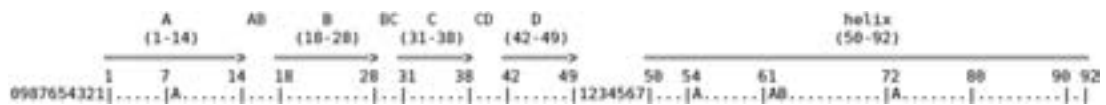
### IMGT Gaps and Additional Positions

Dots in IMGT Protein displays (Fig. 4) and hatched circles or squares in IMGT Collier de Perles for G domain (**IMGT Collier de Perles for the Variable (V), Constant (C), and Groove (G) Domains of IG, TR, MH, IgSF, and MhSF** [Lefranc 2011e]) correspond to missing positions according to the IMGT unique numbering for G domain (Lefranc et al. 2005b).

The gaps are localized in the turns. The AB-TURN (positions 15 to 17) comprises three amino acids in the G-ALPHA1, G-ALPHA2, and G-BETA domains but these positions are unoccupied in the G-ALPHA domains (as well as position 18 of B-STRAND). The BC-TURN (positions 29 and 30) comprises two positions that are occupied in all G-DOMAIN. The CD-TURN (positions 39 to 41) is occupied by three amino acids in the G-ALPHA1 domains and by one amino acid in most of the other G domains (Lefranc et al. 2005b).

The additional position 7A represents a bulge in 3D structures and is present in some G-ALPHA domains, for instance those of the human HLA-DQA1 and HLA-DOA, and mouse H2-AA and H2-DOA chains (Lefranc et al. 2005b). Additional positions at the N terminus of A-STRAND or at the C terminus of D-STRAND can be added if necessary. Thus, two additional positions (1.2 and 1.1) are added at the N terminus of the A-STRAND of the G-ALPHA domains as the presence of these two amino acids was demonstrated by protein sequencing of the HLA-DRA. However the proteolytic cleavage site of the leader peptide (L-REGION) needs to be confirmed experimentally for the other G-ALPHA (Lefranc et al. 2005b). In each [D1] G-DOMAIN (except the G-ALPHA domain of human HLA-DMA and mouse H2-DMA discussed below), the amino acid at position 1 (shown within parentheses in IMGT Protein displays, <http://www.imgt.org>) is encoded by the codon that results from the splicing between the first exon (EX1) that encodes the L-REGION and the second exon (EX2) that encodes [D1] (Lefranc et al. 2005b). Ten amino acids have been added at positions 1.10 to 1.1 of HLA-DMA and H2-DMA but it is necessary to confirm experimentally if they belong, or not, to the mature protein. Four additional positions (49.1 to 49.4) are also observed at the C terminus of D-STRAND of the G-BETA domain of HLA-DMB and H2-DMB1 (Lefranc et al. 2005b).

Interestingly, despite the high sequence divergence, only five additional positions in the helix (54A, 61A, 61B, 72A, and 92A) are necessary to align **any** G domain (Lefranc et al. 2005b). Three of them (61A, 61B, 72A) characterize the G-ALPHA2 and/or G-BETA domains. Indeed, positions 61A and 72A are occupied in the G-ALPHA2 domain, whereas positions 61A, 61B, and 72A are occupied in G-BETA domains (except for the mouse H2-AB chain). The position 92A is only occupied in the HLA-DMA and H2-DMA G-ALPHA domains. It is worthwhile to note that position 54A in G-ALPHA1-LIKE is the only additional position needed to extend the IMGT numbering for G-DOMAIN to the G-LIKE-DOMAIN of the MhSF proteins other than MH, or related proteins of the immune system (RPI)-MH1Like (Lefranc et al. 2005b).



**FIGURE 4.** “IMGT Protein display for G domain” header. The header comprises five lines. Line 1: name of strands (A, B, C, D), turns (AB, BC, CD), and helix. Line 2: start and end positions of strands (1–14), (18–28), (31–38), (42–49), and helix (50–92). Line 3: arrows (for the strands) and line (for the helix). Line 4: positions according to the IMGT unique numbering for G-DOMAIN and G-LIKE-DOMAIN (Lefranc et al. 2005c). Line 5: Pipes for the start and end positions of strands and highlighted positions, dots for the other positions, plus if needed, letters or numbers for additional positions that characterize some G domains (for example: A, AB or 123...) (Lefranc et al. 2005b).



## G Domain Characteristics Comparison

Two cysteine, CYS-11 (in strand A) and CYS-74 (in the helix), are well conserved in the G-ALPHA2 and G-BETA domains where they participate in a disulfide bridge that fastens the helix on the groove floor. The G-ALPHA1 and G-ALPHA domains have a conserved N-glycosylation site at position 86 (N-X-S/T, where N is asparagine, X is any amino acid except proline, S is serine, and T is threonine). An N-glycosylation site is also found at position 86 in the G-ALPHA2 domain of the mouse classical MH1 chains (H2-D1, H2-K1, and H2-L). The G-BETA domains (except for the human HLA-DMB and mouse H2-DMB1 chains) have a conserved N-glycosylation site at position 15 (AB-TURN) (Lefranc et al. 2005b).

Interestingly, the G-ALPHA domains of the HLA-DMA and H2-DMA chains have specific features compared to the other G-ALPHA domains and share common characteristics with the G-ALPHA2 and G-BETA domains: there is a conserved CYS-11\_CYS-74 disulfide bridge, positions 61A and 61B are occupied (as in the G-BETA domains), and there is no N-glycosylation site at position 86 (Lefranc et al. 2005b) (IMGT Protein display in IMGT Repertoire, <http://www.imgt.org>).

Practically, the IMGT unique numbering for positions 1–39 and 73–92 of the G-ALPHA2 domains can be obtained very easily by subtracting 90 from the mature protein numbering (91–129 and 163–182). Between these positions, the two gaps (at positions 40 and 41) and the two insertions (at positions 61A and 72A) are necessary, in the IMGT unique numbering, to allow meaningful sequence and structure alignment and comparison between the G-ALPHA1 and G-ALPHA2 sequences.

## BRIDGING THE GAP BETWEEN IMMUNOGENETICS AND SYSTEMS BIOLOGY

By allowing one to number the V and C domains of the IG, TR, and IgSF other than IG and TR, and the G domain of the MH and MhSF other than MH (Lefranc 1997, 1999; Lefranc et al. 2003, 2005a,b), the IMGT unique numbering has been fundamental in:

- the creation of the IMGT Colliers de Perles for V, C, and G domains (**IMGT Collier de Perles for the Variable (V), Constant (C), and Groove (G) Domains of IG, TR, MH, IgSF, and MhSF** [Lefranc 2011e])
- the definition of the Residue@Position concept (standardized numerotation of amino acid changes and polymorphisms)
- the standardized numerotation of mutations (at the codon and nucleotide level) and definition of alleles
- the delimitation of the FR-IMGT and CDR-IMGT for antibody engineering and antibody humanization
- the comparison of interactions and contact analysis between domains in 3D structures

The IMGT unique numbering bridges the gap between amino acid (and nucleotide) sequences and 3D structures and is essential for domain comparison in molecular immunogenetics (Bertrand et al. 2004; Duprat et al. 2004; Frigoul and Lefranc 2005; Garapati and Lefranc 2007), in structural biology and immunology for receptor/ligand interaction analysis (Kaas and Lefranc 2005; Kaas et al. 2008), and in biotechnology related to antibody engineering and humanization (Lefranc 2009; Ehrenmann et al. 2010a). The IMGT unique numbering is used (i) for the annotation of sequences, genes, and structures in the IMGT databases: IMGT/LIGM-DB (Giudicelli et al. 2006), IMGT/CLL-DB (Brochet 2008), IMGT/GENE-DB (Giudicelli et al. 2005), IMGT/2Dstructure-DB and IMGT/3Dstructure-DB (Kaas et al. 2004; Ehrenmann et al. 2010b), (ii) for the analysis of sequences and structures in the IMGT online tools: IMGT/V-QUEST (Giudicelli et al. 2004; Brochet et al. 2008; Giudicelli and Lefranc 2009), IMGT/JunctionAnalysis (Yousfi Monod et al. 2004; Bleakley et al. 2006), IMGT/HighV-QUEST (Alamyar et al. 2010), IMGT/Phylogene (Elemento and Lefranc 2003), IMGT/GenelInfo (Baum et al. 2004, 2006), IMGT/LIGMotif (Lane et al. 2010), IMGT/DomainGapAlign, IMGT/DomainDisplay and IMGT/Collier de Perles (Ehrenmann et al. 2010b), and (iii) in the data management of the IMGT knowledge web resources: IMGT Protein displays, IMGT Alignments of alleles, IMGT Table of alleles, IMGT Colliers de Perles, etc. (<http://www.imgt.org>) (**IMGT, the International ImMunoGeneTics Information System** [Lefranc 2011a]).

As a major IMGT-ONTOLOGY concept of numerotation, the IMGT unique numbering is used, with the IMGT-ONTOLOGY concepts of classification (**From IMGT-ONTOLOGY CLASSIFICATION Axiom to IMGT Standardized Gene and Allele Nomenclature: For Immunoglobulins (IG) and T Cell Receptors (TR)** [Lefranc 2011d]) and concepts of description (**From IMGT-ONTOLOGY DESCRIPTION Axiom to**

**IMGT Standardized Labels: For Immunoglobulin (IG) and T Cell Receptor (TR) Sequences and Structures [Lefranc 2011c]**), in the definition of monoclonal antibodies (mAb, suffix -mab) and fusion proteins for immune applications (FPIA, suffix -cept) of the World Health Organization/International Nonproprietary Name (WHO/INN) programme (Lefranc 2011f).

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# IMGT Collier de Perles for the Variable (V), Constant (C), and Groove (G) Domains of IG, TR, MH, IgSF, and MhSF

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## INTRODUCTION

The “IMGT Collier de Perles” (or “IMGT\_Collier\_de\_Perles”) concept is a major concept of numerotation (generated from the NUMEROTATION axiom) of IMGT-ONTOLOGY, the global reference in immunogenetics and immunoinformatics, built by IMGT, the international ImMunoGeneTics information system. The “IMGT Collier de Perles” concept, described here, allows standardized two-dimensional (2D) graphical representations of the domains, based on the IMGT unique numbering. Three leafconcepts (a leaf-concept is a concept that corresponds to the finest level of granularity) have been defined: for the variable (V) domain and constant (C) domain of the immunoglobulin superfamily (IgSF) and for the groove (G) domain of the major histocompatibility (MH) superfamily (MhSF). IMGT Colliers de Perles are obtained, starting from V, C, or G domain amino acid sequences, using IMGT/DomainGapAlign and IMGT/Collier de Perles tools. In IMGT/3Dstructure-DB, IMGT Colliers de Perles of V and C domains are provided with hydrogen bonds and those of G domains with IMGT pMH contact analysis. IMGT Colliers de Perles allows one to bridge the gap between sequences and three-dimensional (3D) structures, whatever the species, the IgSF or MhSF protein, or the chain type. They are particularly useful for antibody engineering, sequence-structure analysis, visualization and comparison of positions for mutations, polymorphisms and contact analysis of immunoglobulins (IG), T cell receptors (TR), MH, and related proteins of the immune system (RPI) belonging to the IgSF and MhSF.

## RELATED INFORMATION

A detailed description of IMGT is provided in **IMGT, the International ImMunoGeneTics Information System** (Lefranc 2011a). The NUMEROTATION axiom is an axiom of the Formal IMGT-ONTOLOGY or IMGT-Kaleidoscope (Giudicelli and Lefranc 1999; Lefranc et al. 2004, 2005a, 2008; Duroux et al. 2008). The NUMEROTATION axiom postulates that any molecule, cell, tissue, organ, organism or population, any process and any relation, has to be numbered. The NUMEROTATION axiom has generated the concepts of numerotation of IMGT-ONTOLOGY, the global reference in immunogenetics and immunoinformatics, built by IMGT (<http://www.imgt.org>) (Lefranc et al. 2009). The NUMEROTATION axiom is one of seven axioms of the Formal IMGT-ONTOLOGY, the others being “IDENTIFICATION axiom” (**From IMGT-ONTOLOGY IDENTIFICATION Axiom to IMGT Standardized Keywords: For Immunoglobulins (IG), T Cell Receptors (TR), and Conventional Genes**) (Lefranc 2011b), “DESCRIPTION axiom” (**From IMGT-ONTOLOGY DESCRIPTION Axiom to IMGT Standardized Labels: For Immunoglobulin (IG) and T Cell Receptor (TR) Sequences and Structures**) (Lefranc 2011c), “CLASSIFICATION axiom” (**From IMGT-ONTOLOGY CLASSIFICATION Axiom to IMGT Standardized Gene and Allele Nomenclature: For Immunoglobulins (IG) and T Cell Receptors (TR)**) (Lefranc 2011d), “LOCALIZATION axiom”, “ORIENTATION axiom”, and “OBTENTION axiom”. The IMGT Collier de Perles (Lefranc 1999; Ruiz and Lefranc 2002; Kaas and Lefranc 2007; Kaas et al. 2007) and the IMGT unique numbering (Lefranc 1997, 1999; Lefranc et al. 2003, 2005b,c) (**IMGT Unique Numbering for the Variable (V), Constant (C), and Groove (G) Domains of IG, TR, MH, IgSF, and MhSF**) (Lefranc 2011e) are the two major concepts of numerotation, generated from the NUMEROTATION axiom.

Protocols are available for **IMGT/V-QUEST: IMGT Standardized Analysis of the Immunoglobulin (IG) and T Cell Receptor (TR) Nucleotide Sequences** (Giudicelli et al. 2011), **IMGT/JunctionAnalysis: IMGT**

**Standardized Analysis of the V-J and V-D-J Junctions of the Rearranged Immunoglobulins (IG) and T Cell Receptors (TR)** (Giudicelli and Lefranc 2011), **IMGT/Collier de Perles: IMGT Standardized Representation of Domains (IG, TR, and IgSF Variable and Constant Domains, MH and MhSF Groove Domains)** (Ehrenmann et al. 2011), **IMGT/DomainGapAlign: IMGT Standardized Analysis of Amino Acid Sequences of Variable, Constant, and Groove Domains (IG, TR, MH, IgSF, MhSF)** (Ehrenmann and Lefranc 2011a), and **IMGT/3Dstructure-DB: Querying the IMGT Database for 3D Structures in Immunology and Immunoinformatics (IG or Antibodies, TR, MH, RPI, and FPIA)** (Ehrenmann and Lefranc 2011b).

## BACKGROUND INFORMATION

A standardized approach for immunogenetics and immunoinformatics was strongly needed in the 1990s. The first IMGT Collier de Perles to be put online was the VH domain from mouse E5.2 Fv in December 1997, still available in IMGT Repertoire (<http://www.imgt.org>) in its original drawing. By creating the IMGT Colliers de Perles (Lefranc 1999; Ruiz and Lefranc 2002; Kaas and Lefranc 2007; Kaas et al. 2007), based on the IMGT unique numbering (Lefranc 1997, 1999; Lefranc et al. 2003, 2005b,c), IMGT has bridged the gap between sequences and 3D structures (Lefranc 2005, 2008a,b; Lefranc et al. 2008). IMGT Collier de Perles can be obtained by the users from their own nucleotide sequences using IMGT/V-QUEST (Giudicelli et al. 2004; Brochet 2008; Brochet et al. 2008; Giudicelli and Lefranc 2009), from their amino acid sequences using IMGT/DomainGapAlign (Ehrenmann et al. 2010b), or whether their amino acid sequences are already gapped according to the IMGT unique numbering, using the IMGT/Collier de Perles tool (<http://www.imgt.org>). IMGT Colliers de Perles are available in IMGT/3Dstructure-DB, with hydrogen bonds for V and C domains, for proteins with known 3D structures, and in IMGT/2Dstructure-DB for therapeutic monoclonal antibodies and fusion proteins for immune applications (FPIA) (Edelmann et al. 2010b; Poirion et al. 2010). IMGT Colliers de Perles are widely used for structural biology and immunology (Kaas and Lefranc 2007; Kaas et al. 2007) and biotechnologies related to antibody engineering and humanization (Lefranc 2009; Ehrenmann et al. 2010a).

## IMGT COLLIER DE PERLES FOR V DOMAIN

“IMGT Collier de Perles for V domain” (or “IMGT\_Collier\_de\_Perles\_for\_V\_domain”) is a leafconcept of the “IMGT Collier de Perles” concept of numerotation. It represents the V domain that includes the V-DOMAIN of IG or antibodies and TR and the V-LIKE-DOMAIN of the IgSF proteins other than IG and TR. It is based on the IMGT unique numbering for V domain (Lefranc 1997, 1999; Lefranc et al. 2003) (**IMGT Unique Numbering for the Variable (V), Constant (C), and Groove (G) Domains of IG, TR, MH, IgSF, and MhSF**) (Lefranc 2011e).

The topology and 3D structure of a V-LIKE-DOMAIN are very similar to those of an IG or TR V-DOMAIN (Fig. 1A).

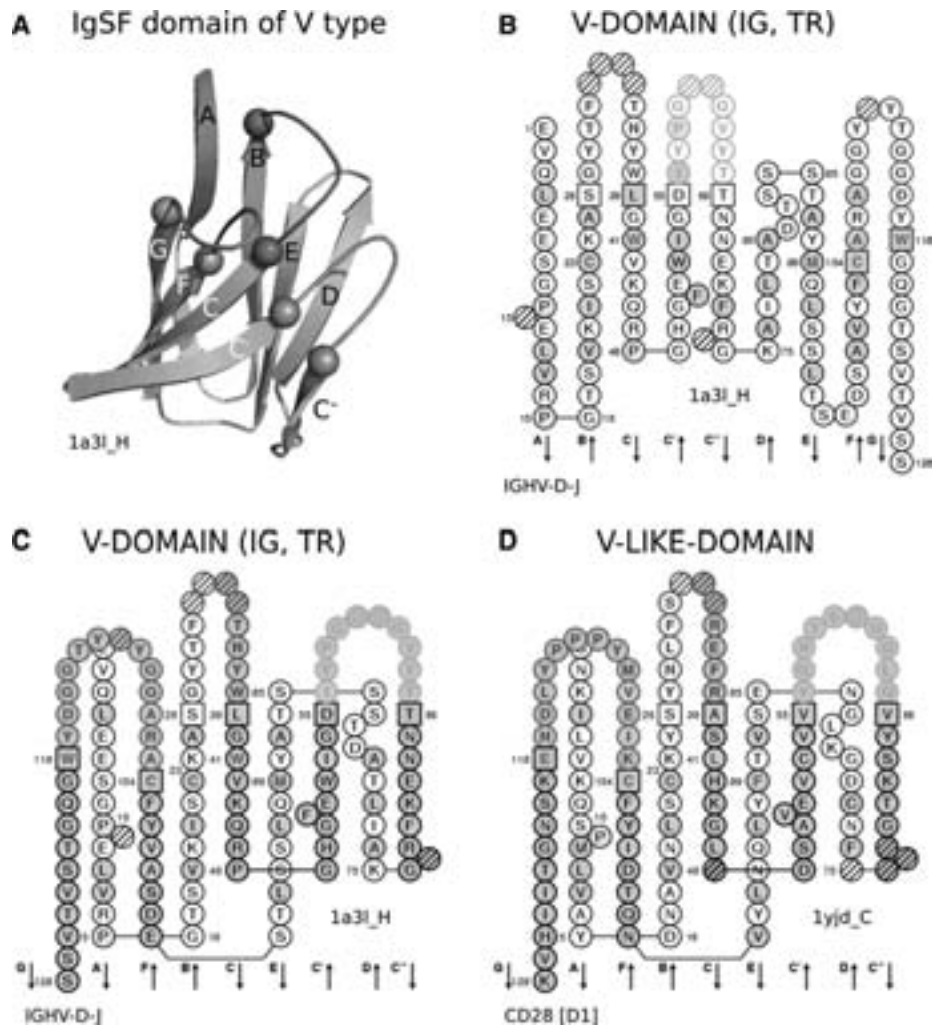
A V domain consists of about 100 amino acids in nine antiparallel  $\beta$  strands (A, B, C, C', C'', D, E, F, and G), linked by  $\beta$  turns (AB, CC', C'' D, DE, and EF) and loops (BC, C'C'', and FG), located on two layers, forming a sandwich of two sheets. The sheets are closely packed against each other through hydrophobic interactions, giving a hydrophobic core and joined together by a disulfide bridge between the B-STRAND in the first sheet and the F-STRAND in the second sheet (Lefranc et al. 2003). Four strands form one sheet and five strands form a second sheet.

It is remarkable that the 3D structure fold of the V domain has been conserved through evolution, despite the sequence divergence between IgSF domains and, even more surprisingly, despite the particularities of IG and TR synthesis compared with the other proteins (Lefranc and Lefranc 2001a,b). Indeed, in a conventional gene, the V-LIKE-DOMAIN is usually encoded by a unique exon, or more rarely by two spliced exons, whereas in IG and TR, the V-DOMAIN results from the rearrangement of two or three types of genes: variable (V) gene, joining (J) gene and, in some loci, diversity (D) gene (Lefranc and Lefranc 2001a,b) (Fig. 2).

The V-LIKE-DOMAIN is usually, as is the IG and TR V-DOMAIN, the most N-terminal (and extracellular) domain of the protein. However, in contrast to the IG and TR V-DOMAIN, which is always unique, the V-LIKE-DOMAIN may be present in several copies in the same protein and interspersed with the C-LIKE-DOMAIN or with domains of other superfamilies.

Based on the IMGT unique numbering, the conserved amino acids always have the same position. For the V domain, five positions are essential for the core structure: cysteine **23** (1st-CYS), tryptophan **41**

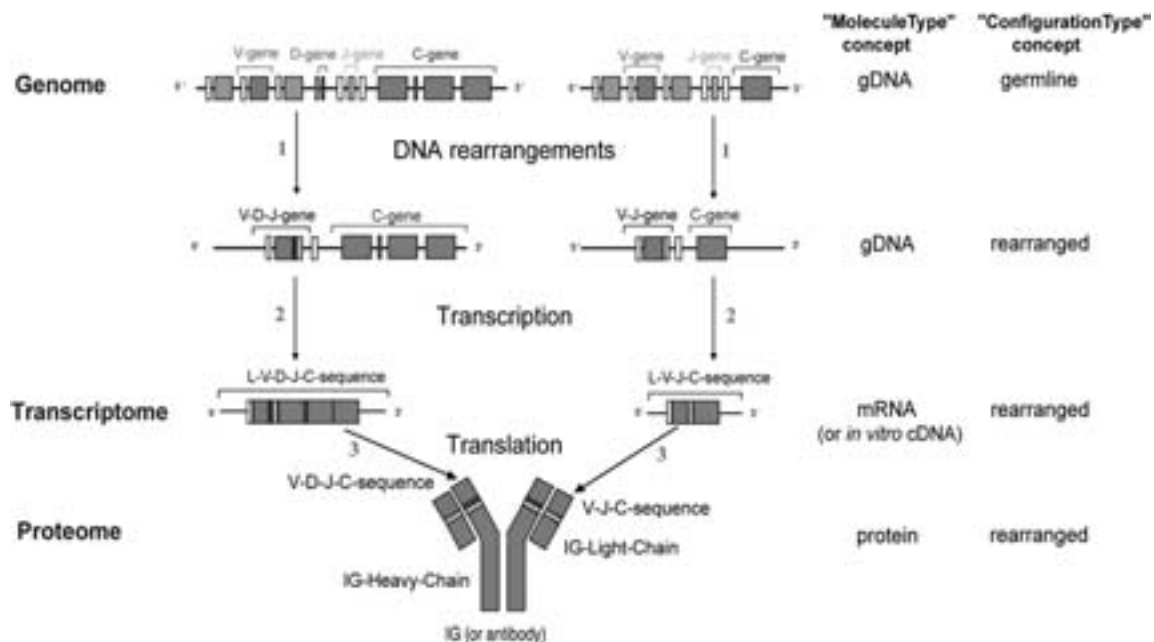




**FIGURE 1.** IMGT Collier de Perles for V domain. (A) Ribbon representation of a V-DOMAIN as an example. A similar topology and 3D structure characterize a V-LIKE-DOMAIN. (B,C) V-DOMAIN on one layer and on two layers, respectively (*Mus musculus* VH [8.8.12]). (D) V-LIKE-DOMAIN on two layers (*Homo sapiens* CD28 [9.9.13]). Amino acids are shown in the one-letter abbreviation. Position at which hydrophobic amino acids (hydropathy index with positive value: I, V, L, F, C, M, A) and tryptophan (W) are found in >50% of analyzed sequences are shown online in blue. All proline (P) are shown online in yellow. The loops BC, C'C'' and FG (corresponding to the CDR-IMGT) are limited by amino acids shown in squares (anchor positions), which belong to the neighboring strands (FR-IMGT). BC loops are represented online in red, C'C'' loops in orange, and FG loops in purple. Hatched circles or squares correspond to missing positions according to the IMGT unique numbering for V domain (Lefranc et al. 2003). Arrows indicate the direction of the  $\beta$  strands and their designations in 3D structures. The IMGT Colliers de Perles on two layers show, in the forefront, the GFCC'C'' strands and, in the back, the ABED strands. IMGT Colliers de Perles on two layers with hydrogen bonds are available in IMGT/3Dstructure-DB (<http://www.imgt.org>): 1a3l\_H, 1yjd\_C.

(CONSERVED-TRP), conserved hydrophobic amino acid **89**, cysteine **104** (2nd-CYS), and "FG anchor" **118** (J-PHE or J-TRP in V-DOMAIN) (Lefranc et al. 2003). In the IG and TR V-DOMAIN, the structurally conserved antiparallel  $\beta$  strands are designated as framework regions (FR-IMGT), whereas the BC, C'C'', and FG loops are designated as complementarity determining regions (CDR-IMGT) (Lefranc et al. 2003). The length (number of amino acids or by extrapolation number of codons, that is the number of occupied positions) of the strands and loops is a crucial and original concept of IMGT-ONTOLOGY. The lengths of the BC (CDR1-IMGT), C'C'' (CDR2-IMGT) and FG (CDR3-IMGT) loops characterize the V domain. Thus, the length of the three loops BC, C'C'', and FG is shown, in number of amino acids (or codons), in brackets and separated by dots. In Figure 1, the lengths are [8.8.12] for the V-DOMAIN CDR-IMGT and [9.9.13] for the V-LIKE-DOMAIN. IMGT Colliers de Perles





**FIGURE 2.** Synthesis of an IG or antibody in humans. A human being may potentially synthesize  $10^{12}$  different antibodies (Lefranc and Lefranc 2001a). (1) DNA rearrangements (is\_rearranged\_into), (2) Transcription (is\_transcribed\_into), (3) Translation (is\_translated\_into). The configuration of C-GENE is "undefined" (ConfigurationType) (IMGT Repertoire, <http://www.imgt.org>).

for V domain can be represented on one layer (Fig. 1B) or two layers (Fig. 1C,D) (Lefranc 1999; Ruiz and Lefranc 2002; Lefranc et al. 2003; Kaas and Lefranc 2007; Kaas et al. 2007). If 3D structures are available, IMGT Colliers de Perles on two layers are provided with hydrogen bonds, in IMGT/3Dstructure-DB (<http://www.imgt.org>) (Ehrenmann et al. 2010b).

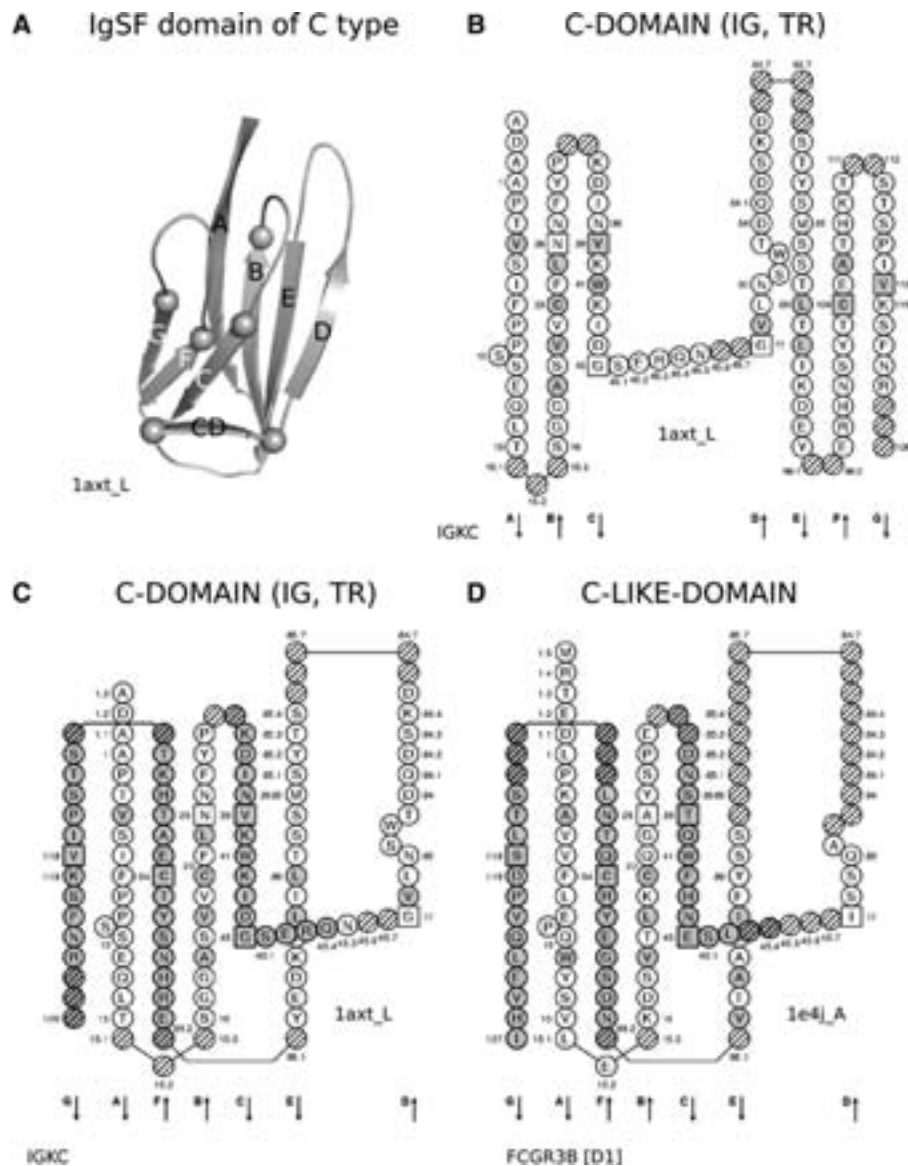
## IMGT COLLIER DE PERLES FOR C DOMAIN

"IMGT Collier de Perles for C domain" (or "IMGT\_Collier\_de\_Perles\_for\_C\_domain") is a leafconcept of the "IMGT Collier de Perles" concept of numerotation. It represents the C domain, which includes the C-DOMAIN of IG or antibodies and TR and the C-LIKE-DOMAIN of IgSF proteins other than IG and TR. It is based on the IMGT unique numbering for C domain (Lefranc et al. 2005c) (**IMGT Unique Numbering for the Variable (V), Constant (C), and Groove (G) Domains of IG, TR, MH, IgSF, and MhSF**) (Lefranc 2011e). The topology and 3D structure of a C-LIKE-DOMAIN are very similar to those of an IG or TR C-DOMAIN (Fig. 3A).

A C domain consists of about 100 amino acids in seven antiparallel  $\beta$  strands (A, B, C, D, E, F, and G), linked by  $\beta$  turns (AB, DE, and EF), a transverse strand (CD), and loops (BC and FG), located in two layers, forming a sandwich of two sheets. The sheets are closely packed against each other through hydrophobic interactions, giving a hydrophobic core, and joined together by a disulfide bridge between the B-STRAND in the first sheet and the F-STRAND in the second sheet (Lefranc et al. 2005c). Four strands form one sheet and three strands form a second sheet.

The C domain has a topology and a 3D structure similar to those of the V domain, but differs by the number of antiparallel  $\beta$  strands (seven instead of nine). By comparison with a V domain, the C'-STRAND, C''-LOOP and C''-STRAND are missing in the C domain and are replaced by the characteristic transverse CD-STRAND that links the two sheets. Depending on the CD-STRAND length, the D-STRAND is in the first or in the second sheet (Lefranc et al. 2005c). Additional positions observed in the C domain define the AB-TURN, DE-TURN, and EF-TURN (Lefranc et al. 2005c).

The IMGT unique numbering for C domain (Lefranc et al. 2005c) is derived from the IMGT unique numbering first described for the V-REGION and for the V domain (Lefranc et al. 2003). The five positions that are essential for the core structure have the same number in the C domain: cysteine **23** (1st-CYS),



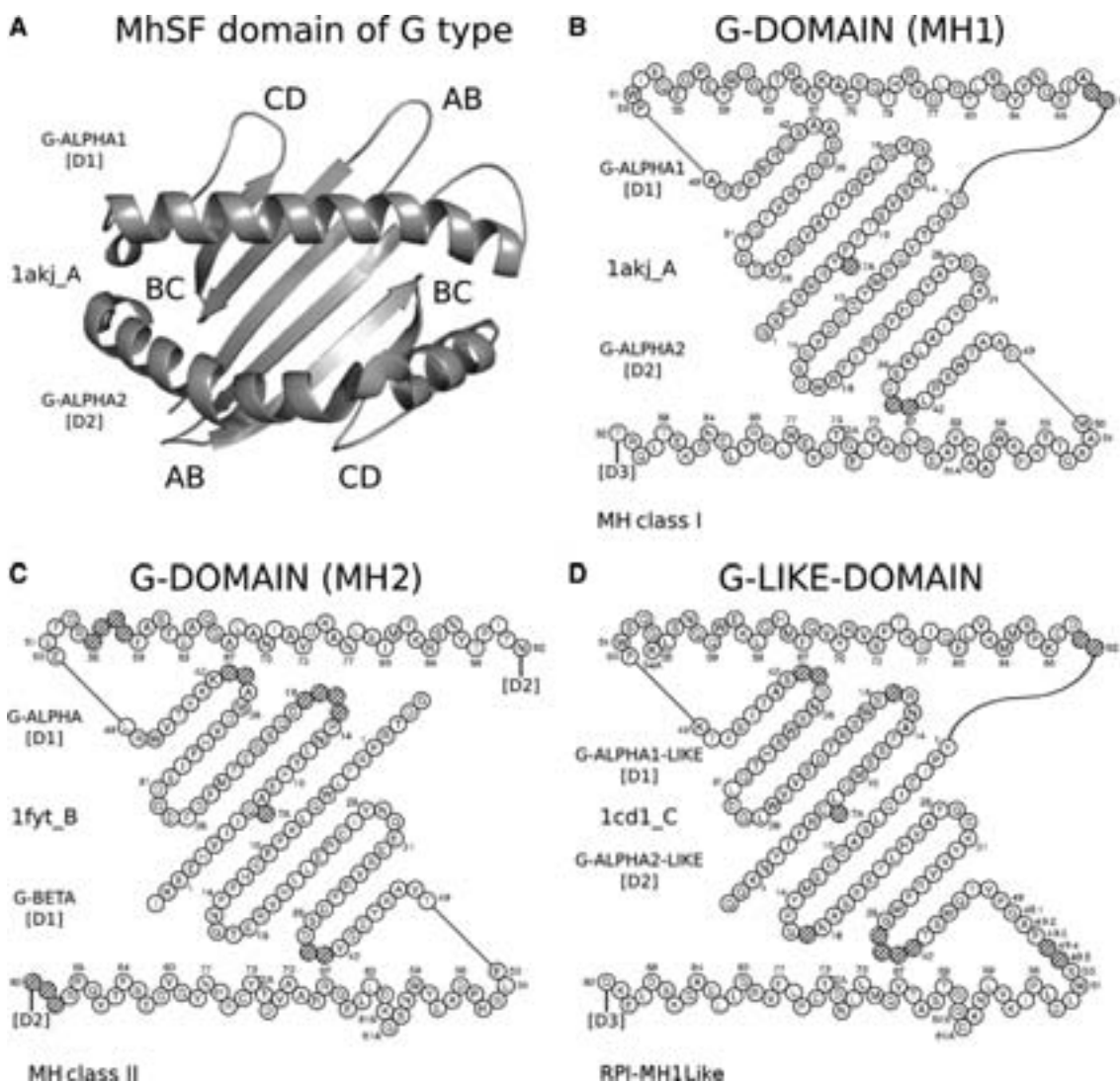
**FIGURE 3.** IMGT Collier de Perles for C domain. (A) Ribbon representation of a C-DOMAIN as an example. A similar topology and 3D structure characterize a C-LIKE-DOMAIN. (B,C) C-DOMAIN on one layer and two layers, respectively (*Mus musculus* C-KAPPA). (D) C-LIKE-DOMAIN on two layers (*Homo sapiens* FCGR3B [D1]). Amino acids are shown in the one-letter abbreviation. Position at which hydrophobic amino acids (hydropathy index with positive value: I, V, L, F, C, M, A) and tryptophan (W) are found in >50% of analyzed sequences are shown in gray. The positions 26, 39, and 104 are shown in squares by homology with the corresponding positions in the V domain. Positions 45 and 77, which delimit the characteristic CD-STRAND of the C domain, and position 118, which corresponds structurally to J-PHE or J-TRP of the IG and TR J REGION (Lefranc and Lefranc 2001a,b), are also shown in squares. Hatched circles correspond to missing positions according to the IMGT unique numbering for C domain (Lefranc et al. 2005c). Arrows indicate the direction of the  $\beta$  strands and their designations in 3D structures. The IMGT Colliers de Perles on two layers show, in the *forefront*, the GFC strands and, in the *back*, the ABE strands. IMGT Colliers de Perles on two layers with hydrogen bonds are available in IMGT/3Dstructure-DB (<http://www.imgt.org>): 1axt\_L, 1e4j\_A.

tryptophan **41** (CONSERVED-TRP), conserved hydrophobic amino acid **89**, cysteine **104** (2nd-CYS), and “FG anchor” **118** (Lefranc et al. 2005c).

IMGT Colliers de Perles for C domain can be represented on one layer (Fig. 3B) or two layers (Fig. 3C, D) (Lefranc et al. 2005c; Kaas and Lefranc 2007; Kaas et al. 2007). If 3D structures are available, IMGT Colliers de Perles on two layers are provided with hydrogen bonds in IMGT/3Dstructure-DB (<http://www.imgt.org>) (Ehrenmann et al. 2010b).

## IMGT COLLIER DE PERLES FOR G DOMAIN

“IMGT Collier de Perles for G domain” (or “IMGT\_Collier\_de\_Perles\_for\_G\_domain”) is a leafconcept of the “IMGT Collier de Perles” concept of numerotation. It represents the G domain that includes the G-DOMAIN of MH and the G-LIKE-DOMAIN of the MhSF proteins other than MH. It is based on the IMGT unique numbering for G domain (Lefranc et al. 2005b) (**IMGT Unique Numbering for the Variable (V), Constant (C), and Groove (G) Domains of IG, TR, MH, IgSF, and MhSF**) (Lefranc 2011e). The topology and 3D structure of a G-LIKE-DOMAIN are very similar to those of an MH G-DOMAIN (Fig. 4A). Two G domains are necessary to form the characteristic groove of the MhSF proteins that



**FIGURE 4.** IMGT Collier de Perles for G domain. (A) Ribbon representation of two G-DOMAINS as an example. A similar topology and 3D structure characterize the G-LIKE-DOMAIN. (B) G-DOMAIN of MH class I (MH1). G-ALPHA1 [D1] and G-ALPHA2 [D2] (*Homo sapiens* HLA-A\*0201). (C) G-DOMAIN of MH class II (MH2). G-ALPHA [D1] and G-BETA [D1] (*Homo sapiens* HLA-DRA\*0101 and HLA-DRB1\*0101). (D) G-LIKE-DOMAIN of RPI-MH1Like. G-ALPHA1-LIKE [D1] and G-ALPHA2-LIKE [D2] (*Mus musculus* CD1D1). Amino acids are shown in the one-letter abbreviation. Hatched circles correspond to missing positions according to the IMGT unique numbering for G domain (Lefranc et al. 2005b). In IMGT Colliers de Perles, position 7A is only displayed in the G-ALPHA, G-ALPHA1, and G-ALPHA1-LIKE domains, and positions 61A and 61B in the G-BETA, G-ALPHA2, and G-ALPHA2-LIKE domains. Position 54A is only occupied in G-ALPHA1-LIKE of RPI-MH1like proteins. Position 92A is only added for HLA-DMA and H2-DMA IMGT Colliers de Perles. Note that the N-terminal end of a peptide in the cleft would be on the left side. IMGT Colliers de Perles are available in IMGT/3Dstructure-DB (<http://www.imgt.org>): 1akj\_A, 1fyt\_B, 1cd1\_C.

comprise the MH (MH class I or MH1, and MH class II or MH2) and the RPI-MH1Like proteins (Lefranc et al. 2005b).

A G domain consists of about 90 amino acids in four antiparallel  $\beta$  strands (A, B, C, and D), linked by turns (AB, BC, CD), forming half of the groove floor, and one  $\alpha$  helix, forming one wall of the groove. For each G domain, the positions that contribute to the groove floor comprise positions 1–49, corresponding to the four strands linked by turns (Lefranc et al. 2005b). The numbering of the G domain helix starts at position 50 and ends at position 92 (Fig. 4).

Interestingly, despite the high sequence divergence, only five additional positions (54A, 61A, 61B, 72A, and 92A) are necessary to align any G-DOMAIN and G-LIKE-DOMAIN (Lefranc et al. 2005b). It is worthwhile to note that position 54A in G-ALPHA1-LIKE is the only additional position needed to extend the IMGT numbering for G-DOMAIN to the G-LIKE-DOMAIN.

The helix (positions 50–92) sits on the  $\beta$  sheet and its axis forms an angle of about 40 degrees with the  $\beta$  strands. Two cysteines, CYS-11 (in strand A) and CYS-74 (in the helix) are well conserved in the G-ALPHA2, G-BETA, and G-ALPHA2-LIKE domains, where they participate to a disulfide bridge that fastens the helix on the groove floor. The IMGT Collier de Perles allows one to describe specific features (Lefranc et al. 2005b; Kaas et al. 2007; Kaas and Lefranc 2007). As an example, the G-ALPHA1 and G-ALPHA domains have a conserved N-glycosylation site at position 86 (N-X-S/T, where N is asparagine, X is any amino acid except proline, S is serine, and T is threonine).

If 3D structures of peptide/MH are available, IMGT Colliers de Perles are provided with IMGT pMH contact analysis, in IMGT/3Dstructure-DB (<http://www.imgt.org>) (Kaas et al. 2004, 2008; Kaas and Lefranc 2005; Ehrenmann et al. 2010b).

## WHAT DO IMGT COLLIERS DE PERLES TEACH US?

1. Any domain represented by an IMGT Collier de Perles is characterized by the **length** of its strands, loops, and turns and, for the G domain, by the length of its helix (Lefranc 1997, 1999; Lefranc et al. 2003, 2005b,c) (**IMGT Unique Numbering for the Variable (V), Constant (C), and Groove (G) Domains of IG, TR, MH, IgSF, and MhSF**) (Lefranc 2011e). The strand, loop, turn, or helix lengths (number of amino acids or by extrapolation number of codons, that is the number of occupied positions) become crucial information that characterizes the domains. This first feature of the IMGT standardization based on the IMGT unique numbering allowed us, for instance, to show that the distinction between the C1, C2, I1, and I2 domain types found in the literature and in the databases to describe the IgSF C domains is unnecessary and moreover inapplicable when dealing with sequences for which no structural data are known (discussed in Lefranc et al. 2005c).
2. A second feature of the IMGT standardization is the comparison of cDNA and/or amino acid sequences with **genomic** sequences, and the identification of the splicing sites, to delimit precisely the domains: a V-LIKE-DOMAIN, a C-DOMAIN, a C-LIKE-DOMAIN, a G-DOMAIN, or a G-LIKE-DOMAIN is usually encoded by a unique exon or more rarely by two spliced exons (Lefranc et al. 2003, 2005b,c). This IMGT standardization for the domain delimitation explains the discrepancies observed with the generalist UniProt/Swiss-Prot database, which identifies domains based on amino acid sequences and does not take into account the genomic information. The IMGT Collier de Perles also addresses the question of the leader region. Indeed, the N-terminal end of the first domain of an IgSF or MhSF chain depends on the proteolytic cleavage site of the leader region (peptide signal), which is rarely determined experimentally. When this site is not known, the IMGT Colliers de Perles start with the first amino acid resulting from the splicing (usually a splicing frame 1) (“Splicing sites” in IMGT Aide-mémoire, <http://www.imgt.org>). For an IG and TR V-DOMAIN, the leader proteolytic site is known (or is extrapolated) and the IMGT Colliers de Perles start with the first amino acid of the V-REGION (Lefranc and Lefranc 2001a,b).
3. The IMGT Colliers de Perles allow a precise visualization of the differences between proteins and/or between species for the IgSF V and C domain strands and loops, and MhSF G domain strands and helix, even in the absence of 3D structures. This has been applied, for example, to the CD28 family members and their B7 family ligands and to the BTLA proteins that belong to the IgSF by their V and/or C domains (Garapati and Lefranc 2007) and to other members of the IgSF (Bertrand et al. 2004; Duprat et al. 2004) and MhSF (Frigoul and Lefranc 2005). The IMGT Colliers de Perles are particularly useful in molecular engineering and antibody humanization design based on CDR grafting. Indeed, they allow one to precisely define the CDR-IMGT and to easily compare the amino acid



sequences of the four FR-IMGT (FR1-IMGT: positions 1–26, FR2-IMGT: 39–55, FR3-IMGT: 66–104, and FR4-IMGT: 118–128) between the mouse (or other species) and the closest human V-DOMAIN. Analyses performed on humanized therapeutic antibodies underline the importance of a correct delimitation of the CDR regions to be grafted (Lefranc 2009; Ehrenmann et al. 2010a).

4. The IMGT Colliers de Perles also allow a comparison to the IMGT Colliers de Perles **statistical profiles** for the human expressed IGHV, IGKV, and IGLV repertoires. These statistical profiles are based on the definition of 11 IMGT amino acid physicochemical characteristics classes that take into account the hydrophathy, volume, and chemical characteristics of the 20 common amino acids (Pommié et al. 2004) (“Amino acids” in IMGT Aide-mémoire, <http://www.imgt.org>). The statistical profiles identified positions that are conserved for the physicochemical characteristics: 41 FR-IMGT positions for the human IGHV and 59 FR-IMGT positions for the human IGKV and IGLV at >80% threshold (see Plate 3 in Pommié et al. 2004). After assignment of the IMGT Collier de Perles amino acids to the IMGT amino acid physicochemical classes, comparison can be made with the statistical profiles of the human expressed repertoires. This comparison is useful to identify potential immunogenic residues at given positions in chimeric or humanized antibodies or to evaluate immunogenicity of therapeutic antibodies.
5. IMGT Colliers de Perles are also of interest when **3D structures** are available. In IMGT/3Dstructure-DB (Ehrenmann et al. 2010b), “IMGT Collier de Perles on 2 layers” are displayed with hydrogen bonds for V and C domains. Clicking on a residue in “IMGT Collier de Perles on one layer” gives access to the corresponding IMGT Residue@Position card, which provides the atom contact types and atom contact categories for that amino acid. IMGT Colliers de Perles display the IMGT pMH contact sites for 3D structures with peptide/MH (pMH) complexes, which can be compared with the pMH contact sites available in IMGT/3Dstructure-DB (Kaas et al. 2004, 2008; Kaas and Lefranc 2005; Ehrenmann et al. 2010b).

The IMGT Colliers de Perles for the V, C, and G domains, based on the IMGT unique numbering, therefore, represent a major step forward for the comparative analysis of the sequences and structures of the IgSF and MhSF domains, for the study of their evolution, and for applications in antibody engineering, IG and TR repertoires in autoimmune diseases and leukemias, pMH contact analysis, and more generally, ligand-receptor interactions involving V, C, and/or G domains.

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# IMGT/V-QUEST: IMGT Standardized Analysis of the Immunoglobulin (IG) and T Cell Receptor (TR) Nucleotide Sequences

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## INTRODUCTION

IMGT/V-QUEST is the integrated online IMGT tool for the standardized analysis of the immunoglobulin (IG) or antibody and T cell receptor (TR) rearranged nucleotide sequences. The analysis of these antigen receptors represents a crucial challenge for the study of the adaptive immune response in normal and disease-related situations. The expressed IG and TR repertoires represent a potential of  $10^{12}$  IG and  $10^{12}$  TR per individual. This huge diversity results from mechanisms that occur at the DNA level during IG molecular synthesis, including the combinatorial rearrangement of the variable (V), diversity (D), and joining (J) genes, the N diversity (V, D, and J exonuclease trimming and addition at random of nucleotides by the terminal deoxynucleotidyl transferase during the V-(D)-J rearrangement) and, for IG, somatic hypermutations. IMGT/V-QUEST, as described here, identifies the V, D, J genes and alleles by alignment with the germline IG and TR gene and allele sequences of the IMGT reference directory. The tool describes the V-REGION mutations and identifies the hot spot positions in the closest germline V gene. IMGT/V-QUEST integrates IMGT/JunctionAnalysis for a detailed analysis of the V-J and V-D-J junctions and IMGT/Automat for a complete sequence annotation of the sequences, and provides IMGT Collier de Perles.

## RELATED INFORMATION

IMGT/V-QUEST (Lefranc 2004; Giudicelli and Lefranc 2005, 2008; Brochet et al. 2008) is part of IMGT, the international ImMunoGeneTics information system, <http://www.imgt.org> (Lefranc et al. 2009). Standardization and IMGT Scientific chart rules are based on the IMGT-ONTOLOGY concepts of identification, classification, description (Giudicelli and Lefranc 1999; Lefranc et al. 2004, 2005a, 2008), and numerotation (Lefranc 1997, 1999; Lefranc et al. 2003, 2005b,c) generated from the axioms of the Formal IMGT-ONTOLOGY or IMGT-Kaleidoscope (Duroux et al. 2008). The concepts of identification led to the IMGT standardized keywords, the concepts of classification to the IMGT standardized gene and allele names, the concepts of description to the IMGT standardized labels, and the concepts of numerotation to the IMGT unique numbering.

A detailed description of IMGT is provided in **IMGT, the International ImMunoGeneTics Information System** (Lefranc 2011a). Information is also available on **From IMGT-ONTOLOGY IDENTIFICATION Axiom to IMGT Standardized Keywords: For Immunoglobulins (IG), T Cell Receptors (TR), and Conventional Genes** (Lefranc 2011b), **From IMGT-ONTOLOGY DESCRIPTION Axiom to IMGT Standardized Labels: For Immunoglobulin (IG) and T Cell Receptor (TR) Sequences and Structures** (Lefranc 2011c), **From IMGT-ONTOLOGY CLASSIFICATION Axiom to IMGT Standardized Gene and Allele Nomenclature: For Immunoglobulins (IG) and T Cell Receptors (TR)** (Lefranc 2011d), **IMGT Unique Numbering for the Variable (V), Constant (C), and Groove (G) Domains of IG, TR, MH, IgSF, and MhSF** (Lefranc 2011e), and **IMGT Collier de Perles for the Variable (V), Constant (C), and Groove (G) Domains of IG, TR, MH, IgSF, and MhSF** (Lefranc 2011f).

In addition, protocols are available for **IMGT/JunctionAnalysis: IMGT Standardized Analysis of the V-J and V-D-J Junctions of the Rearranged Immunoglobulins (IG) and T Cell Receptors (TR)** (Giudicelli and Lefranc 2011), **IMGT/Collier de Perles: IMGT Standardized Representation of Domains (IG, TR, and IgSF Variable and Constant Domains, MH and MhSF Groove Domains)** (Ehrenmann et al. 2011), **IMGT/DomainGapAlign: IMGT Standardized Analysis of Amino Acid Sequences of**

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**Variable, Constant, and Groove Domains (IG, TR, MH, IgSF, MhSF)** (Ehrenmann and Lefranc 2011a), and **IMGT/3Dstructure-DB: Querying the IMGT Database for 3D Structures in Immunology and Immunoinformatics (IG or Antibodies, TR, MH, RPI, and FPIA)** (Ehrenmann and Lefranc 2011b).

## MATERIALS

It is essential that you consult the appropriate Material Safety Data Sheets and your institution's Environmental Health and Safety Office for proper handling of equipment and hazardous materials used in this protocol.

### Equipment

Computer (Internet-connected)

## METHOD

1. Using any modern web browser, access the IMGT Home page, <http://www.imgt.org> and click, in the "IMGT tools" section, on the link to IMGT/V-QUEST.

*This gives access to the IMGT/V-QUEST Welcome page.*

### IMGT/V-QUEST Welcome Page

2. In the IMGT/V-QUEST Welcome page (Fig. 1), locate the following sections:

- i. "Analyze your immunoglobulin (IG) or antibody nucleotide sequences"

*This section shows the species for which the analysis of IG sequences is available.*

*There is no need to specify more precisely the type of IG chain or locus, as the tool will automatically identify whether sequences are IGH, IGK, or IGL by selecting one species or taxon in that section. This works for species with complete IMGT reference directories, such as human (Lefranc 2000a; Lefranc and Lefranc 2001a) and mouse (Giudicelli et al. 2005a). This also holds for the other species or taxons with incomplete IMGT reference directories; however, in those cases, results should be analyzed considering the status of the IMGT reference directory (information on the updates on the web site).*

- ii. "Analyze your T cell Receptor (TR) nucleotide sequences"

*This section shows the species for which the analysis of TR sequences is available.*

*There is no need to specify more precisely the type of TR chain or locus, as the tool will automatically identify whether sequences are TRA, TRB, TRG, or TRD by selecting one species or taxon in that section. This works for species with complete IMGT reference directories, such as human (Lefranc 2000b; Lefranc and Lefranc 2001b) and mouse (Giudicelli et al. 2005a). This also holds true for the other species or taxons with incomplete IMGT reference directories; however, in those cases, results should be analyzed considering the status of the IMGT reference directory (information on the updates on the web site).*

- iii. "Download IMGT reference directory in FASTA format (IG and TR)"

*This section shows the species for which the IMGT reference directory is available.*

*Clicking on one of the links gives access to the data (overview and IMGT/V-QUEST reference directory per species or taxon), if needed.*

3. Choose the IG or TR section (see Step 2), depending on your sequences.
4. Click on the species (e.g., "Human") or taxon that corresponds to the IMGT reference directory against which you want your sequences to be analyzed.

*This will open the IMGT/V-QUEST Search page. That page has a similar display regardless of the choice IG or TR (Step 3) and the species (Step 4).*

### IMGT/V-QUEST Search Page

5. In the IMGT/V-QUEST Search page (Fig. 2), and below the title that reflects your choice IG or TR (Step 3), locate the following.
  - i. "Your selection" that recalls your selection, for example "Human" (Step 4)

# WELCOME ! to IMGT/V-QUEST

THE  
INTERNATIONAL  
IMMUNOGENETICS  
INFORMATION SYSTEM



Citing IMGT/V-QUEST: Brochet, X. et al., Nucl. Acids Res. 36, W503-508 (2008). PMID: 18503082 [more](#)

IMGT/V-QUEST programme version: 3.2.18 (21 March 2011) - IMGT/V-QUEST reference directory release: 201110-1 (7 March 2011)

## Analyse your immunoglobulin (IG) or antibody nucleotide sequences

<ul style="list-style-type: none"><li>Human</li><li>Mouse</li></ul>	<ul style="list-style-type: none"><li>Telocetes</li><li>- Atlantic cod</li><li>- Channel catfish</li><li>- Channel catfish</li><li>- Rainbow trout</li></ul>
<ul style="list-style-type: none"><li>Bat</li><li>Rabbit</li></ul>	<ul style="list-style-type: none"><li>Chondrichthyes</li><li>Shrew</li></ul>

## Analyse your T cell Receptor (TR) nucleotide sequences

<ul style="list-style-type: none"><li>Human</li><li>Mouse</li><li>Camel</li><li>Rainbow trout</li></ul>	<ul style="list-style-type: none"><li>Nonhuman primates</li><li>Bovine</li><li>Dog</li></ul>
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## Download IMGT reference directory in FASTA format (IG and TR)

<ul style="list-style-type: none"><li>Human</li><li>Mouse</li></ul>	<ul style="list-style-type: none"><li>Telocetes</li><li>- Atlantic cod</li><li>- Channel catfish</li><li>- Rainbow trout</li></ul>
<ul style="list-style-type: none"><li>Bat</li><li>Rabbit</li><li>Dog</li><li>Camel</li></ul>	<ul style="list-style-type: none"><li>Chondrichthyes</li><li>Shrew</li><li>Nonhuman primates</li></ul>

FIGURE 1. IMGT/V-QUEST Welcome page.

A note indicates the IMGT reference directory set against which your sequences will be analyzed, e.g., "Human (*Homo sapiens*) IG set," as determined by the tool from Steps 3 and 4. A link provides access to the IMGT reference directory sets.

A link for sequence sets allows one to test IMGT/V-QUEST, if needed (only for human IG sequences).

### ii. The sequence submission section

That section comprises:

Two windows to either type (or copy/paste) your sequence(s) or upload a file, in FASTA format (proceed to Step 6 for entering your sequences).

The "Start" and "Clear the form" buttons.

### iii. "Selection for results"

IMGT/V-QUEST provides the choice between three types of selection for the results:

"A. Detailed view." This displays the results of the analysis for each sequence, individually (proceed to Step 8).

"B. Synthesis view." This displays the alignments of sequences that express the same V gene and allele and allows one to compare the localization of the mutations and the composition of their junctions (proceed to Step 14 for choice of the parameters).

"C. Excel file." This allows the users to get the results of the analysis in a spreadsheet with the choice of 11 sheets (proceed to Step 21 for choice of the parameters).





iv. "Advanced parameters"

"Advanced parameters" allows one to modify the parameters for sophisticated queries or unusual sequences (proceed to Step 11 for choice of "Advanced parameters").

### Entering Nucleotide Sequences for Analysis

6. Copy and paste your sequence(s) in the text area of "Nucleotide sequences," or upload a text file by selecting the option "Or give the path access to a local file containing your sequence(s) in FASTA format" and using the "Browse" button to select the file.

*The analyzed sequences are the V-J or V-D-J regions that encode the variable domain (V-DOMAIN) of the antigen receptors. The analysis is performed for sequences from genomic DNA (gDNA) or complementary DNA (cDNA) without need for the user to specify the molecule type. If the submitted sequences are in reverse orientation, the tool will automatically "complementary reverse" them and will provide the user sequences and the results in the sense orientation. The online IMGT/V-QUEST can analyze up to 50 sequences in a single run. The sequences must be formatted in FASTA format. Note that FASTA headers longer than 30 characters will be truncated in the results.*

7. Depending on the selection you want for your results, proceed to:

- i. Step 8 for A. Detailed view
- ii. Step 14 for B. Synthesis view
- iii. Step 21 for C. Excel file

### IMGT/V-QUEST Detailed View

8. Select the option "A. Detailed view" (selected by default).
9. Select the format of the results, "HTML" (by default) or "Text," and the "Nb of nucleotides per line in alignments" (60 by default).
10. Select, in the light orange box, the displays of the results to be shown.

*There is a choice of 14 different result displays for the analysis of the V-DOMAIN (Lefranc 2004; Giudicelli and Lefranc 2005, 2008; Brochet et al. 2008). The most commonly requested are already selected by default. You can check or uncheck the result displays as needed.*

- i. Alignment for V-GENE
- ii. Alignment for D-GENE
- iii. Alignment for J-GENE
- iv. Results of IMGT/JunctionAnalysis with full list of eligible D-GENE or without list of eligible D-GENE
- v. Sequence of the JUNCTION (nucleotide ["nt"] and amino acid ["AA"])
- vi. V-REGION alignment
- vii. V-REGION translation
- viii. V-REGION protein display
- ix. V-REGION mutation and AA change table
- x. V-REGION mutation and AA change statistics
- xi. V-REGION mutation hot spots
- xii. Sequences of V-, V-J-, or V-D-J-REGION ("nt" and "AA") with gaps, in FASTA and access to IMGT/PhyloGene for V-REGION ("nt")
- xiii. Annotation by IMGT/Automat
- xiv. IMGT Colliers de Perles: link to IMGT/Collier de Perles tool **IMGT/Collier de Perles: IMGT Standardized Representation of Domains (IG, TR, and IgSF Variable and Constant Domains, MH and MhSF Groove Domains)** (Ehrenmann et al. 2011), IMGT Collier de Perles (for a number [nb] of sequences less than five), or no IMGT Collier de Perles

11. Go to “Advanced parameters” at the bottom of the search page and modify the parameters according to the particularities of the sequences, if needed.

*Default values of “Advanced parameters” have been set for classical analysis. They should be modified for sophisticated queries or for unusual sequences (for example, with insertions and/or deletions).*

- i. “Selection of IMGT reference directory set,” with a choice of four sets (“F+ORF,” “F+ORF+in frame P” (by default), “F+ORF including orphans,” or “F+ORF+in frame P including orphans”) where F is functional, ORF is open reading frame, P is pseudogene. This allows sequences to be compared with only relevant sequences (e.g., orphon sequences are relevant for genomic, but not for expressed repertoire studies).

*The selected set can also be chosen either “With all alleles” or “With allele \*01 only.”*

- ii. “Search for insertions and deletions in V-REGION” (“Yes” or “No”).

*If “Yes,” the search is slower and the number of submitted sequences in a single run is limited to 10.*

- iii. “Parameters for IMGT/JunctionAnalysis:” “Nb of accepted D-GENE” (provided for the IGH junctions and for the TRB and TRD junctions, respectively) or “Nb of accepted mutations” in 3’V-REGION, D-REGION, and 5’J-REGION.

- iv. “Parameters for Detailed view:” “Nb of nucleotides to exclude in 5’ of the V-REGION for the evaluation of the nb of mutations” (to avoid, e.g., counting primer specific nucleotides) or “Nb of nucleotides to add (or exclude) in 3’ of the V-REGION for the evaluation of the alignment score” (e.g., in case of low or high exonuclease activity).

12. Click on the “Start” button to launch the analysis.

*“Detailed view” results are displayed in “A. Detailed results for the IMGT/V-QUEST analyzed sequences.” The top of this page indicates the number of analyzed sequences with links to individual results (Fig. 3).*

13. Use the link associated with a sequence name to go directly to the individual results.

*The results include the sequence in FASTA format and a Result summary (Fig. 4 and Fig. 5) and, if selected in Step 10:*

- *The Alignment for V, D, and J genes and alleles (Fig. 6)*
- *The results of IMGT/JunctionAnalysis and the sequence of the JUNCTION (“nt” and “AA”) (Fig. 7)*
- *Different displays of the V-REGION (Fig. 8)*
- *The analysis of the mutations (Fig. 9)*

**THANK YOU**  
for using **IMGT/V-QUEST**

THE  
INTERNATIONAL  
IMMUNOGENETICS  
INFORMATION SYSTEMS



IMGT/V-QUEST programme version: 3.2.18; IMGT/V-QUEST reference directory release: 201110-1

## A. Detailed results for the IMGT/V-QUEST analysed sequences

Number of analysed sequences: 7

seq1 seq2 seq3 seq4 seq5 seq6 seq7

⚠ This release of IMGT/V-QUEST uses [IMGT/JunctionAnalysis](#) for the analysis of the JUNCTION

⚠ Hyphens (-) show nucleotide identity, dots (.) represent gaps

**FIGURE 3.** IMGT/V-QUEST “Detailed view” result page: top of the page. The number of analyzed sequences is indicated. The link associated to a sequence name allows one to go directly to the individual sequence result.

Sequence number 1: seq1

Sequence compared with the human IG set from the IMGT reference directory

```
>seq1
gagggtcagactgttggagctctgggggagggcctggtccagcctggagggctccctgagactc
tccctgtatagcctctgggtccacctccagtagctatccatgacctgggtccggcaggtc
ccaggccaaaggggtggagtgggtggcaagtatccatagacgggaagtatataaatasaag
gtagctccctgagggggcggctcaccatctccagagacactccaaagacagctgtat
ttggaatgacagcctgacagctgaggacagggctgtgtatctgtggaggacagct
tcttttaagcctatgactctctggggccagggaacctgggtcacctctctccagcctcc
accaggggcccctgggtctcccccctggaccctctccaaagagcactctgggggcaaa
ggggcctgggctgctggtaaggactactcccccgaaccggtgacggtgtcgtggac
tcaggcgcctgaccaggggggtgacacctcccccgtgtctccagctccaggaactc
tactcctccagcagctgtgtgacctgacctccagcagcttgggaccagacctacatc
tggacgtgatccaaagcccagcaccaccaggtggcacaagaagttgagccccaaatct
ctgacaaaaatccaca
```

Result summary:	Productive IGH rearranged sequence (no stop codon and in-frame junction)		
V-GENE and allele	IGHV3-30*04	score = 1255	identity = 93.06% (268/288 nt)
J-GENE and allele	IGHJ4*02	score = 204	identity = 91.67% (44/48 nt)
D-GENE and allele by IMGT/JunctionAnalysis	IGHD5-5*01	D-REGION is in reading frame 1	
FR-IMGT lengths, CDR-IMGT lengths and AA JUNCTION	[25.17.38.11]	[8.8.11]	CARTAFFNAYDFW

**FIGURE 4.** IMGT/V-QUEST “Detailed view” result page: *top* part of an individual result with the sequence and the “Result summary.” The *top* part of each individual result indicates the IMGT reference directory set (here, human IG set) against which the sequence was analyzed, and displays the user sequence in FASTA format (a sequence submitted in antisense orientation will be shown as a complementary reverse sequence, that is in V gene sense orientation). The “Result summary” provides a crucial feature that is the evaluation of the user sequence functionality performed by IMGT/V-QUEST: productive (if no stop codon and in-frame junction) or unproductive (if stop codons and/or out-of-frame junction). It also summarizes the main characteristics of the analyzed sequence, which includes (1) the names (with species in the six-letter abbreviation, and the functionality added in the version 3.2.19) of the closest “V-GENE and allele” (e.g., Homsap IGHV3-30\*04 F) and “J-GENE and allele” (e.g., Homsap IGHJ4\*02 F) with the alignment score, the percentage of identity and the ratio “number of identical nucleotides/number of aligned nucleotides,” (2) the name (with species in the six-letter abbreviation, and the functionality) of the closest “D-GENE and allele” (e.g., Homsap IGHV3-30\*04 F) determined by IMGT/JunctionAnalysis (Giudicelli and Lefranc 2011) with the D-REGION reading frame, (3) the framework region (FR)-IMGT lengths (e.g., [25.17.38.11]), the complementarity determining region (CDR)-IMGT lengths (e.g., [8.8.11]), and the amino acid (AA) JUNCTION sequence that characterizes a V domain. Information shown in orange (here, “Productive IGH rearranged sequence” and “93,06%”) are used by clinicians analyzing the level of mutations in chronic lymphocytic leukemia (CLL) IGHV rearranged sequences as a prognostic criterion (Giudicelli and Lefranc 2008). Note that IMGT/V-QUEST provides warnings (data not shown) that appear as notes in red to alert the user if potential insertions or deletions are suspected in the V-REGION (sequences with <85% of identity and/or with different CDR1-IMGT and/or CDR2-IMGT lengths compared with the closest germline V-REGION), or whether other possibilities for the J gene and allele are identified. The seq1 accession number is AB012909 from the IMGT/LIGM-DB database (Giudicelli et al. 2006).

- The localization of the mutation hot spots (Fig. 10)
- The sequences of V-, V-J-, or V-D-J- REGION (“nt” and “AA”) with gaps in FASTA and access to IMGT/PhyloGene for V-REGION (“nt”) (Fig. 11)
- The annotation by IMGT/Automat (Fig. 12)
- The IMGT Collier de Perles or, by default, the link to the IMGT Collier de Perles tool **IMGT/Collier de Perles: IMGT Standardized Representation of Domains (IG, TR, and IgSF Variable and Constant Domains, MH and MhSF Groove Domains)** (Fig. 13; Ehrenmann et al. 2011)

### IMGT/V-QUEST Synthesis View

14. Select the option “B. Synthesis view.”
15. Select the format of the results, “HTML” (by default) or “Text,” and the “Nb of nucleotides per line in alignments” (60 by default).



**1. Alignment for V-GENE and allele identification**

Closest V-REGIONS (evaluated from the V-REGION first nucleotide to the 2nd-CYS codon)

	Score	Identity
<u>L06615</u> IGHV3-30*04	1255	93,06% (248/268 nt)
<u>N83134</u> IGHV3-30*01	1246	92,71% (247/268 nt)
<u>N77300</u> IGHV3-30*09	1246	92,71% (247/268 nt)
<u>N77324</u> IGHV3-30*14	1246	92,71% (247/268 nt)
<u>X92283</u> IGHV3-30-3*01	1246	92,71% (247/268 nt)

**Alignment with FR-IMGT and CDR-IMGT delimitations**

```

seq1
L06615 IGHV3-30*04
N83134 IGHV3-30*01
N77300 IGHV3-30*09
N77324 IGHV3-30*14
X92283 IGHV3-30-3*01

          <----- FR1-IMGT ----->
gaggtgagcgtgtcggagctcggggga...ggcgtggccagcctgggaggtccctgagacctctcctgtatagcctctggattcaccctc
c-----g-----..-----gc-----
c-----g-----..-----gc-----
c-----g-----..-----gc-----
c-----g-----..-----gc-----
c-----g-----..-----gc-----

          CDR1-IMGT <----- FR2-IMGT -----> CDR
... ..agtagctatcctatgacctgggtccggccaggctccaggcaagggcctggagtggtggcaggtatcacatagac...
... ..g-----ca-----gt-----t...
... ..g-----ca-----gt-----t...
... ..g-----ca-----gt-----t...
... ..g-----ca-----gt-----t...
... ..g-----ca-----gt-----t...

2-IMGT <----- FR3-IMGT ----->
..ggaggttataaatataaggttagactccatgag...ggccgactccaccctctccagagaccattccaaagaccagctgtatttggaa
.. ..a-----ct-c-c-g-----t-----c-----c-c-
.. ..a-----ct-c-c-g-----t-----t-g-----c-c-
.. ..a-----ct-c-c-g-----t-----t-g-----c-c-
.. ..a-----ct-c-c-g-----t-----t-g-----c-c-
.. ..ca-----ct-c-c-g-----t-----t-----c-c-

          >----- CDR3-IMGT -----
atgaacagcctgacagctgaggaccagcctgtgtattactgtgcggagcagcttctttaaagcctatgactctctggggccagggaaac
-----g-----aga
-----g-----aga
-----g-----aga
-----g-----aga
-----g-----a

seq1
L06615 IGHV3-30*04
N83134 IGHV3-30*01
N77300 IGHV3-30*09
N77324 IGHV3-30*14
X92283 IGHV3-30-3*01

ctggtccacgctctctccagcctccaccaggggcccatcggctcttcccctggccacctctccaaagaccctctctggggccacc

```

**3. Alignment for J-GENE and allele identification**

Closest J-REGIONS

	Score	Identity
<u>X86355</u> IGHJ4*02	204	91,67% (44/48 nt)
<u>J00256</u> IGHJ4*01	195	89,58% (43/48 nt)
<u>N25625</u> IGHJ4*03	177	85,42% (41/48 nt)
<u>J00256</u> IGHJ5*01	174	82,35% (42/51 nt)
<u>X86355</u> IGHJ5*02	174	82,35% (42/51 nt)

**Alignment**

```

seq1
X86355 IGHJ4*02
J00256 IGHJ4*01
N25625 IGHJ4*03
J00256 IGHJ5*01
X86355 IGHJ5*02

gacagctttctttaagcctatgacctctggggccagggaccctgggtccacgctctctccagcctccaccaggggcccatcggctctccc
.....-ta--t-----a-----
.....-ta--t-----a-----
.....g-ta--t-----a-g-----
.....ac--tgg-tc--c-----a-----
.....ac--tgg-tc--cc-----

```

**FIGURE 6.** IMGT/V-QUEST “Detailed view” result page: “Alignment for V-GENE and allele identification” and “Alignment for J-GENE and allele identification.” If selected in the Search page, the alignments for the V- and J-GENE and allele identification are displayed with the alignment score, the percentage of identity for the five closest genes and alleles and the ratio number of identical nucleotides/number of aligned nucleotides. Dashes indicate identical nucleotides. Dots indicate gaps. FR-IMGT and CDR-IMGT are delimited according to the IMGT unique numbering (Lefranc 1997, 1999; Lefranc et al. 2003). The seq1 accession number is AB012909 from the IMGT/LIGM-DB database (Giudicelli et al. 2006).







**V-REGION mutation and AA change table**

FR-IMGT	CDR-IMGT	FR-IMGT	CDR-IMGT	FR-IMGT	CDR-IMGT
c1>g, Q1>E (+ + -)	g12>c, A20>P (- -)	c110>a, H62>T (+ -)	t07>c	c190>t	a210>g
g23>l, V5>G (+ -)		q110>a, H62>T (+ -)	q192>l, N64>Y (-)	l10>a, I57>K (- -)	g210>+
g70>a, A26>I (+ -)		g163>a, V50>G (- -)		c20>g, H2>K (- -)	a20>c
c71>l, A24>I (+ -)		l54>g, V50>G (- -)		c20>g, A60>V (+ +)	
				g211>a, V71>M (+ -)	
				c26>+, F70>L (+ -)	
				c30>+	
				c30>g, Q60>E (+ +)	
				g28>+, R61>T (- -)	

**V-REGION mutation and AA change statistics**

**Nucleotide (nt) mutations**

IMGT labels	V-REGION	FR-IMGT	CDR-IMGT	FR-IMGT	CDR-IMGT	FR-IMGT	CDR-IMGT
No of positions including IMGT gaps (nt)	317 (33)	75	26	61	30	117	5 (8)
No of nucleotides	291 (29)	75	24	61	24	114	5 (8)
No of identical nucleotides	273	71	23	47	22	105	5
No of mutations	28 (2)	4	1	4	2	9	0 (2)
Mutations	Silent	3 (8)	0	0	0	2	0 (2)
	Non-silent	5	4	1	4	7	0
Transitions	a>g	6 (1)	0	0	0	0	0 (1)
	g>a	3 (8)	1	0	1	0	1 (1)
	c>t	6	1	0	0	3	0
	t>c	2	0	0	0	1	0
Transversions	a>c	1 (2)	0	0	1	0	0 (1)
	c>a	1	0	0	1	0	0
	a>t	1	0	0	0	1	0
	t>a	1	0	0	0	1	0
	g>t	2	0	1	0	0	1
	t>g	3	1	0	0	0	2
	g>t	1	1	0	0	0	0
	t>g	1	0	0	1	0	0

**Amino acid (AA) changes**

IMGT labels	V-REGION	FR-IMGT	CDR-IMGT	FR-IMGT	CDR-IMGT	FR-IMGT	CDR-IMGT	
No of positions including IMGT gaps (AA)	301 (33)	26	12	17	10	39	1 (2)	
No of AA	37 (20)	25	9	17	9	36	1 (2)	
No of identical AA	34 (20)	22	7	15	7	32	1 (2)	
No of AA changes	13	3	1	2	1	6	0	
AA changes	Very similar (+ + +)	8	0	0	0	0	0	
	Similar (+ + -)	(+ + -)	2	1	0	0	1	0
		(+ - +)	3	2	0	0	0	1
	Dissimilar (- - +)	(- - +)	3	0	0	1	0	2
		(- + -)	6	0	0	0	0	0
		(- - -)	6	0	0	0	0	0
	Very dissimilar (- - -)	5	0	1	1	1	2	

**FIGURE 9.** IMGT/V-QUEST “Detailed view” result page: “V-REGION mutation and AA change table,” and “V-REGION mutation and AA change statistics.” The “V-REGION mutation and AA change table” lists the mutations (nt and AA) of the analyzed sequence compared with the closest V-REGION allele. They are described for the V-REGION and for each FR-IMGT and CDR-IMGT, with their nucleotide and codon position according to the IMGT unique numbering (Lefranc 1997, 1999; Lefranc et al. 2003) and for the AA changes according to the IMGT AA classes (Pommié et al. 2004). For example, c1>g, Q1>E (+ + -) means that the nucleotide mutation (c>g) at nt 1 leads to an AA change (Q>E) at codon 1 with the same hydrophathy (+) and volume (+) but with different physicochemical properties (-) classes (Pommié et al. 2004). It is the first time that such qualification of AA replacement is provided. This has led us to identify four types of AA changes: very similar (+++), similar (++-, +-+), dissimilar (- - +, - + -, + - -) and very dissimilar (- - -). The “V-REGION mutation and AA change statistics” comprises two tables, the first one for “Nucleotide (nt) mutations” described as above. For both tables, results are given for the V-REGION and per FR-IMGT and CDR-IMGT. Statistics are calculated up to the 3’ end of the V-REGION identified in the input sequence (this includes the 3’ last two identical nucleotides with the closest germline V-REGION). The numbers in parentheses, in the V-REGION and CDR3-IMGT columns, correspond to the statistics calculated up to the 3’ end of the closest germline V-REGION, and therefore may include nucleotide and amino acid differences due to the junction diversity. The accession number of the analyzed sequence is AB012909 from the IMGT/LIGM-DB database (Giudicelli et al. 2006).

11. V-REGION mutation hot spots (motifs and localizations in germline V-REGION)

(a)t <sub>a</sub> wa		t(a)t tw		(a)g g(c)t(a)t rgw		(a)t(a)g g(c)t wcy	
Motif	Localization	Motif	Localization	Motif	Localization	Motif	Localization
ta	105-106 (CDR1)	tt	82-83 (CDR1)	agct	5-11 (FR1)	agct	5-11 (FR1)
ta	109-110 (CDR1)	tt	88-89 (CDR1)	agta	103-106 (CDR1)	agcc	41-44 (FR1)
ta	114-115 (CDR1)	ta	105-106 (CDR1)	ggct	130-135 (FR2)	agcc	72-75 (FR1)
aa	142-143 (FR2)	ta	109-110 (CDR1)	ggca	139-142 (FR2)	agct	105-109 (CDR1)
ta	165-166 (FR2-CDR2)	ta	114-115 (CDR1)	ggct	146-149 (FR2)	tgct	111-114 (CDR1)
ta	167-168 (CDR2)	tt	164-165 (FR2)	ggca	159-162 (FR2)	tact	196-199 (FR3)
ta	172-173 (CDR2)	ta	167-168 (CDR2)	agta	167-190 (CDR2)	agcc	277-280 (FR3)
aa	186-187 (CDR2)	ta	172-173 (CDR2)	agct	205-208 (FR3)	agct	295-298 (FR3)
ta	189-190 (CDR2)	ta	189-190 (CDR2)	ggct	287-300 (FR3)	tact	307-310 (FR3)
ta	192-193 (CDR2)	ta	192-193 (CDR2)				
aa	194-195 (CDR2)	ta	196-197 (FR3)				
ta	196-197 (FR3)	ta	199-200 (FR3)				
ta	199-200 (FR3)	tt	225-227 (FR3)				
aa	214-215 (FR3)	tt	246-247 (FR3)				
aa	244-245 (FR3)	ta	262-263 (FR3)				
aa	250-251 (FR3)	ta	304-305 (FR3)				
aa	253-254 (FR3)	tt	306-307 (FR3)				
ta	262-263 (FR3)						
aa	269-270 (FR3)						
aa	274-275 (FR3)						
ta	304-305 (FR3)						
ta	307-308 (FR3)						

**FIGURE 10.** IMGT/V-QUEST “Detailed view” result page: “V-REGION mutation hot spots (motifs and localization in germline V-REGION).” This table provides the hot spot motifs identified in the closest germline V-REGION and their localization. The hot spot patterns are (a)t<sub>a</sub> and (a)g|g(c)t(a)t, and their complementary reverse motifs t(a)t and (a)t(a)g|g(c)t. The accession number of the analyzed sequence is AB012909 from the IMGT/LIGM-DB database (Giudicelli et al. 2006).

11. "seq1" V-REGION and V-D-J-REGION

V-REGION nucleotide sequence in FASTA format

```
>seq1
ggggtgagcgtgttgggctctggggg...ggcgtggtcagcctgggggtccctgagc
ctctctctgtatagcctctggcttcccttc...agtagctatcctatgac
tgggtcggcaggctcagggcaggggtggggtgggtggcagctatctatgac...
...ggagctatcaatctaggtagcctccatgag...ggcagctcaccctctcagc
gcaatctcagagacagcgtgtcttggaaatgcaagcctgacagctggggacgggt
gtgatctctgtggag
```

[Analyze this sequence with IMGT/PhyloGene](#)

V-REGION amino acid sequence in FASTA format

```
>seq1
EVQLLESG...GVVQFQRLSLSCLIASDTF...STYPTWVQAIFKLEWFAISIT...
...GSTEYVDSK...GLTISSRHSHNTLLEHSLTAEDTAVTYCA
```

V-REGION amino acid sequence on one line

```
>seq1
EVQLLESG...GVVQFQRLSLSCLIASDTF...STYPTWVQAIFKLEWFAISIT...GSTEYVDSK...GLTISSRHSHNTLLEHSLTAEDTAVTYCA
```

V-D-J-REGION nucleotide sequence in FASTA format

```
>seq1
ggggtgagcgtgttgggctctggggg...ggcgtggtcagcctgggggtccctgagc
ctctctctgtatagcctctggcttcccttc...agtagctatcctatgac
tgggtcggcaggctcagggcaggggtggggtgggtggcagctatctatgac...
...ggagctatcaatctaggtagcctccatgag...ggcagctcaccctctcagc
gcaatctcagagacagcgtgtcttggaaatgcaagcctgacagctggggacgggt
gtgatctctgtggagcagcgtctcttcaagcctatgactcttggggcagggac
ctggtaccgctctcctag
```

V-D-J-REGION amino acid sequence in FASTA format

```
>seq1
EVQLLESG...GVVQFQRLSLSCLIASDTF...STYPTWVQAIFKLEWFAISIT...
...GSTEYVDSK...GLTISSRHSHNTLLEHSLTAEDTAVTYCASTAFFHATDFWQGT
LVTVSS
```

V-D-J-REGION amino acid sequence on one line

```
EVQLLESG...GVVQFQRLSLSCLIASDTF...STYPTWVQAIFKLEWFAISIT...GSTEYVDSK...GLTISSRHSHNTLLEHSLTAEDTAVTYCASTAFFHATDFWQGTLVTVSS
```

**FIGURE 11.** IMGT/V-QUEST “Detailed view” result page: sequences of V-REGION and V-J- or V-D-J-REGION. This section provides nucleotide and amino acid sequences with gaps according to the IMGT unique numbering (Lefranc 1997, 1999; Lefranc et al. 2003) and includes the V-REGION nucleotide sequence in FASTA format with access to the IMGT/PhyloGene tool (Elemento and Lefranc 2003), “V-REGION amino acid on one line,” V-J- or V-D-J-REGION nucleotide and amino acid sequences in FASTA format, V-J- or V-D-J-REGION AA sequence on one line.

### 13. Annotation by IMGT/Automat

Label	Location/Qualifiers
<u>V-D-J-REGION</u>	1..355 /CDR_length="[8.8.11]" /FR_length="[25.17.38.11]" /nucleotide sequence ggagtgcaagctgttggagctctgggggaggcgtgggtccagcctgggaggtccctgagactc tcctgtatagcctctggattcaocttcagtagctatcctatgacctgggtccggccagggt ccaggccaggggcctggagtggtggcaggtatcctatgacgggaagttataaatataag gtagactccatgagggccgactcaccatctccagagacaattccaagacacgctgtat ttggaaatgacagcctgacagctgaggacaggcgtgtgtattactgtgagaggacagct ttctttaagcctatgactctctggggccagggaacctgggtccagctctcctcag /translation EVQLLESGGGVVQPGSRRLRSCIASGFTFSSYFHTWVRQAPGKLEWVASISYDOSTYK VDSNKGRLTISRINSKNTLYLENSLTAEDTAVYYCARAFFNATDFWGQGLTVVSS
<u>V-REGION</u>	1..293 /allele="IGHV3-30*04" /gene="IGHV3-30" /nucleotide sequence ggagtgcaagctgttggagctctgggggaggcgtgggtccagcctgggaggtccctgagactc tcctgtatagcctctggattcaocttcagtagctatcctatgacctgggtccggccagggt ccaggccaggggcctggagtggtggcaggtatcctatgacgggaagttataaatataag gtagactccatgagggccgactcaccatctccagagacaattccaagacacgctgtat ttggaaatgacagcctgacagctgaggacaggcgtgtgtattactgtgagag /translation EVQLLESGGGVVQPGSRRLRSCIASGFTFSSYFHTWVRQAPGKLEWVASISYDOSTYK VDSNKGRLTISRINSKNTLYLENSLTAEDTAVYYCA
<u>FR1-IGT</u>	1..75 /AA_IGT="AA 1 to 26, AA 10 is missing" /nucleotide sequence ggagtgcaagctgttggagctctgggggaggcgtgggtccagcctgggaggtccctgagactc tcctgtatagcctct /translation EVQLLESGGGVVQPGSRRLRSCIAS
<u>1st-CYS</u>	64..66 /nucleotide sequence tgt /translation C
<u>CDR1-IGT</u>	76..99 /AA_IGT="AA 27 to 30, AA 31, 32, 33, 34 are missing" /nucleotide sequence ggattcaocttcagtagctatcct /translation GFTFSSYF
<u>FR2-IGT</u>	100..150 /AA_IGT="AA 39 to 55" /nucleotide sequence atgacctgggtccgacaggctccaggccaggggcgtggagtggtggcaggt /translation ETWVRQAPGKLEWVAS
<u>CONSERVED-TFP</u>	104..108 /nucleotide sequence tgg /translation W
<u>CDR2-IGT</u>	151..174 /AA_IGT="AA 56 to 65, AA 60, 61 are missing" /nucleotide sequence atctcatgacgggaagttataaa /translation ISYDOSTK

**FIGURE 12.** IMGT/V-QUEST “Detailed view” result page: “Annotation by IMGT/Automat”: the results of the analysis of the input sequence by IMGT/Automat (Giudicelli et al. 2003, 2005b) provide a full automatic annotation for the V-J-REGION or V-D-J-REGION using IMGT standardized labels.

### IMGT/V-QUEST Excel File

21. Select the option “C. Excel file.”

22. Select, in the light orange box, the displays of the results to be shown.

*There are 11 sheets proposed for the result display. The “Summary” and “Parameters” sheets are always provided by default.*

- Summary
- IMGT-gapped-nt-sequences



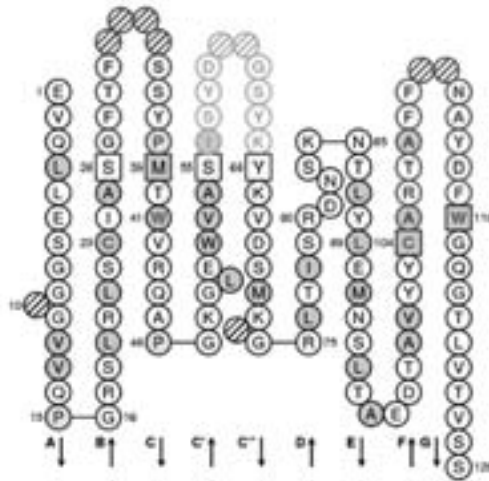
#### 14. IMGT Collier de Perles for seq1

IMGT/V-QUEST identified region: V-D-J-REGION

CDR-IMGT lengths: [8.8.11]

FR-IMGT lengths: [26.17.38.11]

CDR-IMGT and FR-IMGT lengths are based on the [IMGT unique numbering for V-DOMAIN](#)



**FIGURE 13.** IMGT/V-QUEST “Detailed view” result page: IMGT Collier de Perles. This section recalls the identified region (here, V-D-J-REGION) and the CDR-IMGT lengths and FR-IMGT lengths. Depending on the choice selected in option 14 of the IMGT/V-QUEST Search page (see Fig. 2), the “IMGT Collier de Perles” (Ruiz and Lefranc 2002; Kaas et al. 2004, 2007; Kaas and Lefranc 2007) can be displayed either as a “link to IMGT/Collier de Perles tool” **IMGT/Collier de Perles: IMGT Standardized Representation of Domains (IG, TR, and IgSF Variable and Constant Domains, MH and MhSF Groove Domains)** (Ehrenmann et al. 2011) or as a direct “IMGT Collier de Perles (for a nb of sequences <5)” representation integrated into IMGT/V-QUEST results. The IMGT Collier de Perles shown was obtained by selecting this last option. Positions in blue mean that the amino acids of the user sequence at these positions is hydrophobic (hydropathy index with positive value) or is a tryptophan (W), as in 50% or more of analyzed V domains. Positions with red and bold letters indicate the five conserved positions of a V domain: 1st-CYS 23, CONSERVED-TRP 41, hydrophobic 89, 2nd-CYS 104, and J-TRP 118. Anchor positions are in squares. Hatched positions correspond to gaps according to the IMGT unique numbering for V domain (Lefranc 1997, 1999; Lefranc et al. 2003). Prolines are shown in yellow. Arrows indicate the  $\beta$  strands and their direction. The CDR-IMGT lengths of this domain are (8.8.11). The CDR-IMGT are colored as follows: CDR1-IMGT (red), CDR2-IMGT (orange), and CDR3-IMGT (purple) for the IGH (this figure), TRB and TRD V-D-J-REGION, and CDR1-IMGT (blue), CDR2-IMGT (green), and CDR3-IMGT (greenblue) (data not shown) for the IKG, IGL, TRA, and TRG V-J-REGION.

- *nt-sequences*
- *IMGT-gapped-AA-sequences*
- *AA-sequences*
- *Junction*
- *V-REGION-mutation-and-AA-change-table*
- *V-REGION-nt-mutation-statistics*
- *V-REGION-AA-change-statistics*
- *V-REGION-mutation-hot spot*
- *Parameters*

23. Go to “Advanced parameters” at the bottom of the search page and modify the parameters according to the particularities of the sequences, if needed (for details, see Step 11).

24. Click on “Start” to launch the analysis.

*A small window will appear on the screen, giving the choice to “open” or “save” the Excel file.*

## B. Synthesis for the IMGT/V-QUEST analysed sequences

Number of analysed sequences: 7

Sequences compared with the [human\\_IG.fasta](#) from the [IMGT reference directory](#)

Summary table:

Sequence ID	V-GENE and allele	Functionality	V-REGION score	V-REGION Identity % (nt)	J-GENE and allele	D-GENE and allele	D-REGION reading frame	CDR1-IMGT lengths	AA JUNCTION	JUNCTION frame
seq1	IGHV1-69*01	Productive	1273	93.75% (270/288 nt)	IGHJ4*01 or IGHJ4*02 or IGHJ4*03	IGHD3-22*01	2	[8.8.12]	CAKQYGSSTYYAYW	in-frame
seq2	IGHV3-30*03, or IGHV3-30*18	Productive	1417	99.31% (285/288 nt)	IGHJ6*02	IGHD3-16*02	2	[8.8.23]	CAKDLDDYVWGSYRHSRYTYGMDFW	in-frame
seq3	IGHV3-30*04	Productive	1255	93.06% (269/288 nt)	IGHJ4*02	IGHD5-6*01	1	[8.8.11]	CARTAFFNAYDFW	in-frame
seq4	IGHV3-30*04	Productive	1435	100.00% (288/288 nt)	IGHJ4*02	IGHD6-19*01	2	[8.8.15]	CARDRSIAVAQYVYDFYW	in-frame
seq5	IGHV3-30*04	Productive	1409	98.96% (285/288 nt)	IGHJ4*02	IGHD6-13*01	2	[8.8.17]	CARDSSYPPGIAAAGVYVW	in-frame
seq6	IGHV3-30*04	Productive	1425	99.65% (287/288 nt)	IGHJ1*01	IGHD3-10*01	2	[8.8.16]	CARGRTKGSQRPQYFQHW	in-frame
seq7	IGHV3-9*01	Unproductive (stop codons)	1417	99.31% (285/288 nt)	IGHJ4*02	IGHD3-10*01	2	[8.8.17]	CAKDIQGGKYSGSGSGLD*W	in-frame

Results of IMGT/JunctionAnalysis for : [IGH junctions](#)

Alignment with the closest alleles:

The analysed sequences are aligned with the closest allele (with number of aligned sequences in parentheses):

[IGHV3-69\\*01 \(1\)](#) [IGHV3-30\\*03 \(1\)](#) [IGHV3-30\\*04 \(6\)](#) [IGHV3-9\\*01 \(1\)](#)

**FIGURE 14.** IMGT/V-QUEST “Synthesis view” result page: *top* of the page. The number of analyzed sequences is indicated. The Summary table displays one row for each input sequence, including: (1) the name of the sequence (Sequence ID), (2) the name of the closest V-GENE and allele (a note may appear with the V-GENE and allele name when the V-REGION score is very low (<200), and/or the percentage of identity is <85% and/or, compared with that of V, when input sequence has different CDR1-IMGT and/or CDR2-IMGT lengths: the alignment for this sequence has to be checked in “A. Detailed view,” using the advanced parameters “Search for insertions and deletions in V-REGION”), (3) the functionality of the sequence (when found, the presence of stop codons is indicated), (4) the V-REGION score, (5) the V-REGION percentage of identity and ratio number of identical nucleotides (nt)/number of aligned nt, (6) the name of the closest J-GENE and allele (notes may appear with the J-GENE and allele name to indicate that other possibilities exist for the choice of the J-GENE and allele name), and (7) provided according to the IMGT/JunctionAnalysis results, the D-GENE and allele name, the D reading frame, the CDR-IMGT lengths, the AA JUNCTION, and the JUNCTION frame. In the absence of results of IMGT/JunctionAnalysis, only the AA JUNCTION defined by IMGT/V-QUEST is displayed. The seq1, seq2, seq3, seq4, seq5, seq6, and seq7 correspond to the DQ100777, AB021524, AB012909, AB021511, AB021516, AB021514, and AB021539 accession numbers, respectively, from the IMGT/LIGM-DB database (Giudicelli et al. 2006).

25. To see the results, click on “open.”

*The Excel file comprises 11 sheets. These can be viewed in the supplementary material (Results\_Excel.doc).*

## DISCUSSION

IMGT/V-QUEST (<http://www.imgt.org>) provides a highly detailed and standardized analysis of the IG and TR rearranged sequences: it describes the main characteristics of user sequences according to the axioms and concepts of IMGT-ONTOLOGY (Giudicelli et al. 1999; Duroux et al. 2008). IMGT/V-QUEST is constantly updated based on the results of biological and clinical research. Improvements can be followed through the program version number and the reference directory release number indicated in the search pages and the list of the corresponding changes found in the corresponding documentations. New alleles annotated in IMGT are integrated in the IMGT reference directory as soon as they are confirmed experimentally and are publicly available in the generalist databases. IMGT/HighV-QUEST (Alamyar et al. 2010, 2011), the high-throughput version of IMGT/V-QUEST for Next Generation Sequencing (NGS) that analyzes up to 150,000 sequences per run, uses the IMGT/V-QUEST program and standards, and provides results with the same degree of resolution and high quality (results











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# IMGT/JunctionAnalysis: IMGT Standardized Analysis of the V-J and V-D-J Junctions of the Rearranged Immunoglobulins (IG) and T Cell Receptors (TR)

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## INTRODUCTION

IMGT/JunctionAnalysis is the online IMGT tool for the detailed and standardized analysis of the junctions between the variable (V), diversity (D), and joining (J) genes (V-J and V-D-J junctions) of the rearranged immunoglobulin (IG) or antibody and T cell receptor (TR) variable domains. The V-(D)-J junctions comprise the rearranged CDR3-IMGT and its anchors 2nd-CYS 104 and J-PHE or J-TRP 118. The diversity of the junctions that determines the antigen receptor specificity results from complex molecular mechanisms that occur at the DNA level during the IG and TR synthesis and create combinatorial diversity, N-diversity and, for IG, somatic hypermutations. The annotation of V-J or V-D-J junctions in rearranged IG and TR sequences represents a huge challenge due to its uniqueness and complexity. IMGT/JunctionAnalysis has been a major breakthrough by providing, for the first time, a very detailed and accurate analysis of the junctions. The tool, whose use is described here, identifies the D genes in the IGH, TRB, and TRD junctions, the trimmed nucleotides (nt) at the end of the genes which recombine, and the palindromic P regions in the absence of gene trimming. It delimits the N regions that result from the N-diversity, calculates the ratio of G+C nucleotides in the N regions, and evaluates the number of somatic hypermutations for each gene within the junction.

## RELATED INFORMATION

IMGT/JunctionAnalysis (Lefranc 2004; Yousfi Monod et al. 2004; Giudicelli and Lefranc 2005, 2008) is part of IMGT, the international ImMunoGeneTics information system, <http://www.imgt.org> (Lefranc et al. 2009). A detailed description of IMGT is provided in **IMGT, the International ImMunoGeneTics Information System** (Lefranc 2011a). Standardization and IMGT Scientific chart rules are based on the IMGT-ONTOLOGY concepts of identification, classification, description (Giudicelli and Lefranc 1999; Lefranc et al. 2004, 2005a, 2008) and numerotation (Lefranc 1997, 1999; Lefranc et al. 2003, 2005b, c) generated from the axioms of the Formal IMGT-ONTOLOGY or IMGT-Kaleidoscope (Duroux et al. 2008). The concepts of identification led to the IMGT standardized keywords (**From IMGT-ONTOLOGY IDENTIFICATION Axiom to IMGT Standardized Keywords: For Immunoglobulins (IG), T Cell Receptors (TR), and Conventional Genes**) (Lefranc 2011b), the concepts of description to the IMGT standardized labels (**From IMGT-ONTOLOGY DESCRIPTION Axiom to IMGT Standardized Labels: For Immunoglobulin (IG) and T Cell Receptor (TR) Sequences and Structures**) (Lefranc 2011c), the concepts of classification to the IMGT standardized gene and allele names (**From IMGT-ONTOLOGY CLASSIFICATION Axiom to IMGT Standardized Gene and Allele Nomenclature: For Immunoglobulins (IG) and T Cell Receptors (TR)**) (Lefranc 2011d), and the concepts of numerotation to the IMGT unique numbering (**IMGT Unique Numbering for the Variable (V), Constant (C), and Groove (G) Domains of IG, TR, MH, IgSF, and MhSF**) (Lefranc 2011e), and IMGT Colliers de Perles (**IMGT Collier de Perles for the Variable (V), Constant (C), and Groove (G) Domains of IG, TR, MH, IgSF, and MhSF**) (Lefranc 2011f).

In addition, protocols are available for **IMGT/V-QUEST: IMGT Standardized Analysis of the Immunoglobulin (IG) and T Cell Receptor (TR) Nucleotide Sequences** (Giudicelli et al. 2011),

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Cite as: Cold Spring Harb Protoc; 2011; doi:10.1101/pdb.prot5634

IMGT/Collier de Perles: IMGT Standardized Representation of Domains (IG, TR, and IgSF Variable and Constant Domains, MH and MhSF Groove Domains) (Ehrenmann et al. 2011), IMGT/Domain-GapAlign: IMGT Standardized Analysis of Amino Acid Sequences of Variable, Constant, and Groove Domains (IG, TR, MH, IgSF, MhSF) (Ehrenmann and Lefranc 2011a), and IMGT/3Dstructure-DB: Querying the IMGT Database for 3D Structures in Immunology and Immunoinformatics (IG or Antibodies, TR, MH, RPI, and FPIA) (Ehrenmann and Lefranc 2011b).

## MATERIALS

It is essential that you consult the appropriate Material Safety Data Sheets and your institution's Environmental Health and Safety Office for proper handling of equipment and hazardous materials used in this protocol.

### Equipment

Computer (Internet-connected)

## METHOD

1. Using any modern web browser, access the IMGT Home page, <http://www.imgt.org> and click, in the "IMGT tools" section, on the link to IMGT/JunctionAnalysis.

*This gives access to the IMGT/JunctionAnalysis Welcome page.*

### IMGT/JunctionAnalysis Welcome Page

2. In the IMGT/JunctionAnalysis Welcome page (Fig. 1), locate at the top of the page:

"Species" (drop-down list)

"Locus" (radio buttons): IGH, IGK, IGL, TRA, TRB, TRG, TRD

*IMGT/JunctionAnalysis provides in a single run a standardized analysis of an unlimited number of IG and TR V-J and V-D-J junctions from the same locus and the same species. Analysis of IG and TR junctions of mice and humans can be performed exhaustively. Analysis of junctions of other species (rat, rabbit, trout) becomes progressively more available as genomic sequences are annotated in IMGT.*

3. Select the species and the locus.

4. Locate the sequence submission section.

*That section comprises:*

*Two windows to either type (or copy/paste) your sequence(s) or upload a file, in FASTA format.*

*The "Start" and "Clear the form" buttons.*

*Because the junctions include only a short part of the 3' end of the V (3'V-REGION) and of the 5' end of the J (5'J-REGION) (Bleakley et al. 2006, 2008), the tool cannot identify the V and J genes and alleles and, therefore, the corresponding gene and allele names must be provided and a specific format is required (detailed in Step 5).*

5. Check the format of your junction nucleotide sequences. Keep the following points in mind:
  - i. The required format is the FASTA format. Each JUNCTION nucleotide sequence must be preceded by the following information:
    - Identifier ("input"), with a 10-character maximum length. This identifier can be a sequence name, an accession number, a clone name, etc.
    - The name of the V-GENE and allele according to the IMGT gene name nomenclature (Lefranc 2000a,b, 2007, 2008; Lefranc and Lefranc 2001a,b; Giudicelli et al. 2005a).
    - The name of the J-GENE and allele according to the IMGT gene name nomenclature (Lefranc 2000a,b, 2007, 2008; Lefranc and Lefranc 2001a,b; Giudicelli et al. 2005a).

# WELCOME ! to IMGT/JunctionAnalysis

THE  
INTERNATIONAL  
IMMUNOGENETICS  
INFORMATION SYSTEM®



Citing IMGT/JunctionAnalysis: Youdi Monod, M. et al., *Bioinformatics*, 20, (379-385 (2004)). PMID: 15262823 [www](#)

IMGT/JunctionAnalysis programme version: 2.0.1 (21 March 2011) - IMGT/JunctionAnalysis reference directory release: 201102-1 (7 March 2011)

## Analyse the JUNCTION of your IG and TR nucleotide sequences

Sequence sets to test IMGT/JunctionAnalysis are available [here](#)

### Sequence submission

Species:

Locus:  IGH  IGK  IGL  
 TRA  TRB  TRG  TRD

Type (or copy/paste) your JUNCTION nucleotide sequence(s) into the box below (click [here](#) for required format):

```
>seq1,IGHV1-3*01,IGHJ4*02
tgtgagagagactctatcggtagcttgcctactcgg
>seq2,IGHV3-10*04,IGHJ4*02
tgtgagagagagcgtttcttcaagcctcagctctcgg
>seq3,IGHV3-21*01,IGHJ5*02
tgtgagagaggtacagtaggctcctcggtagccctcgg
>seq4,IGHV3-23*01,IGHJ5*02
```

Or give the path access to a local file containing your sequences:

Click here for an example of IMGT/JunctionAnalysis results.

### Display results

List of all eligible D-GENE:  Yes  No

Colored IMGT AA classes and histogram:  Yes  No

Output order:  Same order as input  
 CDR3-MGT length decreasing order  
 CDR3-MGT length increasing order

### Advanced parameters

5' and 3' ends of the JUNCTION:  Default  
 May start and/or end with any codon

Number of D-GENEs for IGH, TRB and TRD JUNCTION:

Number of accepted mutations:  in 3V-REGION  
 in D-REGION  
 in 5J-REGION

Delimitation of 3V-REGION, D-REGION and 5J-REGION:  Default  
 Stop trimming with the first encountered identical nucleotide

D-GENE choice (if several have the same score):  The less mutated one  
 The longest one  
 The one more upstream in the locus

FIGURE 1. IMGT/JunctionAnalysis Welcome page.

- ii. There is no limitation in the number of sequences to be analyzed in a single search, but a new line should be started for each sequence:

>Input1, V-GENE and allele name, J-GENE and allele name  
nucleotide sequence (in uppercase or lowercase)

>Input2, V-GENE and allele name, J-GENE and allele name  
nucleotide sequence (in uppercase or lowercase)

For instance,

>Seq1,IGHV7-4-1\*02,IGHJ4\*02



```
tgtgcgagagaagatagcaatggctacaaaatattgactactgg
>Seq2, IGHV1-69*06, IGHJ5*02
tgtgcgagagggggggtaaggtcgaattttggagtggttcatgggtactgggtcagaccctgg
```

- iii. If the V-GENE allele or J-GENE allele is unknown, the IMGT/JunctionAnalysis tool accepts a “?” character instead of the allele number (ex: IGHV1-2\*?) and will run the search against the allele \*01 by default.
  - iv. If there are several proposed V-GENE and/or J-GENE, the different V-GENE and allele names, and/or J-GENE and allele names have to be separated by the “/” character (ex: IGHV1-2\*01/IGHV1-3\*?/IGHV1-18\*02, IGHJ1\*01/IGHJ2\*01). The IMGT/JunctionAnalysis tool will run the search against the first V-GENE and allele and J-GENE and allele listed.
6. Select either “Type (or copy/paste) your JUNCTION nucleotide sequence(s) into the box below” and type (or copy and paste) the junction nucleotide sequences to be analyzed in the text area, or “Or give the path access to a local file containing your sequences” and upload a text file by using the “Browse” button to select the file.
  7. For more information, locate “Click here for an [example](#) of IMGT/JunctionAnalysis” (option for first time users or for educational purposes):
    - Click on “[example](#)” to see the format for submission.
    - Select the radiobutton to automatically launch the analysis for the set of sequences indicated in the documentation.

## Display Results

8. Locate the “Display results” section (Fig. 1). Perform Steps 9–11 for choosing options of the display, or if not needed, proceed to Step 12.
9. Choose, if preferred, not to display the “List of all eligible D-GENE.”

*This option allows one to visualize all D genes that match a junction and to compare their score. It is displayed by default when only one junction is analyzed in the run, but can be eliminated.*
10. Choose, if preferred, not to display the “Colored IMGT AA classes and histogram.”

*A display of the “Colored IMGT AA classes and histogram,” with colors of the AA according to the 11 IMGT physico-chemical AA classes (Pommié et al. 2004) (IMGT Aide-mémoire>Amino acids, <http://www.imgt.org>), is provided by default but can be eliminated.*
11. Choose, if preferred, an “Output order” in “CDR3-IMGT length decreasing order” or in “CDR3-IMGT length increasing order.”

*The results in “JUNCTION alignments with translation and IMGT AA classes” may be displayed in “Same order as input” (default), “CDR3-IMGT length decreasing order,” or “CDR3-IMGT length increasing order.”*

## Advanced Parameters

12. Locate the “Advanced parameters” section (Fig. 1). Perform Steps 13–17 for modifying parameters, or if not needed, proceed directly to Step 18.

*Advanced parameters are usually not needed with classical junctions. However, going through Steps 13–17 (even without modifying the parameters) will allow you to be aware of all the possibilities offered by the tool.*
13. Check “5’ and 3’ ends of the JUNCTION” of your sequences and, if needed, select “May start and/or end with any codon.”

*“Default” means that, as normally expected, your junction nucleotide sequences start in 5’ with 2nd-CYS 104 codon (V-REGION cysteine [“tgt” or “tgc”] codon) and end in 3’ with J-PHE or J-TRP 118 codon (J-REGION phenylalanine [“ttt” or “ttc”] or tryptophan [“tgg”]) codon). The conserved amino acids (AA) 2nd-CYS and J-PHE or J-TRP at positions 104 and 118, according to the IMGT unique numbering for V-DOMAIN (Lefranc 1997, 1999; Lefranc et al. 2003), are indeed the anchors of the CDR3-IMGT that delimit the JUNCTION.*

*If one and/or the other of these anchor codon(s) is(are) mutated in some of your sequences, you can modify this parameter by selecting the other alternative “May start and/or end with any codon.”*

14. Modify, if needed, the “Number of D-GENE (for IGH, TRB, and TRD JUNCTION)” to be searched in the drop-down list.  
*Three loci IGH, TRB, and TRD loci have D genes in their V-D-J junction. “Default” means that the maximum number of D genes to be searched is one for IGH, one for TRB, and three for TRD.*  
*You can modify the number of D genes to be searched in the junctions from 0 to 3 by selecting a number in the drop-down list.*
15. Modify, if needed, the “Number of accepted mutations,” in the corresponding 3'V-REGION, D-REGION, and 5'J-REGION drop-down lists.  
*The maximum number of accepted mutations in the 3' V-REGION, D-REGION, and 5' J-REGION depends on the locus. By default, no mutation is accepted for the TR junctions. For IGH, two, four, and two mutations are accepted in the 3' V-REGION, D-REGION, and 5' J-REGION, respectively. For the IGK and IGL loci, the maximum number of accepted mutations is seven in the 3' V-REGION and seven in the 5' J-REGION.*  
*You can modify these numbers from 0 to 10 in the corresponding 3' V-REGION, D-REGION, and 5' J-REGION drop-down lists.*
16. Select, if needed, for the “Delimitation of 3'V-REGION, D-REGION and 5'J-REGION,” “Stop trimming with the first encountered identical nucleotide.”  
*Rules have been defined to identify the trimmed (deleted) nucleotides at the 3' end of the 3' V-REGION, 5' and 3' ends of the D-REGION(s), and 5' end of the 5' J-REGION (Bleakley et al. 2006, 2008).*  
*“Default” is “using patterns.” This indicates that the tool identifies as trimmed the nucleotides that match the patterns “m,” “m-” or “mm--” (at the 3' end of the 3' V-REGION and D-REGION), and the patterns “m,” “-m,” or “--mm” (at the 5' end of the 5' J-REGION and D-REGION) (where “m” indicates a mutation and “-” indicates an identical nucleotide by comparison with the corresponding closest germline alleles). The alternative is “Stop trimming with the first encountered identical nucleotide.”*
17. Select, if needed, for the “D-GENE choice (if several have the same score),” “The longest one,” or “The one more upstream in the locus.”  
*This criterion is used to discriminate between D regions with identical highest scores. “Default” is “The less mutated one.” The other alternatives are “The longest one” or “The one more upstream in the locus.”*
18. Click on the “Start” button to launch the analysis.  
*This leads to the display of the IMGT/JunctionAnalysis Results page (Fig. 2 and Fig. 3).  
 See Discussion for further details.*

## DISCUSSION

### The IMGT/JunctionAnalysis Results Page

The IMGT/JunctionAnalysis Results page provides the following:

1. A brief summary at the top of the page (Fig. 2).

It recalls the locus and species (as selected in Step 3).

It provides, for information, a link to the Locus representation in the IMGT Repertoire.

It recalls the values of the parameters used by the tool for the analysis:

- “Maximum number of accepted mutations” (see Step 15)
- “Deletion limits:” e.g., “using patterns” (default) (see Step 16)
- “Best D-GENE choice for a same score:” e.g., “less mutations” (default) (see Step 17)

2. “Analysis of the JUNCTION” (Fig. 2).

The “Analysis of the JUNCTION” provides the results of the analysis of the junctions at the nucleotide level. The junctions are displayed according to the order of the sequence submissions with the names of the input sequences and names of the V and J genes and alleles as provided by the user (in IMGT/JunctionAnalysis integrated in IMGT/V-QUEST (Lefranc 2004; Giudicelli and Lefranc 2005, 2008; Brochet et al. 2008) (**IMGT/V-QUEST: IMGT Standardized Analysis of the Immunoglobulin (IG) and T Cell Receptor (TR) Nucleotide Sequences** [Giudicelli et al. 2011], that information is from

## Analyse the JUNCTION of your IG and TR nucleotide sequences

### Results

Level: IGH  
Species: Homo sapiens  
IMGT Repertoire file: [IGL01.1000000000](#)

Maximum number of accepted mutations:  
V-REGION: 2, D-REGION: 4, J-REGION: 2  
Deletion limits:  
using patterns  
Best D-GENE choice for a same score:  
less mutations

### Analysis of the JUNCTION

Click on mutated (underlined) nucleotide to see the original one

Input	V name	3'-V-REGION	P N1	D-REGION	D name	5'-J-REGION	D name	5'-J-REGION	Result	Score	Dist	Ngc
#1	seq1	<u>10001-2153</u>	tgctgcaagcaga	ant	...ctttggtg	...ctttggtg	10001-2153	10001-2153	0	0	0	0.73
#2	seq2	<u>10001-2070</u>	tgctgcaagc...	g	...atagctgcttca	...atagctgcttca	10001-2070	10001-2070	0	2	0	0.73
#3	seq3	<u>10001-2170</u>	tgctgcaagc...	g	...gttagctggtt	...gttagctggtt	10001-2170	10001-2170	0	2	0	0.71
#4	seq4	<u>10001-2170</u>	tgctgcaagc...	tcctctctctct	...gatctgctggtt	...gatctgctggtt	10001-2170	10001-2170	0	1	1	0.710
#5	seq5	<u>10001-2070</u>	tgctgcaagc...	agggagaggt	...tgggggtt	...tgggggtt	10001-2070	10001-2070	1	1	0	0.710
#6	seq6	<u>10001-2170</u>	tgctgcaagc...	g	...cttagctgctggtt	...cttagctgctggtt	10001-2170	10001-2170	0	0	0	0.71
#7	seq7	<u>10001-2170</u>	tgctgcaagc...	g	...cttagctgctggtt	...cttagctgctggtt	10001-2170	10001-2170	0	2	0	0.71
#8	seq8	<u>10001-1870</u>	tgctgcaagc...	g	...cttagctgctggtt	...cttagctgctggtt	10001-1870	10001-1870	0	1	0	0.71
#9	seq9	<u>10001-2170</u>	tgctgcaagc...	g	...cttagctgctggtt	...cttagctgctggtt	10001-2170	10001-2170	0	0	0	0.71
#10	seq10	<u>10001-2070</u>	tgctgcaagc...	g	...cttagctgctggtt	...cttagctgctggtt	10001-2070	10001-2070	1	0	0	0.71
#11	seq11	<u>10001-2170</u>	tgctgcaagc...	agggagaggt	...gttagctgctggtt	...gttagctgctggtt	10001-2170	10001-2170	0	3	0	0.710

**FIGURE 2.** IMGT/JunctionAnalysis Results page: top of the page and “Analysis of the JUNCTION.” Nucleotides of each region identified in a JUNCTION are displayed: 3’V-REGION, P regions (P), N regions (N1 and N2), D-REGION, and 5’J-REGION. Dots represent trimmed nucleotides. D name (IGH D-GENE and allele) is identified by IMGT/JunctionAnalysis. Underlined nucleotides are mutated compared with the germline region (the original nucleotide can be seen by clicking on the mutated one). Sequences seq1–seq11 correspond to the junction of sequences with accession numbers AJ399822, AB012909, AB027443, AF015123, AB021512, AB021521, AB063654, AB063683, AB021537, AB027433, AB027433, respectively, from the IMGT/LIGM-DB database (Giudicelli et al. 2006).

IMGT/V-QUEST analysis). Nucleotides of each region identified in a JUNCTION are displayed. Dots in 3’V-REGION, D-REGION, and 5’J-REGION indicate nucleotides trimmed in the rearranged sequence, by comparison to the corresponding germline 3’V-REGION, D-REGION, and 5’J-REGION.

Results provided by IMGT/JunctionAnalysis comprise:

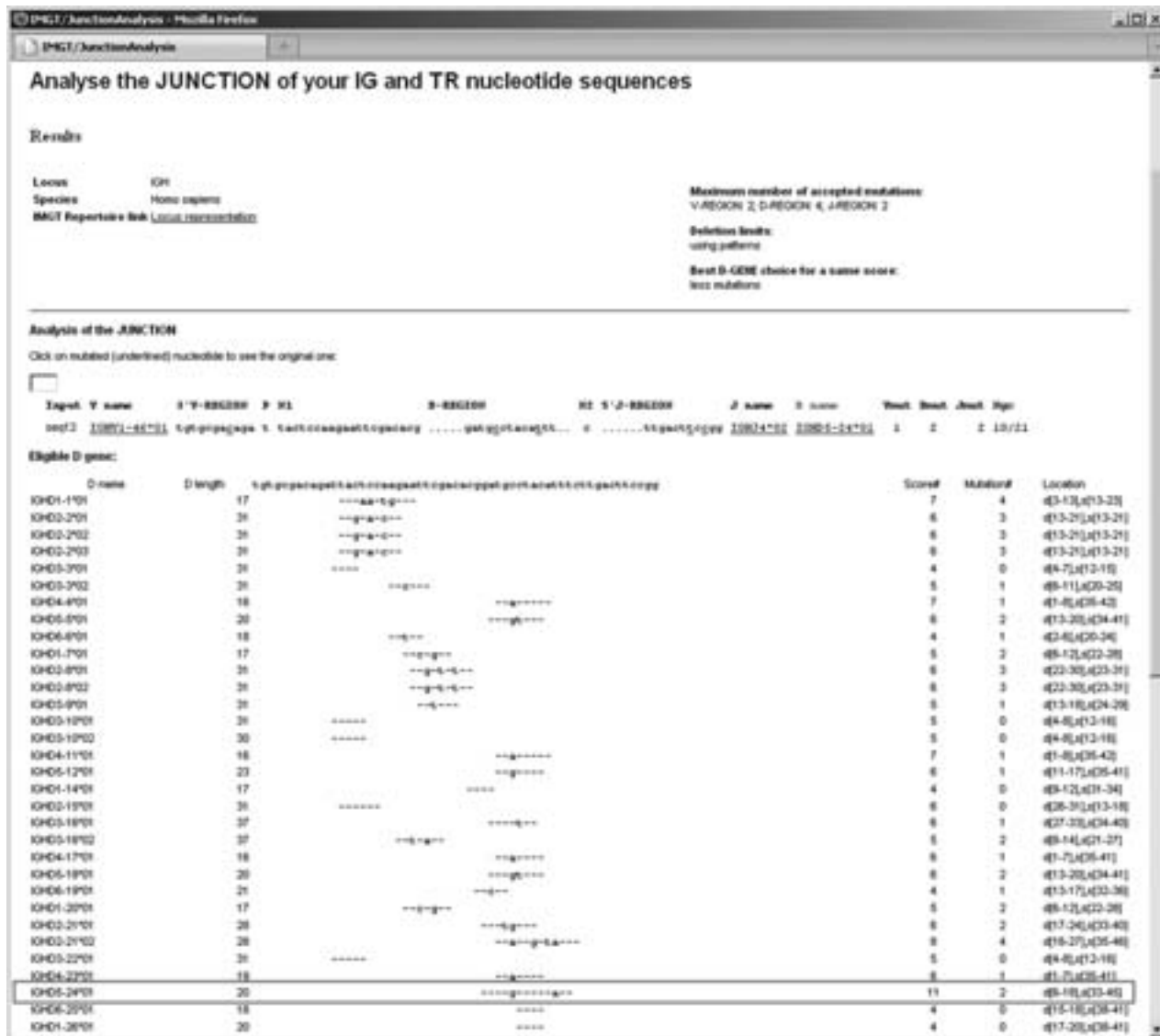
- the display of the 3’V-REGION, D-REGION, and 5’J-REGION, with trimmed nucleotides shown by gaps;
- the D-GENE and allele name for IGH, TRB, and TRD loci;
- the N regions (N or N1, N2... if several, numbered from left to right) and P regions (P) (there is no numbering for the P-REGION);
- the number of mutations found in the 3’V-REGION, D-REGION, and 5’J-REGION (Vmut, Dmut, and Jmut, respectively), by comparison to the corresponding germline allele sequences;
- the ratio of the number of G+C nucleotides to the total number of N region nucleotides (Ngc).

You can click on a mutated (underlined) nucleotide to see the original one in the closest germline region.

Figure 3 shows the results obtained with an example of customization of Advanced parameters on one junction sequence.

### 3. “JUNCTION alignments with translation and IMGT AA classes” (Fig. 4).

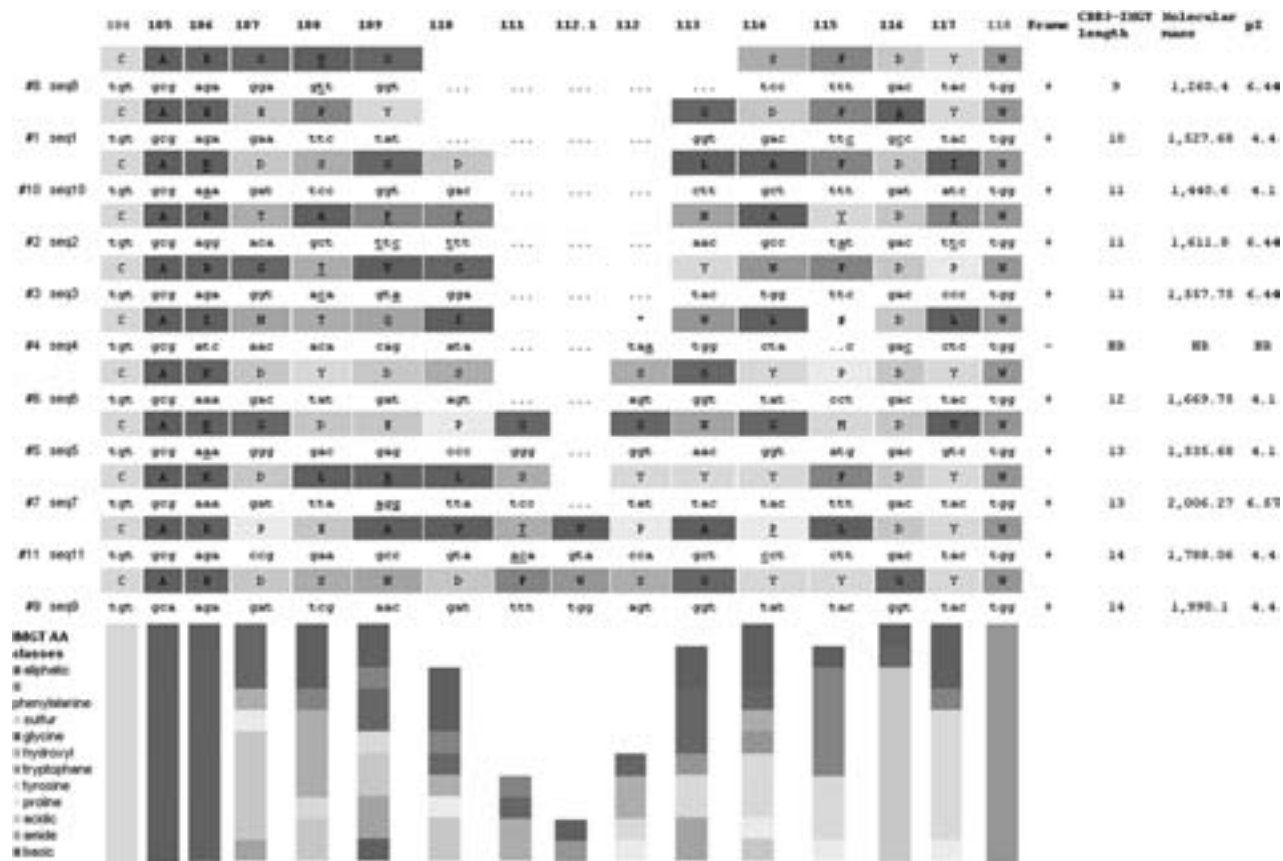
The “JUNCTION alignments with translation and IMGT AA classes” (Fig. 4) provides the results of the analysis of the junctions at the AA level. Each JUNCTION nucleotide sequence is translated in amino



**FIGURE 3.** IMGT/JunctionAnalysis Results page “Analysis of the JUNCTION” with an example of customization of advanced parameters. In this example, the parameter “5’ and 3’ ends of the JUNCTION” has been set to “May start and/or end with any codon” to allow the analysis of that IGH junction that ends in 3’ with “cgg” instead of the classical “tgg” due to a mutation. Because there is only one sequence analyzed in that run, the list of eligible D genes is displayed by default. The D gene and allele with the best score is shown in the red rectangle. The location d[6-18],s[33-45] means that the nucleotides from position 6 to 18 of the germline IGHD5-24\*01 D-REGION match the nucleotides of the junction sequence from position 33 to 45. Seq12 corresponds to the junction of the sequence with accession number AF006519 from the IMGT/LIGM-DB database (Giudicelli et al. 2006).

acid sequences. In the case of frameshifts, gaps indicated by one or two dots are inserted to maintain the J-REGION reading frame and to facilitate sequence comparison. Codons and AA are numbered according to the IMGT unique numbering for V-DOMAIN (Lefranc 1997, 1999; Lefranc et al. 2003). The numbering is made according to the longest JUNCTION obtained in the results. Colors of the AA classes are according to the 11 IMGT physicochemical AA classes (Pommié et al. 2004). Note that:

- Gaps inserted in JUNCTION may split a D-REGION or a J-REGION, since the gaps are localized at the top of the CDR3-IMGT loops and depend on the CDR3-IMGT lengths and not on the sequence alignment.
- “\*” indicates a STOP-CODON.



**FIGURE 4.** IMGT/JunctionAnalysis Results page: “JUNCTION alignments with translation and IMGT AA classes.” On the *top*, the junctions are displayed (by “CDR3-IMGT length increasing order”). AA colors are according to the 11 IMGT physicochemical classes (Pommié et al. 2004). (\*) a STOP-CODON (in seq4). In the “Frame” column: (+) an in-frame junction; (–) an out-of-frame junction. In the case of frame-shifts, gaps indicated by one or two dots are inserted to maintain the J-REGION reading frame and to facilitate sequence comparison and the sign (#) is indicated in the AA sequence (in seq 4). Underlined nucleotides and amino acids are mutated compared with the germline genes (the original nucleotide or amino acids can be seen by clicking on the mutated one). For each junction, the length of the CDR3, the molecular mass, and the isoelectric point (pI) are shown on the *right*. NR indicates that the information is not relevant for out-of-frame junctions. On the *bottom*, a histogram represents the repartition of the 11 IMGT physicochemical AA classes (Pommié et al. 2004) in the junctions analyzed in the same run. Same sequences as in Figure 2.

- “#” indicates a frameshift.
- “+” and “–” at the end of the line indicates “in-frame” and “out-of-frame” JUNCTION, respectively.
- The CDR3-IMGT length, Molecular mass and pI values are indicated as “NR” (not relevant) in case of “out-of-frame” JUNCTION.

You can click on mutated nt or changed AA (underlined) to see the original nucleotide or amino acid in the closest germline region.

### Uses of IMGT/JunctionAnalysis

IMGT/JunctionAnalysis (<http://www.imgt.org>) provides a highly detailed and standardized analysis of the IG and TR JUNCTION sequences: it describes precisely, at the nucleotide level, all characteristics of the junctions according to the complex mechanisms of IG and TR synthesis. IMGT/JunctionAnalysis is currently available for the analysis of junctions from species whose IG and TR loci are extensively studied (human, mouse). Other species (rat, rabbit, trout, etc.) are added as the genomic sequences are annotated in IMGT. Standardization of the results is based on the axioms and concepts of



IMGT-ONTOLOGY (Giudicelli and Lefranc 1999; Lefranc et al. 2004, 2005a, 2008; Duroux et al. 2008). IMGT/JunctionAnalysis has been integrated in IMGT/V-QUEST (Lefranc 2004; Giudicelli and Lefranc 2005, 2008; Brochet et al. 2008) (**IMGT/V-QUEST: IMGT Standardized Analysis of the Immunoglobulin (IG) and T Cell Receptor (TR) Nucleotide Sequences**) (Giudicelli et al. 2011) and in IMGT/HighV-QUEST (Alamyar et al. 2010, 2011). This allows the IMGT users to get, in a single tool, the identification of the V and J gene and allele names of IG and TR rearranged sequence, and the detailed analysis of the JUNCTION according to IMGT/JunctionAnalysis. In its version integrated in IMGT/V-QUEST and in IMGT/HighV-QUEST, IMGT/JunctionAnalysis used the default parameters. The benefit of the IMGT/JunctionAnalysis stand-alone version, available at the IMGT Home page (<http://www.imgt.org>), is that parameters can be modified by the users. Sequences of the junctions are provided by IMGT/V-QUEST (as “Input for IMGT/JunctionAnalysis” in “Sequences of the JUNCTION (“nt” and “AA”)”) for independent analysis in the stand-alone IMGT/JunctionAnalysis. Owing to the accuracy of the results, both tools are used by IMGT/Automat for the automatic annotation of the IG and TR cDNA sequences of IMGT/LIGM-DB (Giudicelli et al. 2003; 2005b). Results obtained by the users have important implications in basic and medical research (Stamatopoulos et al. 2007; Giudicelli and Lefranc 2008), antibody and TR specificity, and in biotechnology related to antibody engineering (Lefranc et al. 2008; Lefranc 2009; Ehrenmann et al. 2010).

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# IMGT/Collier de Perles: IMGT Standardized Representation of Domains (IG, TR, and IgSF Variable and Constant Domains, MH and MhSF Groove Domains)

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## INTRODUCTION

IMGT/Collier de Perles is the IMGT online tool that allows one to draw standardized IMGT two-dimensional (2D) graphical representations of protein domains, or IMGT Colliers de Perles, starting from the user's own domain amino acid (AA) sequences. The use of this tool is described here. IMGT Colliers de Perles can be drawn for three domain types: the variable (V) domain and constant (C) domain of the immunoglobulin (IG) and T cell receptor (TR) and other members of the immunoglobulin superfamily (IgSF), and the groove (G) domain of the major histocompatibility (MH) and other members of the MH superfamily (MhSF). Sequences have to be gapped according to the IMGT unique numbering, using, for example, IMGT/DomainGapAlign. Resulting IMGT Colliers de Perles allow one to quickly visualize amino acids, which are important for the three-dimensional (3D) structural configuration, and to delimit the standardized framework regions (FR-IMGT) and complementarity determining regions (CDR-IMGT) of the IG and TR V-DOMAIN. The length of the strands, loops, and turns and, for the G type, the length of the helix represented in IMGT Colliers de Perles become crucial information in the domain characterization.

## RELATED INFORMATION

IMGT/Collier de Perles (Ruiz and Lefranc 2002; Kaas and Lefranc 2007; Kaas et al. 2007) is a tool of IMGT <http://www.imgt.org> (Lefranc et al. 2009) using standardization based on the IMGT-ONTOLOGY concepts of identification, classification, description (Giudicelli and Lefranc 1999; Lefranc et al. 2004, 2005a, 2008) and numerotation (Lefranc 1997, 1999; Lefranc et al. 2003, 2005b,c), generated from the axioms of the Formal IMGT-ONTOLOGY or IMGT-Kaleidoscope (Duroux et al. 2008). The concepts of identification led to the IMGT standardized keywords, the concepts of classification to the IMGT standardized gene and allele names, the concepts of description to the IMGT standardized labels, and the concepts of numerotation to the IMGT unique numbering.

A detailed description of IMGT is provided in **IMGT, the International ImMunoGeneTics Information System** (Lefranc 2011a). Information is also available on **From IMGT-ONTOLOGY IDENTIFICATION Axiom to IMGT Standardized Keywords: For Immunoglobulins (IG), T Cell Receptors (TR), and Conventional Genes** (Lefranc 2011b), **From IMGT-ONTOLOGY DESCRIPTION Axiom to IMGT Standardized Labels: For Immunoglobulin (IG) and T Cell Receptor (TR) Sequences and Structures** (Lefranc 2011c), **From IMGT-ONTOLOGY CLASSIFICATION Axiom to IMGT Standardized Gene and Allele Nomenclature: For Immunoglobulins (IG) and T Cell Receptors (TR)** (Lefranc 2011d), **IMGT Unique Numbering for the Variable (V), Constant (C), and Groove (G) Domains of IG, TR, MH, IgSF, and MhSF** (Lefranc 2011e), and **IMGT Collier de Perles for the Variable (V), Constant (C), and Groove (G) Domains of IG, TR, MH, IgSF, and MhSF** (Lefranc 2011f).

In addition, protocols are available for **IMGT/V-QUEST: IMGT Standardized Analysis of the Immunoglobulin (IG) and T Cell Receptor (TR) Nucleotide Sequences** (Giudicelli et al. 2011), **IMGT/JunctionAnalysis: IMGT Standardized Analysis of the V-J and V-D-J Junctions of the Rearranged Immunoglobulins (IG) and T Cell Receptors (TR)** (Giudicelli and Lefranc 2011),

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IMGT/DomainGapAlign: IMGT Standardized Analysis of Amino Acid Sequences of Variable, Constant, and Groove Domains (IG, TR, MH, IgSF, MhSF) (Ehrenmann and Lefranc 2011a), and IMGT/3Dstructure-DB: Querying the IMGT Database for 3D Structures in Immunology and Immunoinformatics (IG or Antibodies, TR, MH, RPI, and FPIA) (Ehrenmann and Lefranc 2011b).

## MATERIALS

It is essential that you consult the appropriate Material Safety Data Sheets and your institution's Environmental Health and Safety Office for proper handling of equipment and hazardous materials used in this protocol.

### Equipment

Computer (Internet-connected).

## METHOD

The IMGT/Collier de Perles tool web application at the IMGT website (<http://www.imgt.org>) allows one to “Make Your Own IMGT Colliers de Perles” for V domain, C domain, or G domain starting from a user's amino acid sequence. This protocol describes how to draw “IMGT Colliers de Perles” using the IMGT/Collier de Perles tool.

### IMGT Collier de Perles for V Domain

1. Using any modern web browser, go to the IMGT Home page <http://www.imgt.org>.
2. Click on the “IMGT/Collier de Perles” link in the IMGT tools section of the IMGT Home page. *This will take you to the IMGT/Collier de Perles tool web application (Fig. 1). By default, the web application is configured for V type domains.*
3. Locate the “Domain type” drop-down list: “Variable (V)” is displayed. *“Variable (V)” allows one to draw IMGT Colliers de Perles for variable domains, which include the V-DOMAIN of IG and TR and the V-LIKE-DOMAIN of IgSF other than IG and TR.*

WELCOME !  
to IMGT/Collier-de-Perles

THE INTERNATIONAL IMMUNOGENETICS INFORMATION SYSTEM

Im Muno Gene Tics  
Information system  
<http://www.imgt.org>

Make Your Own IMGT Collier de Perles

Domain type: Variable (V) [v]

Number of layers: 1 [1]

CDR-IMGT color type: DRAW AS THE TREE [v]

Background color: 50% hydrophobic positions [v]

Domain sequence: DIVYKESGP...ELLPQETL...LITSPQFSL...TYPSHWQDPKQLEKAW...  
DME...DWRYPALK...SLTTSKDTSSQVFLK...DASHTQIPATYTCARRSD...  
YQMFDMGGDTLYVA

Position	Length	Numbering start

Amino acid insertions: [text input] [ADD]

CDR-IMGT length: 1 [1]

Your domain title: [text input]

Draw!

FIGURE 1. IMGT/Collier de Perles Welcome page. By default the page is configured for a V domain.

4. Locate the “Number of layers” drop-down list. Select “1” or “2,” depending on the number of layers you want for the display of the IMGT Collier de Perles. Default is “1.”

*This choice is only available for V and C domains.*

5. Locate the “CDR-IMGT color type” drop-down list, and select “1 (RPI,IGH,TRB,TRD)” or “2 (IGK,IGL,TRA,TRG),” depending on the locus to which the sequence belongs.

*This choice is only available for V domains.*

*For “1 (RPI,IGH,TRB,TRD),” CDR-IMGT regions are colored as follows: CDR1-IMGT (red), CDR2-IMGT (orange), and CDR3-IMGT (purple).*

*For “2 (IGK,IGL,TRA,TRG),” CDR-IMGT regions are colored as follows: CDR1-IMGT (blue), CDR2-IMGT (green), and CDR3-IMGT (greenblue).*

6. Locate the “Background color” drop-down list, and select one of the 10 lines, depending on the property you want to highlight in your IMGT Collier de Perles, by comparison with previously analyzed domains of the same locus (Pommié et al. 2004).

*This choice is only available for V domains. Ten selections are provided (the first one by default):*

- “50% Hydrophobic positions:” positions that have a hydrophobic amino acid (hydropathy index with positive value) or a tryptophan (W) like 50% or more of analyzed V domains (blue positions in IMGT Collier de Perles).
- “IGH 80% hydropathy classes:” positions with amino acids that have the same hydropathy class, like 80% or more of analyzed IGH V domains (three classes).
- “IGK 80% hydropathy classes:” positions with amino acids that have the same hydropathy class, like 80% or more of analyzed IGK V domains (three classes).
- “IGL 80% hydropathy classes:” positions with amino acids that have the same hydropathy class, like 80% or more of analyzed IGL V domains (three classes).
- “IGH 80% volume classes:” positions with amino acids that have the same volume class, like 80% or more of analyzed IGH V domains (five classes).
- “IGK 80% volume classes:” positions with amino acids that have the same volume class, like 80% or more of analyzed IGK V domains (five classes).
- “IGL 80% volume classes:” positions with amino acids that have the same volume class, like 80% or more of analyzed IGL V domains (five classes).
- “IGH 80% physicochemical classes:” positions with amino acids that have the same physicochemical class, like 80% or more of analyzed IGH V domains (11 classes).
- “IGK 80% physicochemical classes:” positions with amino acids that have the same physicochemical class, like 80% or more of analyzed IGK V domains (11 classes).
- “IGL 80% physicochemical classes:” positions with amino acids that have the same physicochemical class, like 80% or more of analyzed IGL V domains (11 classes).

7. Locate the “Domain sequence” text area, and enter your gapped sequence obtained, for example, from **IMGT/DomainGapAlign (IMGT/DomainGapAlign: IMGT Standardized Analysis of Amino Acid Sequences of Variable, Constant, and Groove Domains (IG, TR, MH, IgSF, MhSF)** (Ehrenmann and Lefranc 2011a).

*A complete IMGT Collier de Perles of an IG or TR V-DOMAIN includes the CDR3-IMGT and the FR4-IMGT. The FR4-IMGT is composed of at least nine or 10 AA beyond the phenylalanine F (J-PHE 118) or tryptophan W (J-TRP 118) of the motif F/W-G-X-G that characterizes the J-REGION.*

*If the length of the CDR3-IMGT of your sequence is less than 13, you need to add gaps at the top of the loop (Lefranc et al. 2003; Lefranc 2011e) <http://www.imgt.org/textes/IMGTScientificChart/Numbering/IMGTIGVsuperfamily.html>. If the CDR3-IMGT is equal to or greater than 13, the tool will automatically adjust the number of positions to the length (provided that you fill in the required field in Step 9).*

8. Locate the “Amino acid insertions” text area, and enter, if needed, amino acid insertions.

*This option is only needed if you deal with an unusual sequence characterized by insertions compared with the reference sequences. This can be the case for the IgSF V-LIKE-DOMAINS that have amino acid insertions compared with the IG and TR V-DOMAIN. The program for renumbering is based on the Smith and Waterman alignment, which uses IMGT*

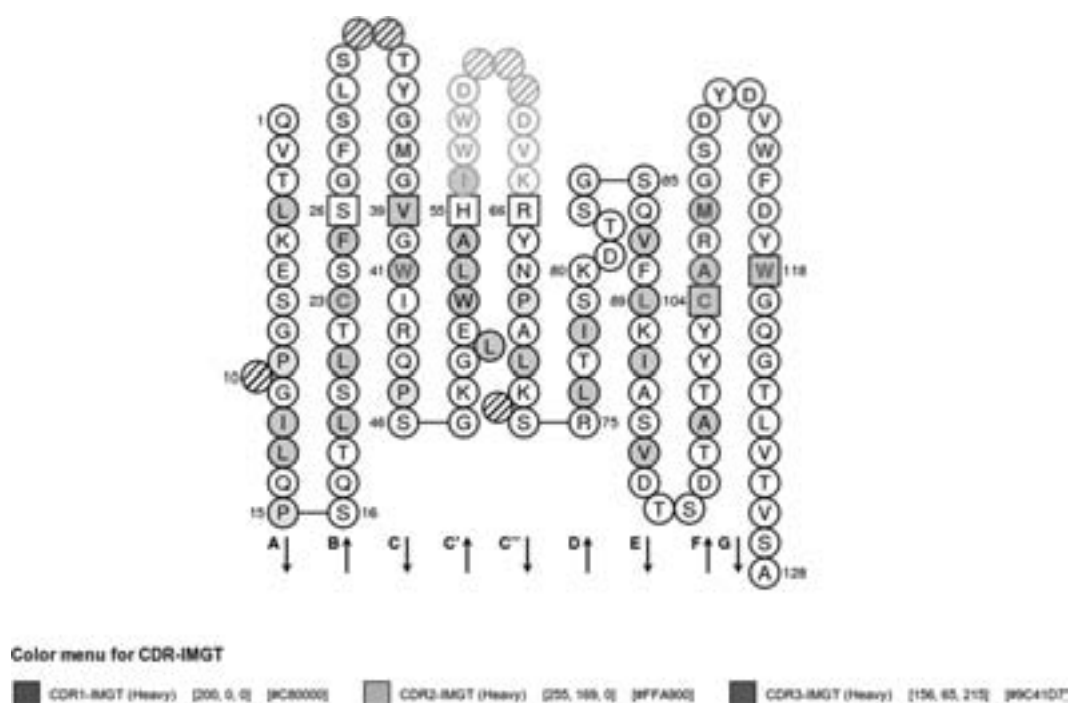


insertions as new amino acids and strongly penalizes the opening of gaps in the closest reference sequence. To add an insertion, enter the position that precedes the insertion, the length (that is the number of amino acids to add) and the numbering label(s) of the novel positions: for examples "84,1,84A" means that, following 84, one additional amino acid labeled 84A has to be added; "84,3,84A,84B,84C" means that, following 84, three additional amino acids labeled 84A, 84B, and 84C have to be added. Then click "ADD." Repeat the process if several insertions have to be entered.

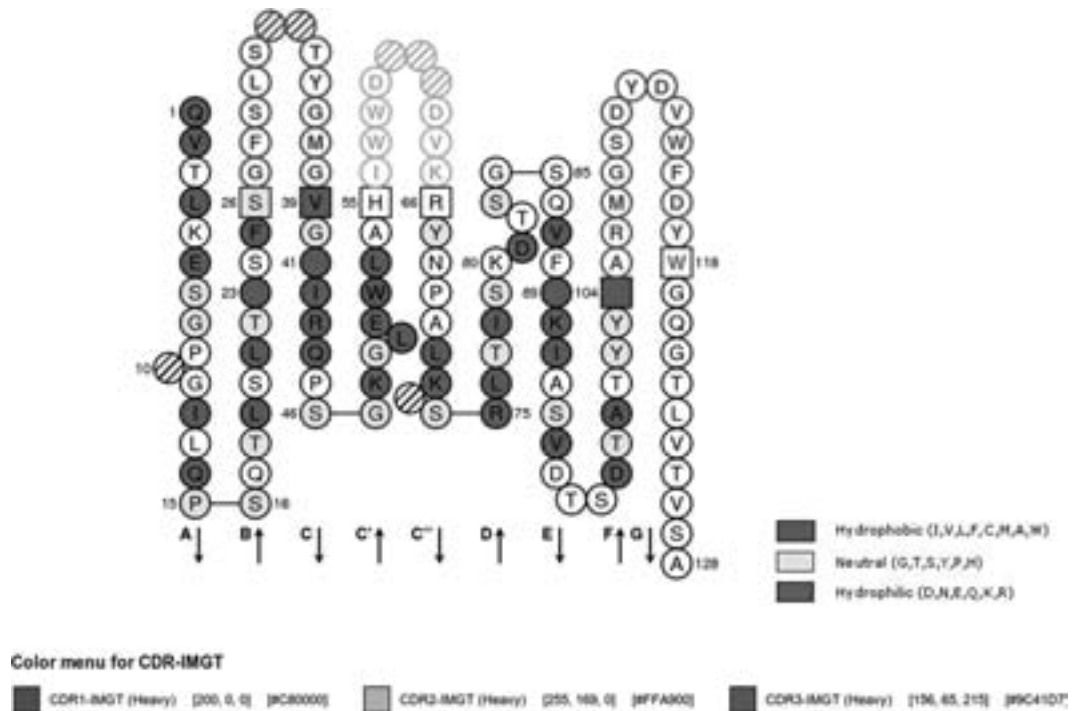
9. Locate the "CDR3-IMGT length" text area, and enter the length of the CDR3-IMGT (required).
10. (Optional) Locate the "Your domain title" text area, and enter a name that you would like to be displayed in the figure.
11. Press the "Draw !" button to visualize the IMGT Collier de Perles for V domain.  
*This will return a page showing the IMGT Collier de Perles taking into account your choices (for examples, see Figs. 2–6). In the case of unusual sequences, it is possible to proceed by trial and error, adding or deleting gaps manually, in the "Domain sequence" text area (Step 7).*

### IMGT Collier de Perles for C Domain

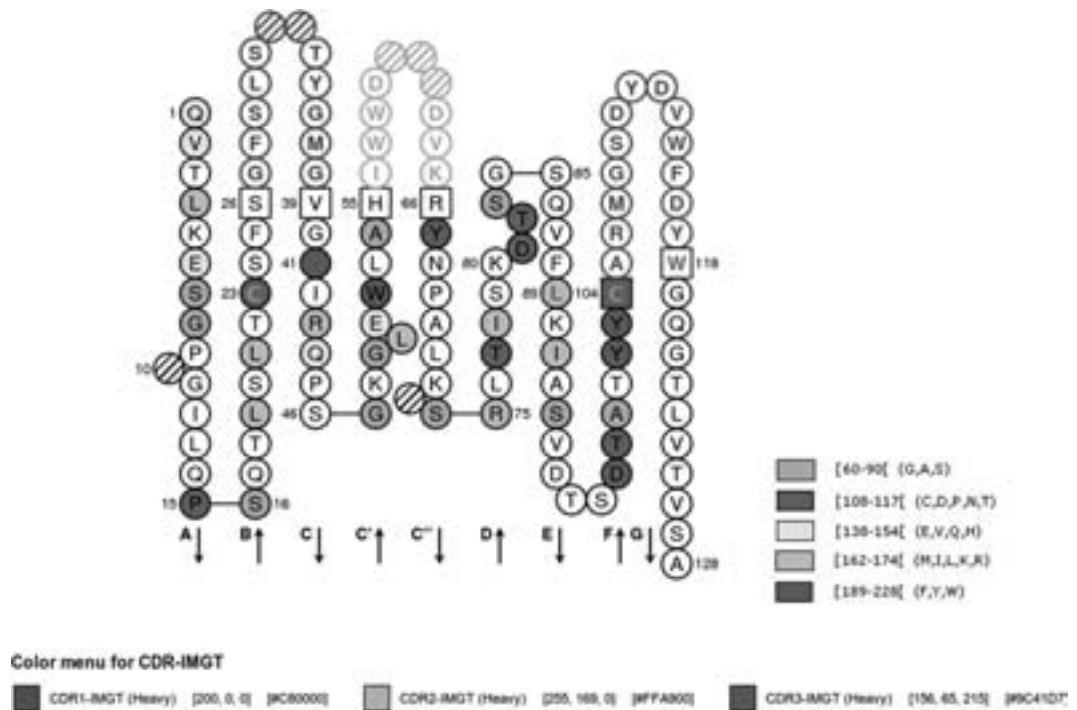
12. Follow Steps 1 and 2 above.
13. Locate the "Domain type" drop-down list, and select the domain type "Constant (C)." This will change the page of the web application (Fig. 7).  
*"Constant (C)" allows one to draw IMGT Colliers de Perles for constant domains, which include the C-DOMAIN of IG and TR and the C-LIKE-DOMAIN of IgSF other than IG and TR.*



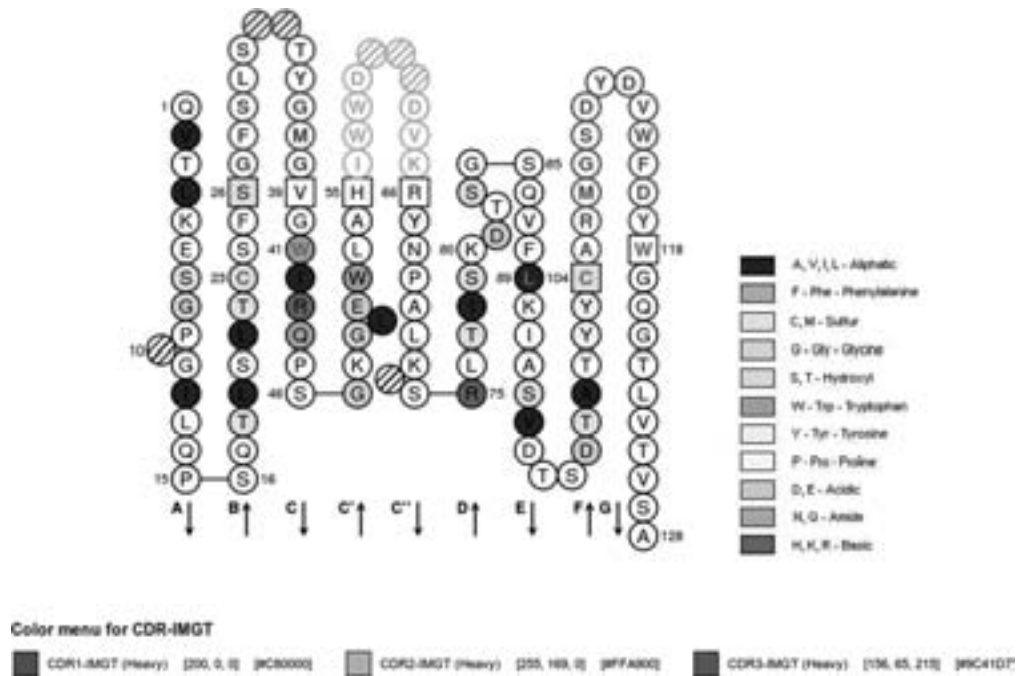
**FIGURE 2.** IMGT Collier de Perles of a V domain, on one layer, obtained after selection of "1 (RPI,IGH,TRB,TRD)" in CDR-IMGT color type, the domain belonging to an IGH chain, and by default "50% Hydrophobic positions." Positions in blue mean that the amino acid of the user sequence at these positions is hydrophobic (hydropathy index with positive value) or is a tryptophan (W), like in 50% or more of analyzed V domains. Positions with red and bold letters indicate the five conserved positions of a V domain: 1st-CYS 23, CONSERVED-TRP 41, hydrophobic 89, 2nd-CYS 104, and J-TRP 118. Anchor positions are in squares. Hatched positions correspond to gaps according to the IMGT unique numbering for V domain (Lefranc 1997, 1999; Lefranc et al. 2003). Prolines are shown in yellow. Arrows indicate the  $\beta$  strands and their direction. The CDR-IMGT lengths of this domain are [10.7.13].



**FIGURE 3.** IMGT Collier de Perles of a V domain with “IGH 80% hydropathy classes” positions. Colored positions (three classes) mean that the property of the amino acids of the user sequence at these positions is found in >80% of the analyzed IGH V domains.



**FIGURE 4.** IMGT Collier de Perles of a V domain with “IGH 80% volume classes” positions. Colored positions (five classes) mean that the property of the amino acids of the user sequence at these positions is found in >80% of the analyzed IGH V domains.

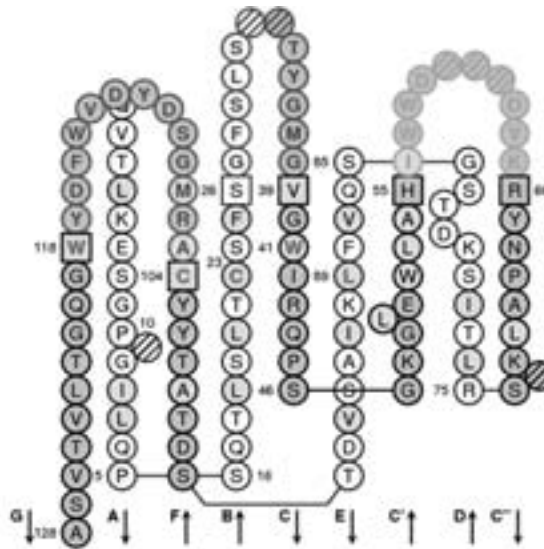


**FIGURE 5.** IMGT Collier de Perles of a V domain with “IGH 80% physicochemical classes” positions. Colored positions (11 classes) mean that the property of the amino acids of the user sequence at these positions is found in >80% of the analyzed IGH V domains.

14. Locate the “Number of layers” drop-down list. Select “1” or “2,” depending on the number of layers you want for the display of the IMGT Collier de Perles. Default is “1.”  
*This choice is only available for V and C domains.*
15. Locate the “Domain sequence” text area, and enter your gapped sequence obtained, for example, from IMGT/DomainGapAlign.
16. Locate the “Amino acid insertions” text area, and enter, if needed, amino acid insertions.  
*This option is only needed if you deal with an unusual sequence characterized by insertions compared with the reference sequences. This can be the case for IgSF C-LIKE-DOMAIN that have amino acid insertions compared with the IG and TR C-DOMAIN. See Step 8 for more information.*
17. (Optional) Locate the “Your domain title” text area, and enter a name that you would like to be displayed in the figure.
18. Press the “Draw !” button to visualize the IMGT Collier de Perles for C domain.  
*This will return a page showing the IMGT Collier de Perles, taking into account your choices (e.g., see Figs. 8 and 9). In the case of unusual sequences, it is possible to proceed by trial and error, adding or deleting gaps manually in the “Domain sequence” text area (see Step15).*

### IMGT Collier de Perles for G Domain

19. Follow Steps 1 and 2 above.
20. Locate the “Domain type” drop-down list, and select the domain type “Groove (G).” This will change the page of the web application (Fig. 10).  
*“Groove (G)” allows one to draw IMGT Colliers de Perles for groove domains, which include the G-DOMAIN of MH class I (or MH1) and class II (or MH2) and the G-LIKE-DOMAIN of MhSF other than MH (or related proteins of the immune system [RPI]-MH1 Like). As a groove is made of 2 G domains, the sequences of the two domains for the IMGT Colliers de Perles are entered in the same page.*



**Color menu for CDR-IMGT**

CDR1-IMGT (heavy) [200, 9, 0] [KCB000]
  CDR2-IMGT (heavy) [255, 169, 0] [MFA000]
  CDR3-IMGT (heavy) [156, 65, 215] [K9C41D?]

**FIGURE 6.** IMGT Collier de Perles of a V domain, on two layers, obtained after selection of “2” in Number of layers, “1 (RPI, IGH,TRB,TRD)” in CDR-IMGT color type, the domain belonging to an IGH chain, and by default “50% Hydrophobic positions.” Positions in blue mean that the amino acid of the user sequence at these positions is hydrophobic (hydropathy index with positive value) or is a tryptophan (W), like in 50% or more of analyzed V domains. Positions with red and bold letters indicate the five conserved positions of a V domain: 1st-CYS 23, CONSERVED-TRP 41, hydrophobic 89, 2nd-CYS 104, and J-TRP 118. Anchor positions are in squares. Hatched positions correspond to gaps according to the IMGT unique numbering for V domain (Lefranc 1997, 1999; Lefranc et al. 2003). Prolines are shown in yellow. Arrows indicate the  $\beta$  strands and their direction. The CDR-IMGT lengths of this domain are [10.7.13].

21. Locate “Link G domains” and click on the button if your sequences are domains of MH1 or RPI-MH1Like, to have them linked.
22. Locate the “Domain sequence 1” text area, and enter the gapped sequence obtained, for example, from IMGT/DomainGapAlign, for the first G domain.  
*For MH1, the sequence is that of the G-ALPHA1, for MH2, that of G-ALPHA, and for RPI-MH1Like, that of G-ALPHA1-LIKE.*

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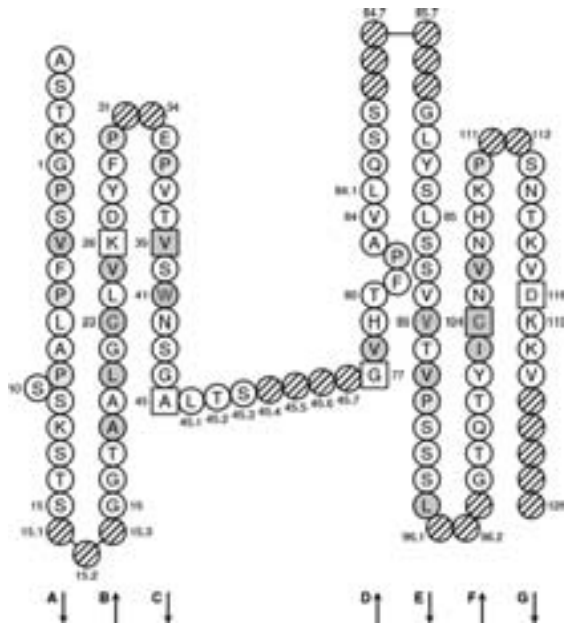
**Make Your Own IMGT Collier de Perles**

- Domain type:
- Number of layers:
- Domain sequence:
 

...ASTKSPSYPLAPGASTS...SITIALDGLWDFP...EPTVSMGIAL  
 TS...SREYFWLSS...ALYLSGWTYFSSL...SPTYLMMW  
 P...SHTGDKKY
- Amino acid insertions:
 

Position	Length	Residue(s)
<input type="text"/>	<input type="text"/>	<input type="text"/>
- Your domain title:

**FIGURE 7.** IMGT/Collier de Perles page for a C domain.



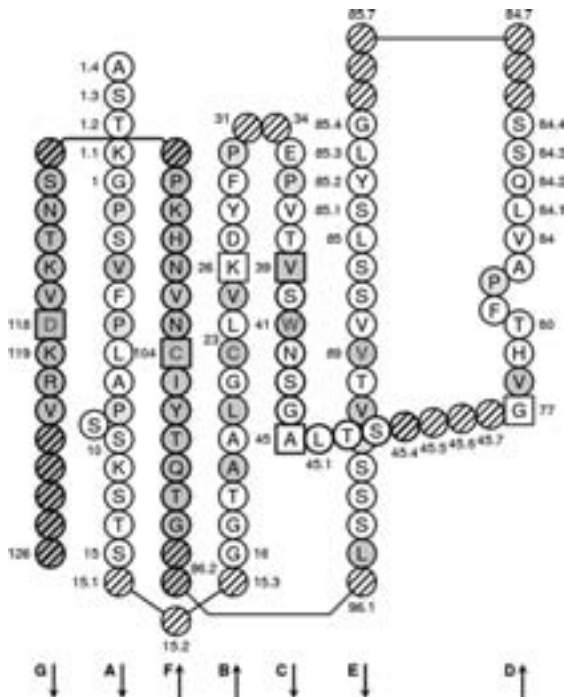
**FIGURE 8.** IMGT Collier de Perles of a C domain on one layer. The default is “50% Hydrophobic positions.” Positions in blue mean that the amino acid of the user sequence at these positions is hydrophobic (hydropathy index with positive value) or is a tryptophan (W), as in 50% or more of analyzed C domains. Positions with red and bold letters are by analogy with a V domain. In a C domain they correspond to the two conserved cysteines, 1st-CYS 23 and 2nd-CYS 104, and to the usually hydrophobic positions 41 and 89, whereas position 118 is not conserved. Anchor positions are in squares. Hatched positions correspond to gaps according to the IMGT unique numbering for C domain (Lefranc et al. 2005b). Prolines are shown in yellow. Arrows indicate the  $\beta$  strands and their direction.

23. Locate the “Amino acid insertions” text area, and enter, if needed, AA insertions.

*This option is only needed if you deal with unusual sequences characterized by insertions compared with the reference sequences. This can be the case for RPI-MH1Like G-LIKE-DOMAIN that have amino acid insertions compared with the MH G-DOMAIN. See Step 8 for more information.*

*Note that position 7A of G-ALPHA1 and positions 61A, 61B, and 72A of G-ALPHA2 are entered by default, as these insertions are characteristic of these domains, respectively.*

24. (Optional) Locate the “Your domain title 1” text area, and enter a name that you would like to be displayed in the figure for the first G domain.



**FIGURE 9.** IMGT Collier de Perles of a C domain on two layers. The default is “50% Hydrophobic positions.” Positions in blue mean that the amino acid of the user sequence at these positions is hydrophobic (hydropathy index with positive value) or is a tryptophan (W), like in 50% or more of analyzed C domains. Positions with red and bold letters indicate four conserved positions of a C domain: 1st-CYS 23, CONSERVED-TRP 41, hydrophobic 89, and 2nd-CYS 104, whereas position 118 (not conserved in a C domain) is only shown by analogy with a V domain. Anchor positions are in squares. Hatched positions correspond to gaps according to the IMGT unique numbering for C domain (Lefranc et al. 2005b). Prolines are shown in yellow. Arrows indicate the  $\beta$  strands and their direction.



Make Your Own IMGT Collier de Perles

The screenshot shows the web interface for creating an IMGT Collier de Perles for G domains. It features several sections with input fields and buttons:

- Domain type:** A dropdown menu set to "G-DOMAIN".
- Link G domains:** A text input field containing "1".
- Domain sequence 1:** A text area containing a gapped amino acid sequence: ".....LQAGRY\_FYTAQKAPQKLEPRF LNVVYDVFQVFQSSAQR  
TQRF.....PACDSEPEKQWTELEFETETETVEKLEKLVYDGEA".
- Amino acid insertions:** A table with columns "Position", "Length", and "Numbering table". Below it are input fields for "1" and "5", and a "ADD" button.
- Your domain title 1:** A text input field containing "G-ALPHA1".
- Domain sequence 2:** A text area containing a gapped amino acid sequence: ".....EAKTDPHFQGLPQKLLAKHQLAKGKYSALMEY LLSW  
TAAR.....TRADTTPKQAA RAREKARVLEGLVENEKFLYKMKETS  
RQ".
- Amino acid insertions:** A table with columns "Position", "Length", and "Numbering table". Below it are input fields for "41", "2", "10", and "1", and a "ADD" button.
- Your domain title 2:** A text input field containing "G-ALPHA2".

At the bottom, there are "Draw" and "Reset" buttons.

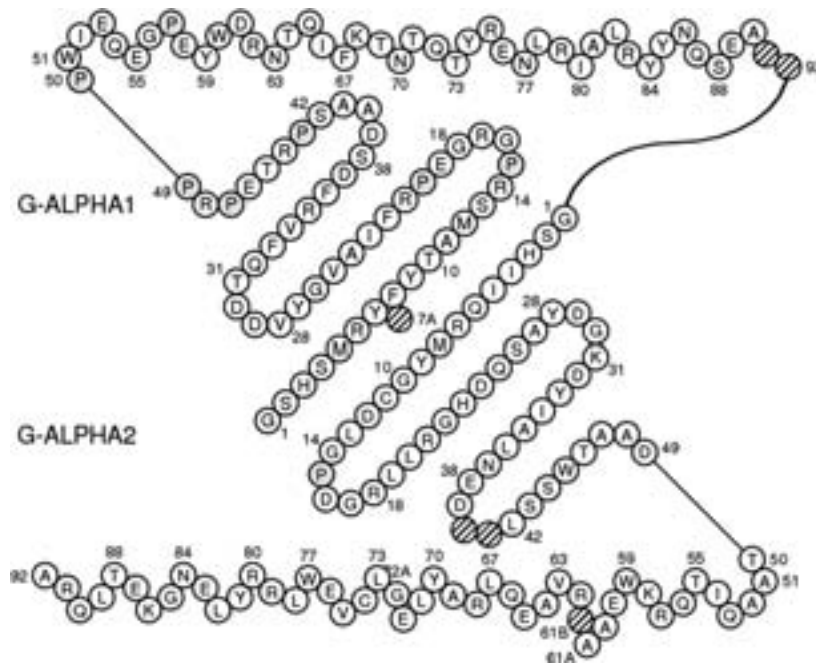
FIGURE 10. Page: IMGT/Collier de Perles for G domains.

25. Locate the "Domain sequence 2" text area, and enter the gapped sequence obtained, for example, from IMGT/DomainGapAlign, for the second G domain.  
*For MH1, the sequence is that of the G-ALPHA2, for MH2, that of G-BETA, and for RPI-MH1 Like, that of G-ALPHA2-LIKE.*
26. Locate the "Amino acid insertions" text area, and enter, if needed, AA insertions.  
*See Step 23 for more information.*
27. (Optional) Locate the "Your domain title 2" text area, and enter a name that you would like to be displayed in the figure for the second G domain.
28. Press the "Draw !" button to visualize the IMGT Collier de Perles for G domain.  
*This will return a page showing the IMGT Collier de Perles taking into account your choices (Fig. 11). In the case of unusual sequences, it is possible to proceed by trial and error, adding or deleting gaps manually, in the "Domain sequence" text area (see Step 22).*

## DISCUSSION

The IMGT Colliers de Perles allow a precise visualization of the interspecies differences for the IgSF V and C domain strands and loops and MhSF G domain strands and helix, even in the absence of 3D structures (Lefranc et al. 2008). The IMGT Colliers de Perles are particularly useful in molecular engineering and antibody humanization design based on CDR grafting to visualize the CDR-IMGT (Lefranc 2009; Ehrenmann et al. 2010a) and to easily compare the amino acid sequences of the four FR-IMGT between the murine or nonhuman primate and the closest human V-DOMAIN (Pelat et al. 2008). A recent analysis performed on humanized antibodies used in oncology underlines the importance of correct delimitation of the CDR regions to be grafted (Magdelaine-Beuzelin et al. 2007). The IMGT Colliers de Perles also allow a comparison to the IMGT Collier de Perles statistical profiles for the human expressed IGHV, IGKV, and IGLV repertoires (Pommié et al. 2004).

The IMGT Colliers de Perles for the V, C, and G domains, based on the IMGT unique numbering (Lefranc 1997, 1999; Lefranc et al. 2003, 2005b,c) therefore represent a major step forward for the comparative analysis of the sequences and structures of the IgSF and MhSF domains (Kaas et al. 2004; **IMGT/DomainGapAlign: IMGT Standardized Analysis of Amino Acid Sequences of Variable, Constant, and Groove Domains (IG, TR, MH, IgSF, MhSF)** (Ehrenmann and Lefranc 2011a; **IMGT/3Dstructure-DB: Querying the IMGT Database for 3D Structures in Immunology and**



**FIGURE 11.** IMGT Colliers de Perles of G domains of MH1. The domains are linked as selected by the user. Hatched positions correspond to gaps according to the IMGT unique numbering for G domain (Lefranc et al. 2005c). Prolines are shown in yellow.

**Immunoinformatics (IG or Antibodies, TR, MH, RPI, and FPIA)** (Ehrenmann and Lefranc 2011b; Ehrenmann et al. 2010b) and the study of their evolution. In basic and clinical research, the IMGT Colliers de Perles provide new insights in the analysis of the IG and TR repertoires in autoimmune diseases and leukemias (Stamatopoulos et al. 2007), in the study of interactions between IG and antigen (IG/Ag) and between TR, peptide and MH (TR/pMH), and in antibody engineering and humanization for therapeutic applications (Lefranc 2009; Ehrenmann et al. 2010a).

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# IMGT/DomainGapAlign: IMGT Standardized Analysis of Amino Acid Sequences of Variable, Constant, and Groove Domains (IG, TR, MH, IgSF, MhSF)

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## INTRODUCTION

IMGT/DomainGapAlign is the IMGT online tool for the analysis of amino acid (AA) sequences and two-dimensional (2D) structures of domains. The use of this tool is described here. Three domain types can be analyzed: the variable (V) and constant (C) domains of the immunoglobulins (IG) and T cell receptors (TR) and other immunoglobulin superfamily (IgSF) proteins, and the groove (G) domain of major histocompatibility (MH) and other MH superfamily (MhSF) proteins. IMGT/DomainGapAlign creates gaps in the user amino acid sequences and delimits, for the V and C domains, the framework regions (FR-IMGT) and complementarity determining regions (CDR-IMGT), and, for the G domains, the strands and helix, according to the IMGT unique numbering. The tool provides alignments with the closest sequences from the IMGT domain directory of IMGT/DomainDisplay. For the IG and TR V domains that result from V-(D)-J rearrangement, the tool provides alignments with the translation of the IMGT/GENE-DB germline V and joining (J) genes. IMGT/DomainGapAlign displays the amino acid changes, highlights them in IMGT Colliers de Perles, and provides tables with their detailed description, according to the IMGT rules. Several amino acid sequences can be analyzed simultaneously, provided that they belong to the same domain type.

## RELATED INFORMATION

IMGT/DomainGapAlign (Ehrenmann et al. 2010b) is part of IMGT, the international ImMunoGeneTics information system, <http://www.imgt.org> (Lefranc et al. 2009). Standardization and IMGT Scientific chart rules are based on the IMGT-ONTOLOGY concepts of identification, classification, description (Giudicelli and Lefranc 1999; Lefranc et al. 2004, 2005a, 2008) and numerotation (Lefranc 1997, 1999; Lefranc et al. 2003, 2005b,c) generated from the axioms of the Formal IMGT-ONTOLOGY or IMGT-Kaleidoscope (Duroux et al. 2008). The concepts of identification led to the IMGT standardized keywords, the concepts of classification to the IMGT standardized gene and allele names, the concepts of description to the IMGT standardized labels, and the concepts of numerotation to the IMGT unique numbering.

The IMGT/DomainGapAlign tool web application at the IMGT website allows one to align the user amino acid domain sequences, to identify, in terms of sequence identity, the closest V, C, and G domains in the IMGT domain reference directory (Lefranc 2010d; Giudicelli et al. 2005), to create gaps according to the IMGT unique numbering (Lefranc 1997, 1999; Lefranc et al. 2003, 2005b,c), to highlight differences with the closest reference(s), and to generate IMGT Colliers de Perles (**IMGT/Collier de Perles: IMGT Standardized Representation of Domains (IG, TR, and IgSF Variable and Constant Domains, MH and MhSF Groove Domains)**) (Ehrenmann et al. 2011). Several amino acid sequences can be analyzed simultaneously, provided that they belong to the same domain type (V, C, or G).

A detailed description of IMGT is provided in **IMGT, the International ImMunoGeneTics Information System** (Lefranc 2011a). Information is also available on **From IMGT-ONTOLOGY IDENTIFICATION Axiom to IMGT Standardized Keywords: For Immunoglobulins (IG), T Cell Receptors (TR), and Conventional Genes** (Lefranc 2011b), **From IMGT-ONTOLOGY DESCRIPTION Axiom to IMGT Standardized Labels: For Immunoglobulin (IG) and T Cell Receptor (TR) Sequences and Structures** (Lefranc 2011c), **From IMGT-ONTOLOGY CLASSIFICATION Axiom to IMGT Standardized**

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**Gene and Allele Nomenclature: For Immunoglobulins (IG) and T Cell Receptors (TR)** (Lefranc 2011d), **IMGT Unique Numbering for the Variable (V), Constant (C), and Groove (G) Domains of IG, TR, MH, IgSF, and MhSF** (Lefranc 2011e), and **IMGT Collier de Perles for the Variable (V), Constant (C), and Groove (G) Domains of IG, TR, MH, IgSF, and MhSF** (Lefranc 2011f).

In addition, protocols are available for **IMGT/V-QUEST: IMGT Standardized Analysis of the Immunoglobulin (IG) and T Cell Receptor (TR) Nucleotide Sequences** (Giudicelli et al. 2011), **IMGT/Junction Analysis: IMGT Standardized Analysis of the V-J and V-D-J Junctions of the Rearranged Immunoglobulins (IG) and T Cell Receptors (TR)** (Giudicelli and Lefranc 2011), and **IMGT/3Dstructure-DB: Querying the IMGT Database for 3D Structures in Immunology and Immunoinformatics (IG or Antibodies, TR, MH, RPI, and FPIA)** (Ehrenmann and Lefranc 2011).

## MATERIALS

It is essential that you consult the appropriate Material Safety Data Sheets and your institution's Environmental Health and Safety Office for proper handling of equipment and hazardous materials used in this protocol.

### Equipment

Computer (Internet-connected)

## METHOD

### Querying IMGT/DomainGapAlign Tool

1. Using any modern web browser, go to the IMGT Home page <http://www.imgt.org>.
2. Click on the "IMGT/DomainGapAlign" link in the "IMGT tools" section of the IMGT Home page. *This will take you to the IMGT/DomainGapAlign tool web application (Fig. 1)*
3. Locate the text area at the top of the page. Paste your amino acid sequences in FASTA format. Alternatively, you can upload a file.

*A precise delimitation of the domain sequences is not required; however, if the sequence contains several domains, the sequence should be split between the different domains. Several domains can be analyzed simultaneously, but each*

WELCOME !  
to **IMGT/DomainGapAlign**  
THE INTERNATIONAL IMMUNOLOGICAL INFORMATION SYSTEM

Align and "IMGT-gap" your domain amino acid sequence

Paste your protein sequences in FASTA format below

Sequences names must be different !

Upload a file:  None

Select a domain type:

Select a species:

Display alignments:

Alignment:

Putting gaps in the sequences:

Use priority for queries:

FIGURE 1. IMGT/DomainGapAlign Welcome page.



sequence must have a distinct name. If the limits and the numbers of domains of an amino acid sequence are unknown, you can progressively analyze the protein, shortening the sequence once a domain has been identified by the tool. (Note that the first domain identified by the tool is not necessarily the first one in the protein).

4. Locate the “Domain type” drop-down list, and select the domain type (V, C, or G) corresponding to the sequences to analyze.
  - V for “variable” domain (V-DOMAIN of IG and TR and V-LIKE-DOMAIN of IgSF other than IG and TR)
  - C for “constant” domain (C-DOMAIN of IG and TR and C-LIKE-DOMAIN of IgSF other than IG and TR)
  - G for “groove” domain (G-DOMAIN of MH and G-LIKE-DOMAIN of MhSF other than MH)
5. Locate the “Species” drop-down list, and select a Species.

*If the selection is “All species,” the IMGT/DomainGapAlign tool will propose the result(s), taking into account the best Smith and Waterman scores and percentage of identity. If the selection is forced with a species, such as “Homo sapiens,” the IMGT/DomainGapAlign tool will propose the result(s), taking into account the species, the best Smith and Waterman score(s), and percentage of identity.*
6. Locate the “Displayed alignments” drop-down list and select the number of alignments to display (by default 3). Tick off the checkbox if you want to display the IMGT Colliers de Perles.
7. Check the “Options” section if you would like to modify the parameters.

*The alignment is performed by a modified Smith-Waterman algorithm with an asymmetrical management of insertions between the two sequences. By default, the gap penalty is –5 for the user sequence and –20 for the reference sequence.*
8. Press the “Align and IMGT-gap my sequence(s)” button to launch the analysis.

*This will return the results page.*

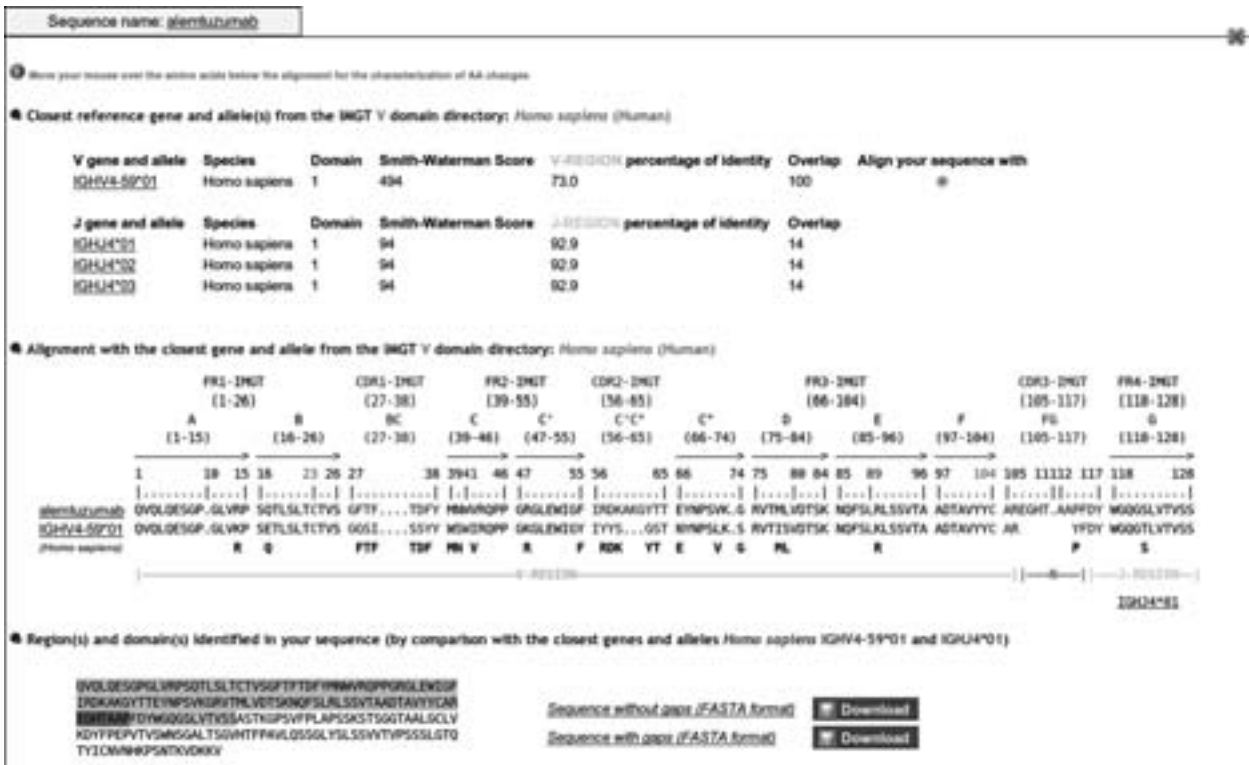
## V Domain Analysis Using IMGT/DomainGapAlign

9. Follow Steps 1–8 above, selecting “V” in Step 4.

*The results display is similar for a V-DOMAIN or a V-LIKE-DOMAIN, despite the huge difference in term of biosynthesis. This strengthens the performance of the tool that respects the structural similarities, while taking into account, for a V-DOMAIN, the particularities of the IG and TR rearrangement and delimitations of the V-(D)-J-REGION.*

The results page for a V-DOMAIN shows the following.

- i. At the top (Fig. 2):
  - The sequence name (as provided by the user).
  - The Closest reference gene and allele(s) from the IMGT domain directory (gene and allele name, Species, Domain number, Smith-Waterman Score, V-REGION percentage of identity, and Overlap score). Note that if several closest alleles are identified (same percentage of identity) or if several alignments to display are chosen, the user can select the display of each corresponding alignment.
  - The Alignment(s) with the domain of the closest gene and allele from the IMGT domain directory.
  - The Region(s) and domain(s) identified in your sequence (by comparison with the closest genes and alleles) using IMGT color menu.
  - Sequence with or without gaps in FASTA format (HTML page or downloadable).
- ii. Then, the Results summary (by comparison with the closest gene and allele) (Fig. 3):
  - Sequence name, V-REGION identity percentage, CDR-IMGT lengths, Number of different AA in CDR1- and CDR2-IMGT, FR-IMGT lengths, Number of different AA in FR-IMGT, Total number of AA changes in V-DOMAIN.
  - The characteristics of the amino acid changes in FR-IMGT (FR1–FR4) and CDR-IMGT (CDR1 and CDR2) (Lefranc 1999, 2011e; Lefranc et al. 2003), according to the IMGT physico-chemical classes (Pommié et al. 2004). Note that the CDR3 that is not relevant for that analysis is not included.



**FIGURE 2.** IMGT/DomainGapAlign top of the results page for a V-DOMAIN. The user V-DOMAIN AA sequence is aligned with the closest germline V-REGION and J-REGION, with IMGT gaps and delimitations of the FR-IMGT and CDR-IMGT according to the IMGT unique numbering (Lefranc et al. 2003). In this example, the user sequence is the V-DOMAIN of the heavy chain (VH) of the monoclonal antibody alemtuzumab. The V-REGION and J-REGION of the alemtuzumab VH is identified as having 73% and 92.9% identity with the *Homo sapiens* IGHV4-59\*01 and IGHJ4\*01, respectively.

Results summary (by comparison with the closest genes and alleles IGHV4-59\*01 and IGHJ4\*01)

Sequence name	V-REGION identity percentage	CDR-IMGT lengths	Number of different AA in CDR1- and CDR2-IMGT	FR-IMGT lengths	Number of different AA in FR-IMGT	Total number of AA changes in V-DOMAIN
alemtuzumab	73.0%	[8, 10, 12]	11	[25, 17, 38, 11] = 91 AA	14	25

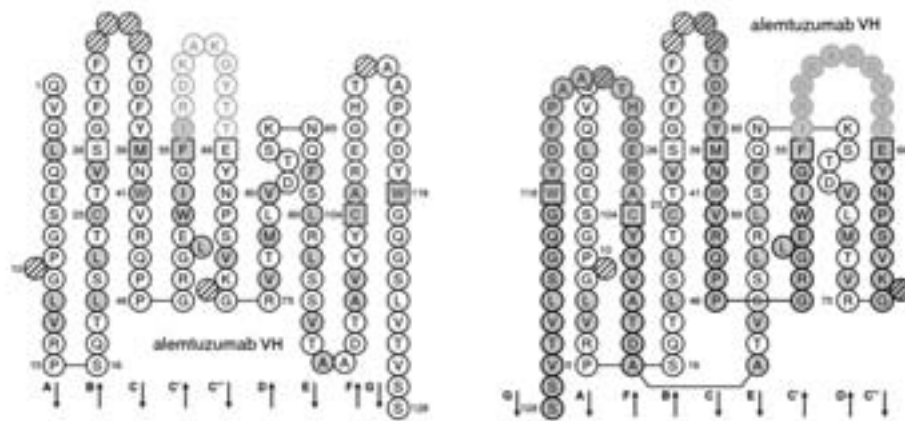
CDR-IMGT	Number of different AA	AA changes
CDR1-IMGT (27-38)	6	G28>F (- - -) very dissimilar S29>T (* - +) similar D30>F (* - -) dissimilar G33>T (* - +) similar S36>D (- - -) very dissimilar Y37>F (- * -) dissimilar
CDR2-IMGT (56-65)	5	Y57>R (- - -) very dissimilar Y58>D (- - -) very dissimilar S59>K (- - -) very dissimilar G63>Y (* - -) dissimilar S64>T (* - +) similar

FR-IMGT	Number of different AA	AA changes
FR1-IMGT (1-26)	2	K14>R (* * *) very similar E17>Q (* * -) similar
FR2-IMGT (39-55)	5	W39>M (* - -) dissimilar S40>N (- - -) very dissimilar I42>V (* - +) similar K48>R (* * +) very similar Y53>F (- * -) dissimilar
FR3-IMGT (66-104)	6	N66>E (* - -) dissimilar L71>V (* - +) similar S74>G (* * -) similar I75>M (* * -) similar S79>L (- - -) very dissimilar K90>R (* * *) very similar
FR4-IMGT (118-129)	1	T123>G (* - +) similar

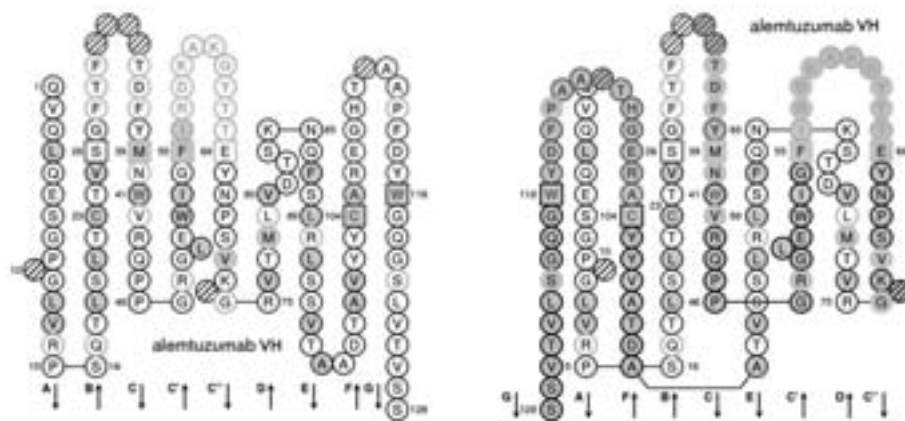
**FIGURE 3.** IMGT/DomainGapAlign Results summary for a V-DOMAIN.

■ IMGT Colliers de Perles for alemtuzumab

● On one and two layers



● On one and two layers with AA changes



**FIGURE 4.** IMGT/DomainGapAlign IMGT Colliers de Perles for a V-DOMAIN. IMGT Collier de Perles on one and two layers are on the *left* and *right* side of the figure, respectively. A display of amino acid changes (pink border) is shown in the *bottom* section of the figure. Positions in blue mean that the amino acid of the user sequence at these positions is hydrophobic (hydropathy index with positive value) or is a tryptophan (W), like in 50% or more of analyzed V domains. Positions with red and bold letters indicate the five conserved positions of a V domain: 1st-CYS 23, CONSERVED-TRP 41, hydrophobic 89, 2nd-CYS 104, and J-TRP 118. Anchor positions are in squares. Hatched positions correspond to gaps according to the IMGT unique numbering for V domain (Lefranc 1997, 1999; Lefranc et al. 2003). Prolines are shown in yellow. Arrows indicate the  $\beta$  strands and their direction. The CDR-IMGT lengths of this domain are [8.10.12]. IMGT/DomainGapAlign identified the domain as a VH, and therefore used the corresponding IMGT color menu for CDR-IMGT: red (CDR1-IMGT), orange (CDR2-IMGT), and purple (CDR3-IMGT).

- iii. If selected in Step 6, IMGT Colliers de Perles on one and two layers, and with or without AA changes (Fig. 4).

*The results page for a V-LIKE-DOMAIN shows a similar display. As an example, Figure 5 shows the top of the results page for the human CTLA4 aligned with the mouse CTLA4 (selection of "Mus musculus" in Step 5). The main difference is that the percentage of identity is calculated for the complete V domain (instead of being calculated for the V-REGION in the case of a V-DOMAIN).*

## C Domain Analysis Using IMGT/DomainGapAlign

- 10. Follow Steps 1–8 above, selecting "C" in Step 4.

*The results page for a C domain is similar for a C-DOMAIN or a C-LIKE-DOMAIN.*

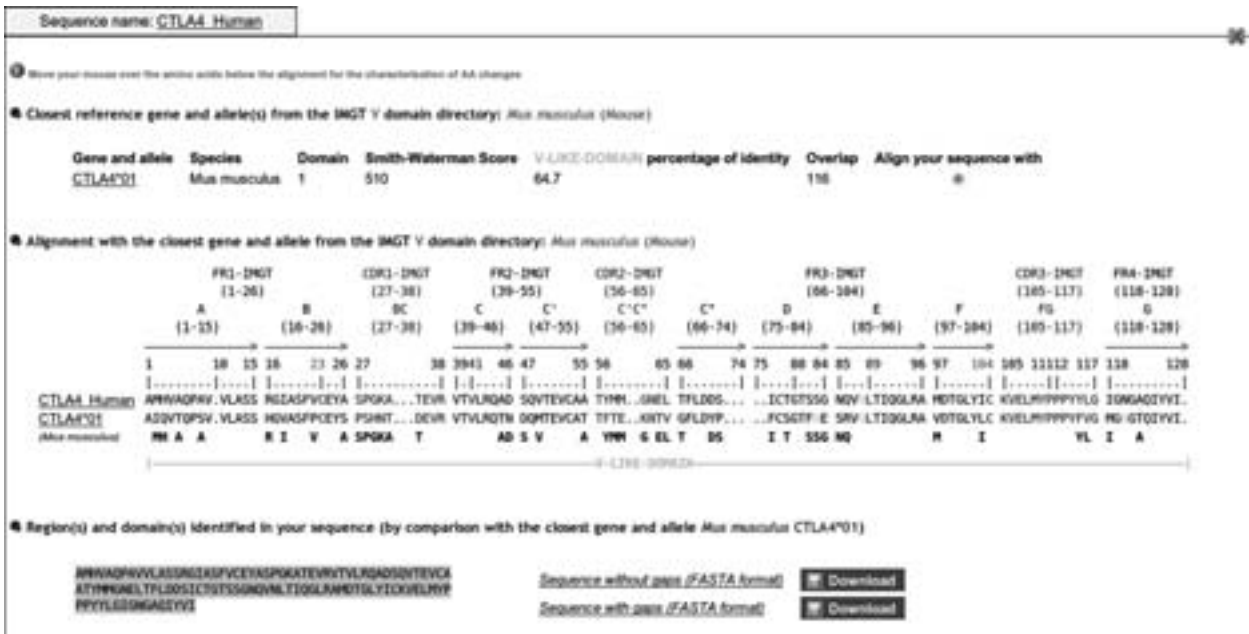


FIGURE 5. IMGT/DomainGapAlign top of the results page for a V-LIKE-DOMAIN.

The results page shows the following.

i. At the top (Fig. 6):

- The sequence name (as provided by the user).
- The Closest reference gene and allele(s) from the IMGT domain directory (gene and allele name, Species, Domain number, percentage of identity, C-DOMAIN, or C-LIKE-DOMAIN

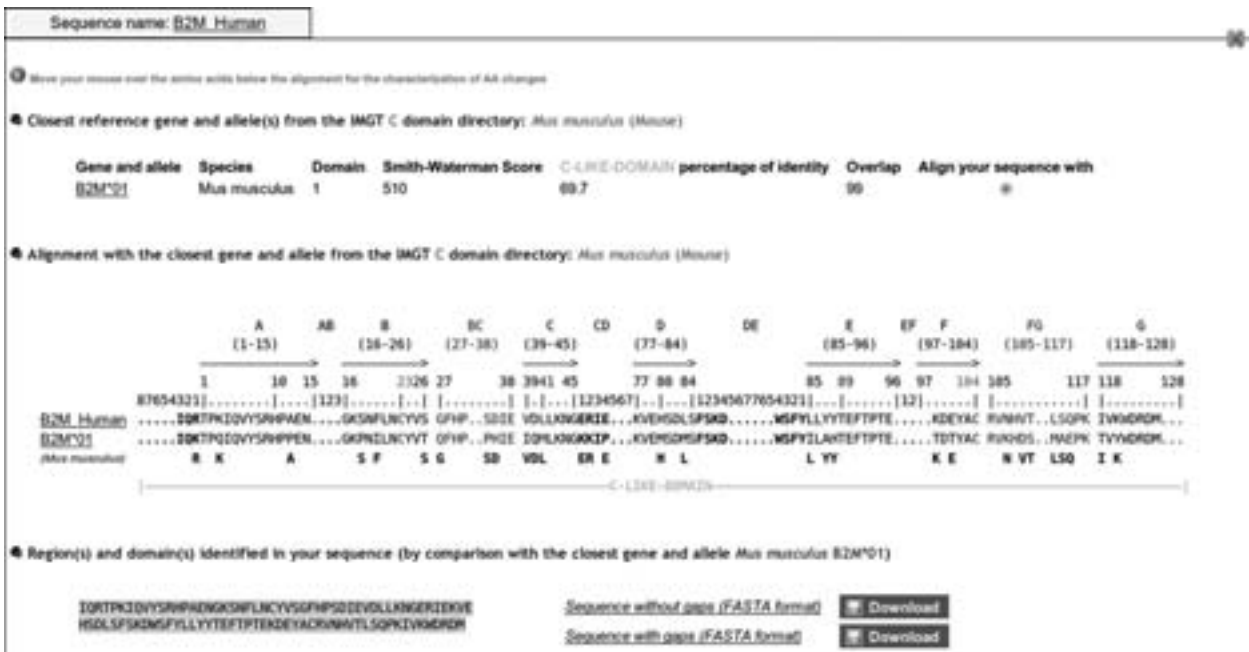


FIGURE 6. IMGT/DomainGapAlign top of the results page for a C type domain. In this example, the user sequence is the *Homo sapiens*  $\beta$ -2-microglobulin (B2M) domain. The B2M domain (*Homo sapiens*) is identified as having 69.7% identity with the *Mus musculus* B2M.

percentage of identity and Overlap score). Note that if several closest alleles are identified (same Smith and Waterman score) or if several alignments to display are chosen, the user can select the display of each corresponding alignment.

- The Alignment(s) with the domain of the closest gene and allele from the IMGT domain directory.
- The C-DOMAIN or C-LIKE-DOMAIN identified in your sequence (by comparison with the closest genes and alleles) colored according to IMGT color menu.
- Sequence with or without gaps in FASTA format (HTML page or downloadable).

ii. Then, the Results summary (by comparison with the closest gene and allele) (Fig. 7):

- Sequence name, C-DOMAIN, or C-LIKE-DOMAIN identity percentage, Total number of AA changes.
- The characteristics of the amino acid changes in strands (A, B, C, CD, D, E, F, G), turns (AB, DE, EF) and loops (BC, FG) (Lefranc et al. 2005b; Lefranc 2011e) according to the IMGT physicochemical classes (Pommié et al. 2004).

iii. If selected in Step 6, IMGT Colliers de Perles on one and two layers, and with or without amino acid changes (Fig. 8).

### G Domain Analysis Using IMGT/DomainGapAlign

11. Follow Steps 1–8 above, selecting “G” in Step 4.

*The results page for a G domain is similar for a G-DOMAIN or a G-LIKE-DOMAIN.*

■ Results summary (by comparison with the closest gene and allele B2M\*01)

Sequence name	C-LIKE-DOMAIN identity percentage	Total number of AA changes in C-LIKE-DOMAIN
B2M_Human	69.7%	30

Strands	Number of different AA	AA changes
A strand (1,9-1,1,1-15)	3	K1,1>R (* + +) very similar Q9>K (* - -) dissimilar P12>A (- - -) very dissimilar
B strand (16-26)	3	P16>S (* - -) dissimilar Q3>F (* - -) dissimilar T26>S (* - +) similar
C strand (39-45)	3	I39>V (* - *) similar Q45>D (* - -) dissimilar M41>L (* + -) similar
CD strand (45,1-45,7)	3	K45,1>E (* - -) dissimilar K45,2>R (* + +) very similar P45,4>E (- - -) very dissimilar
D strand (77-84)	2	M80>H (- - -) very dissimilar M83>L (* + -) similar
E strand (85-96)	3	I85>L (* + *) very similar A87>Y (- - -) very dissimilar H88>Y (* - -) dissimilar
F strand (97-104)	2	T99>K (- - -) very dissimilar T101>E (- - -) very dissimilar
G strand (118-128)	2	T118>I (- - -) very dissimilar Y120>K (- - -) very dissimilar

Turns	Number of different AA	AA changes
AB turn (15,1-15,3)	0	-
DE turn (84,1-84,7, 85,7-85,1)	0	-
EF turn (96,1-96,2)	0	-

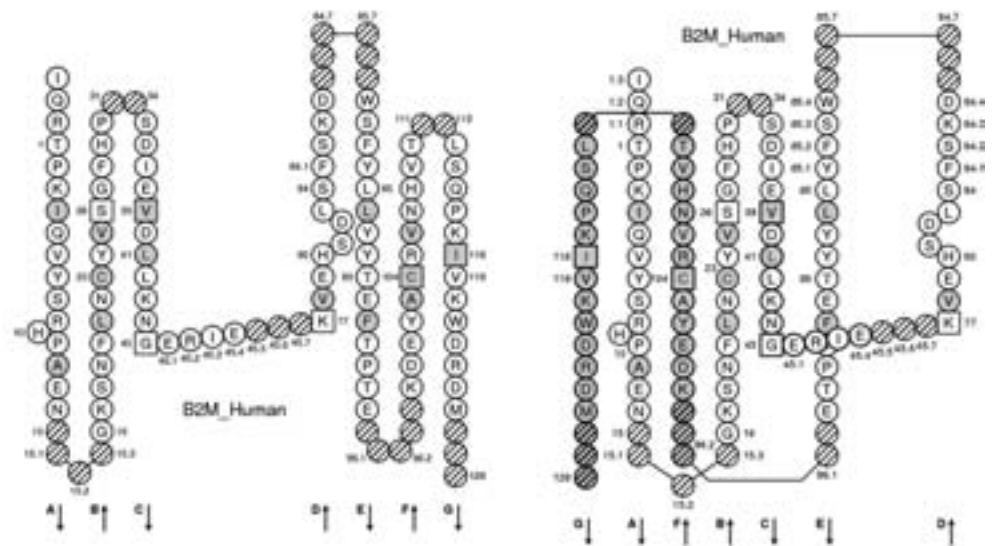
  

Loops	Number of different AA	AA changes
BC loop (27-36)	3	Q27>G (- - -) very dissimilar P33>S (* - -) dissimilar H34>D (- - -) very dissimilar
FG loop (105-117,111,1-111,6,112,1-112,6)	6	K107>N (* - -) dissimilar D109>V (- - -) very dissimilar S110>T (* - *) similar M113>L (* + -) similar A114>S (- + -) dissimilar E115>Q (* + -) similar

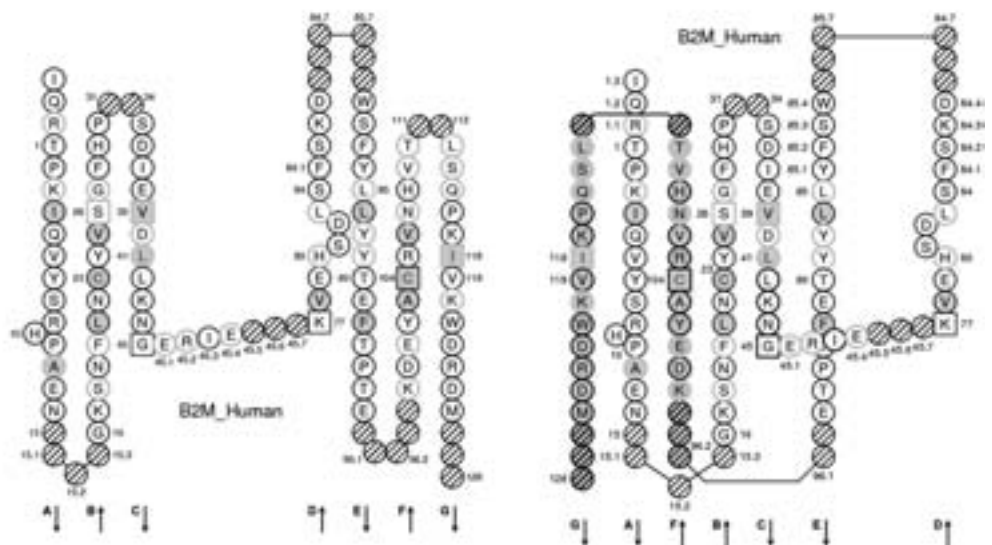
FIGURE 7. IMGT/DomainGapAlign Results summary for a C-LIKE-DOMAIN.



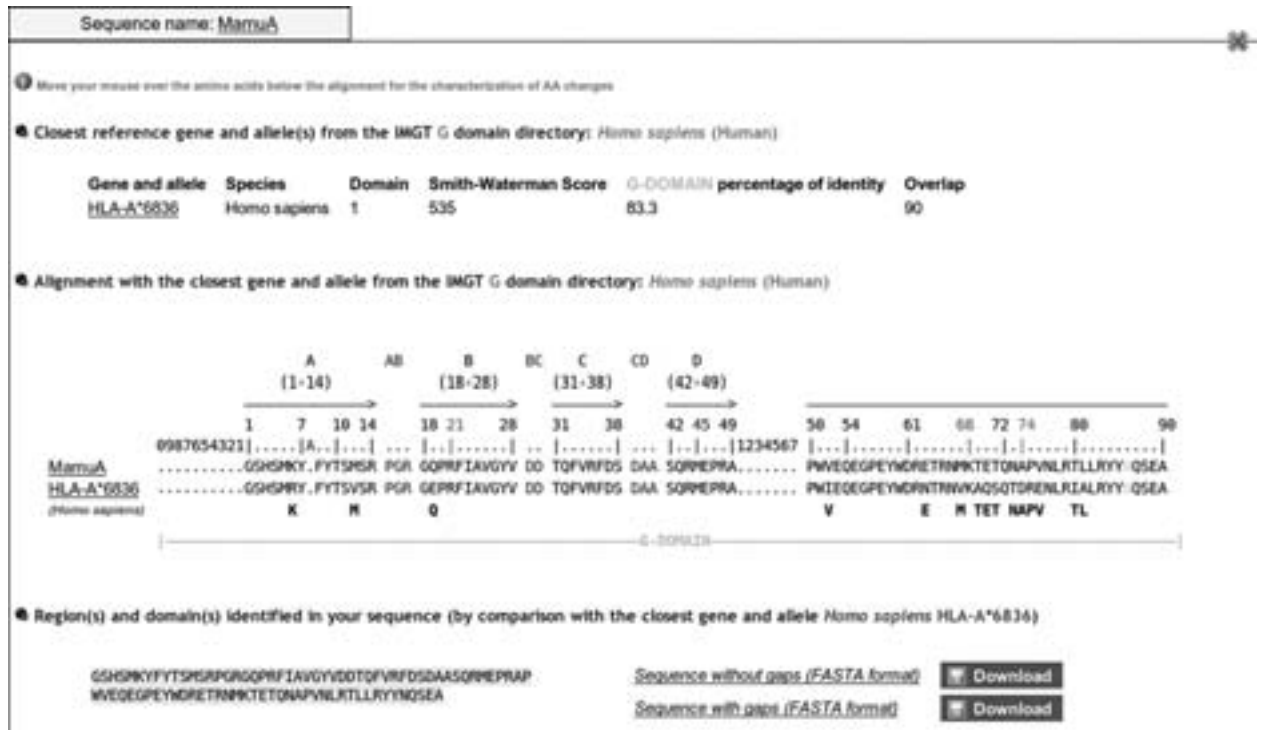
On one and two layers



On one and two layers with AA changes



**FIGURE 8.** IMGT/DomainGapAlign IMGT Colliers de Perles for a C-LIKE-DOMAIN. IMGT Colliers de Perles on one and two layers are on the *left* and *right* side of the figure, respectively. A display of amino acid changes (pink border) is shown in the *bottom* section of the figure. Positions in blue mean that the amino acid of the user sequence at these positions is hydrophobic (hydropathy index with positive value) or is a tryptophan (W), like in 50% or more of analyzed C domains. Positions with red and bold letters are by analogy with a V domain. In a C domain they correspond to the two conserved cysteines, 1st-CYS 23 and 2nd-CYS 104, and to the usually hydrophobic positions 41 and 89 (here 89 is occupied by tyrosine, a neutral amino acid) (Pommié et al. 2004), whereas position 118 is not conserved. Anchor positions are in squares. Hatched positions correspond to gaps according to the IMGT unique numbering for C domain (Lefranc et al. 2005b). Prolines are shown in yellow. Arrows indicate the  $\beta$  strands and their direction.



**FIGURE 9.** IMGT/DomainGapAlign *top* of the results page for a G domain, G-ALPHA1. G-ALPHA1 is the first domain of an MH1 chain. In this example, the user sequence is the G-ALPHA1 of MamuA (Macmul MH1-A) of the rhesus monkey *Macaca mulatta* aligned with the G-ALPHA1 of the closest human HLA-A allele.

The results page shows:

i. At the top (Fig. 9):

- The sequence name (as provided by the user).
- The Closest reference gene and allele(s) from the IMGT domain directory (gene and allele name, Species, Domain number, percentage of identity, G-DOMAIN, or G-LIKE-DOMAIN percentage of identity and Overlap score). Note that if several closest alleles are identified (same Smith and Waterman score) or if several alignments to display are chosen, the user can select the display of each corresponding alignment.
- The Alignment(s) with the domain of the closest gene and allele from the IMGT domain directory.
- The G-DOMAIN or G-LIKE-DOMAIN identified in your sequence (by comparison with the closest genes and alleles) colored according to IMGT color menu.
- Sequence with or without gaps in FASTA format (HTML page or downloadable).

ii. Then, the Results summary (by comparison with the closest gene and allele) (Fig. 10):

- Sequence name, G-DOMAIN or G-LIKE-DOMAIN identity percentage, Total number of AA changes.
- The characteristics of the amino acid changes in strands (A, B, C, D), turns (AB, BC, CD), and in the helix (Lefranc et al. 2005c; Lefranc 2011e), according to the IMGT physico-chemical classes (Pommié et al. 2004).

iii. If selected in Step 6, the IMGT Collier de Perles (Fig. 11).

● Results summary (by comparison with the closest gene and allele HLA-A\*6836)

Sequence name	G-DOMAIN identity percentage	Total number of AA changes in G-DOMAIN
MamuA	83.3%	15

Turns	Number of different AA	AA changes
AB turn (15-17)	0	-
BC turn (29-30)	0	-
CD turn (39-41)	0	-

Strands	Number of different AA	AA changes
A strand (1.9-1.1,1-14)	2	R6>K (+ + +) very similar V13>M (+ - -) dissimilar
B strand (18-28)	1	E20>Q (+ + -) similar
C strand (31-38)	0	-
D strand (42-49,49.1-49.5)	0	-

Helix	Number of different AA	AA changes
Helix (50-92)	12	I53>V (+ - +) similar
		N62>E (+ - -) dissimilar
		V66>M (+ - -) dissimilar
		A68>T (- - -) very dissimilar
		Q69>E (+ + -) similar
		S70>T (+ - +) similar
		T72>N (- + -) dissimilar
		D72>A (- - -) very dissimilar
		R73>P (- - -) very dissimilar
		E74>V (- + -) dissimilar
I78>T (- - -) very dissimilar		
A79>L (+ - +) similar		

FIGURE 10. IMGT/DomainGapAlign Results summary for a G domain, G-ALPHA1.

● IMGT Colliers de Perles for MamuA (determined by IMGT/DomainGapAlign as G-ALPHA1)

● On one layer

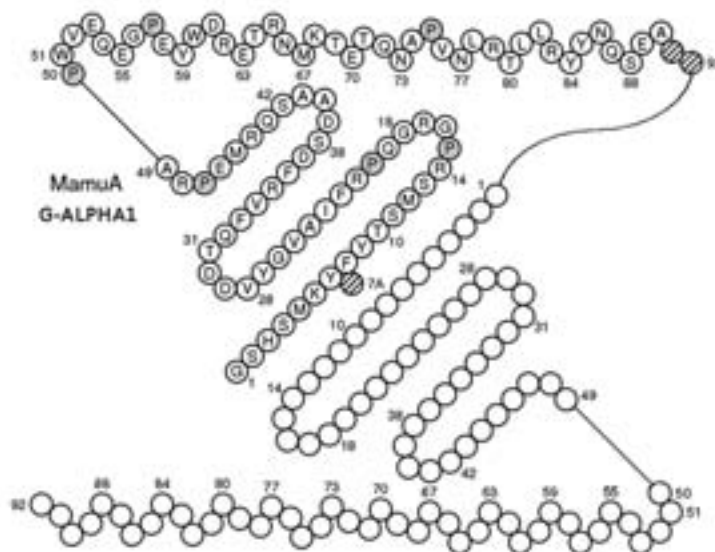
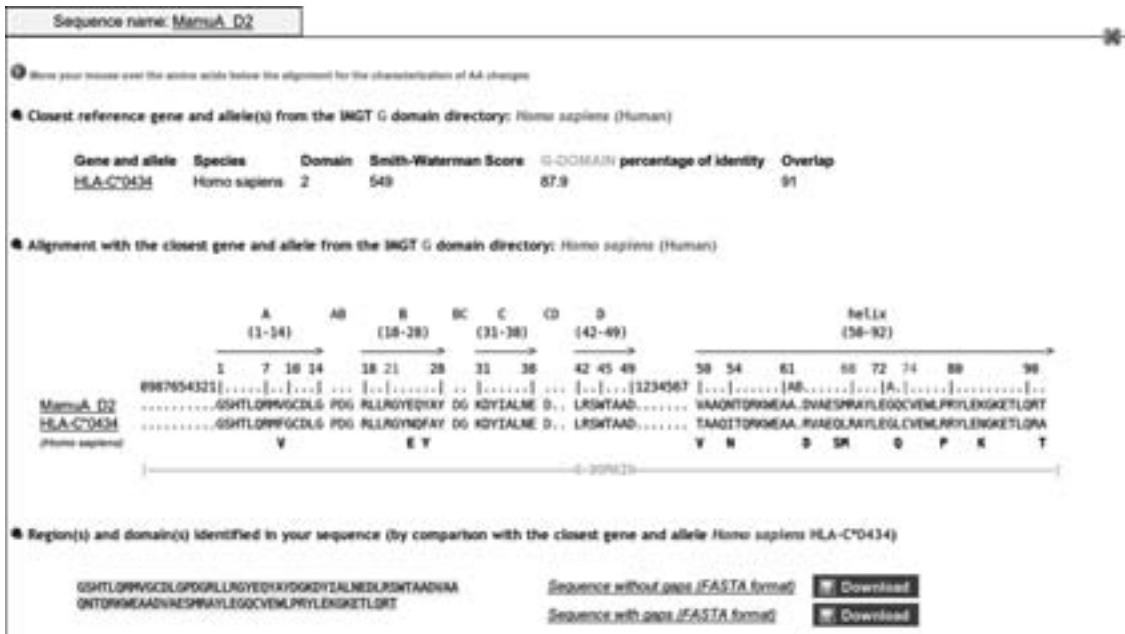


FIGURE 11. IMGT/DomainGapAlign IMGT Collier de Perles for a G domain, G-ALPHA1. A G-ALPHA1 is automatically located in the first half of the figure (for an MH2, the domain automatically displayed in the first half of the figure is G-ALPHA). Hatched positions correspond to gaps according to the IMGT unique numbering for G domain (Lefranc et al. 2005c). Prolines are shown in yellow.



**FIGURE 12.** IMGT/DomainGapAlign top of the results page for a G domain, G-ALPHA2. G-ALPHA2 is the second domain of an MH1 chain. In this example, the user sequence is the G-ALPHA2 of MamuA (Macmul MH1-A) of the rhesus monkey *Macaca mulatta* aligned with the G-ALPHA2 of the closest human HLA-C allele.

Sequence analysis results are similar regardless of the D domain description (G-ALPHA1, G-ALPHA2, G-ALPHA, G-BETA, G-ALPHA1-LIKE, G-ALPHA2-LIKE). Thus, Figure 12 and Figure 13 (G-ALPHA2) are similar to Figure 9 and Figure 10 (G-ALPHA1), respectively. Depending on the G domain description determined by IMGT/DomainGapAlign, the IMGT Collier de Perles is automatically located in the top part for G-ALPHA1 (Fig. 11), G-ALPHA and G-ALPHA1-LIKE (data not shown), or in the bottom part for G-ALPHA2 (Fig. 14), G-BETA, and G-ALPHA2-LIKE (data not shown).

Results summary (by comparison with the closest gene and allele HLA-C\*0434)

Sequence name	G-DOMAIN identity percentage	Total number of AA changes in G-DOMAIN
MamuA_D2	87.9%	11

Turns	Number of different AA	AA changes
AB turn (15-17)	0	-
BC turn (29-30)	0	-
CD turn (39-41)	0	-

Strands	Number of different AA	AA changes
A strand (1-9-1.1,1-14)	1	F9>V (* -) dissimilar
B strand (18-28)	2	N24>E (* -) dissimilar F26>Y (- * -) dissimilar
C strand (31-38)	0	-
D strand (42-49,49.1-49.5)	0	-

Helix	Number of different AA	AA changes
Helix (50-92)	8	T50>V (- -) very dissimilar I54>N (- -) very dissimilar R62>D (* -) dissimilar Q66>S (- -) very dissimilar L67>M (* +) similar L73>Q (- -) very dissimilar R79>P (- -) very dissimilar N84>K (* -) dissimilar

**FIGURE 13.** IMGT/DomainGapAlign Results summary for a G domain, G-ALPHA2.





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# IMGT/3Dstructure-DB: Querying the IMGT Database for 3D Structures in Immunology and Immunoinformatics (IG or Antibodies, TR, MH, RPI, and FPIA)

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## INTRODUCTION

IMGT/3Dstructure-DB is the three-dimensional (3D) structure database of IMGT, the international ImMunoGenetics information system that is acknowledged as the global reference in immunogenetics and immunoinformatics. IMGT/3Dstructure-DB contains 3D structures of (1) antigen receptors that comprise immunoglobulins (IG) or antibodies and T cell receptors (TR), (2) major histocompatibility (MH) proteins of class I (MH1) and class II (MH2), (3) peptide/MH (pMH) complexes (pMH1, pMH2), (4) antigen receptor/antigen complexes (IG/Ag, TR/pMH), and (5) related proteins of the immune system (RPI) belonging to the immunoglobulin and MH superfamilies (IgSF and MhSF, respectively) and found in complexes with IG, TR, or MH. IMGT/3Dstructure-DB data are annotated according to the IMGT criteria, using IMGT/DomainGapAlign, and based on the IMGT-ONTOLOGY axioms and concepts. The use of IMGT/3Dstructure-DB is described here. It provides IMGT gene and allele identification (CLASSIFICATION), region and domain delimitations (DESCRIPTION), and amino acid (AA) positions according to the IMGT unique numbering (NUMEROTATION). IMGT/3Dstructure-DB provides the closest genes and alleles that are expressed in the AA sequences of the 3D structures, by aligning these sequences with the IMGT domain reference directory.

## RELATED INFORMATION

IMGT/3Dstructure-DB (Kaas et al. 2004; Ehrenmann et al. 2010a) is the 3D structure database of IMGT, <http://www.imgt.org> (Lefranc et al. 2009). Standardization and IMGT Scientific chart rules are based on the IMGT-ONTOLOGY concepts of identification, classification, description (Giudicelli and Lefranc 1999; Lefranc et al. 2004, 2005a, 2008) and numerotation (Lefranc 1997, 1999; Lefranc et al. 2003, 2005b,c) generated from the axioms of the Formal IMGT-ONTOLOGY or IMGT-Kaleidoscope (Duroux et al. 2008). The concepts of identification led to the IMGT standardized keywords, the concepts of classification to the IMGT standardized gene and allele names, the concepts of description to the IMGT standardized labels, and the concepts of numerotation to the IMGT unique numbering.

A detailed description of IMGT is provided in **IMGT, the International ImMunoGeneTics Information System** (Lefranc 2011a). Information is also available in **From IMGT-ONTOLOGY IDENTIFICATION Axiom to IMGT Standardized Keywords: For Immunoglobulins (IG), T Cell Receptors (TR), and Conventional Genes** (Lefranc 2011b), **From IMGT-ONTOLOGY DESCRIPTION Axiom to IMGT Standardized Labels: For Immunoglobulin (IG) and T Cell Receptor (TR) Sequences and Structures** (Lefranc 2011c), **From IMGT-ONTOLOGY CLASSIFICATION Axiom to IMGT Standardized Gene and Allele Nomenclature: For Immunoglobulins (IG) and T Cell Receptors (TR)** (Lefranc 2011d), **IMGT Unique Numbering for the Variable (V), Constant (C), and Groove (G) Domains of IG, TR, MH, IgSF, and MhSF** (Lefranc 2011e), and **IMGT Collier de Perles for the Variable (V), Constant (C), and Groove (G) Domains of IG, TR, MH, IgSF, and MhSF** (Lefranc 2011f).

In addition, protocols are available for **IMGT/V-QUEST: IMGT Standardized Analysis of the Immunoglobulin (IG) and T Cell Receptor (TR) Nucleotide Sequences** (Giudicelli et al. 2011), **IMGT/JunctionAnalysis: IMGT Standardized Analysis of the V-J and V-D-J Junctions of the Rearranged**

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Immunoglobulins (IG) and T Cell Receptors (TR) (Giudicelli and Lefranc 2011), IMGT/Collier de Perles: IMGT Standardized Representation of Domains (IG, TR, and IgSF Variable and Constant Domains, MH and MhSF Groove Domains) (Ehrenmann et al. 2011), and IMGT/DomainGapAlign: IMGT Standardized Analysis of Amino Acid Sequences of Variable, Constant, and Groove Domains (IG, TR, MH, IgSF, MhSF) (Ehrenmann and Lefranc 2011).

## MATERIALS

It is essential that you consult the appropriate Material Safety Data Sheets and your institution's Environmental Health and Safety Office for proper handling of equipment and hazardous materials used in this protocol.

### Equipment

Computer (Internet-connected).

## METHOD

### Querying the IMGT/3Dstructure-DB Database

1. Using any modern web browser, go to the IMGT Home page <http://www.imgt.org>.
2. Click the "IMGT/3Dstructure-DB" link in the IMGT databases section of the IMGT Home page.

*This will take you to the IMGT/3Dstructure-DB database web application (Fig. 1).*

*Six sections are available in the IMGT/3Dstructure-DB query page: "Search by Entry code or Molecule name (receptor or ligand)," "Search for complexes," "Search by IMGT entry type using IMGT-ONTOLOGY concepts," "Search by Resolution, Release date or Experimental method," "Search by bibliographical references," and "Chain alignment." These sections are independent of each other, and thus, each user query only concerns one of these six sections. Choose*

**WELCOME !**  
**to IMGT/3Dstructure-DB**  
THE INTERNATIONAL IMMUNOGENETICS INFORMATION SYSTEMS

Im Muno Gene Tics Information system  
<http://www.imgt.org>

**IMGT/3Dstructure-DB Query page**

Citing IMGT/3Dstructure-DB : Kaas, G. et al. Nucleic Acids Res., 32, D208 - D210 (2004) PMID: 14501395 Abstract Full text

Today is Friday, July 16 2010  
IMGT/3Dstructure-DB contains 2300 entries  
1071 - IGH  
04 - IGH  
335 - Kabat

Program version: 3.3.0 (06/07/2010) - Database release: 20100714 (06/07/2010)

● Search by Entry code or Molecule name (receptor or ligand)

● Entry code (IG, TR, MH, MhSF)  ● Molecule name (receptor or ligand)

Search Reset

● Search for complexes

Paratope/epitope ● IG/Ag ● TRpMH1 ● TRpMH2 Peptide/MH ● pMH1 ● pMH2

Ligand category  Peptide length

Search Reset

FIGURE 1. IMGT/3Dstructure-DB Welcome and Query page.

selection A (Steps 3–5), B (Steps 6–9), C (Steps 10–13), D (Steps 14–17), or E (Steps 18–21) and then continue to Step 24, or selection F (Steps 22–23).

#### **Selection A: Search by Entry Code or Molecule Name**

3. Locate the “Entry code” or the “Molecule name (receptor or ligand)” text areas.
4. Enter an entry code (PDB, INN, PROTEIN) in “Entry code” text area or a Molecule name (receptor or ligand) in “Molecule name” text area.

*The IMGT/3Dstructure-DB entry code (ID) is identical to either the PDB code, INN code, or PROTEIN code. PDB code is an alphanumeric code (ex: 1ao7), INN code is a numeric code (ex: 7637), while PROTEIN code is an alphanumeric code (ex : p00054). An AutoComplete behavior is available for this field. AutoComplete works in the following manner: as you type in the beginning of a PDB code, previous entries are pulled from a storage area and you may elect to simply select one of these entries. By selecting an entry, you no longer have to input the string because AutoComplete finishes entering the string for you.*

*An IMGT molecule name is a name retrieved from the literature, a name modified by IMGT, or a name added by IMGT. The search by IMGT molecule name is case insensitive.*

5. Proceed to Step 24.

#### **Selection B: Search for Complexes**

6. Locate the “Paratope/epitope” or “Peptide/MH” radio buttons, and select complexes you want to retrieve.

*Search by Paratope/epitope can retrieve entries containing the following complexes:*

*IG/Ag: Immunoglobulin/antigen*

*TR/pMH1: T cell Receptor/peptide/MH1*

*TR/pMH2: T cell Receptor/peptide/MH2*

*Search by Peptide/MH can retrieve entries containing the following complexes:*

*pMH1: peptide/MH1*

*pMH2 peptide/MH2*

7. Locate the “Ligand category” radio button, and select the ligand category for which you want to retrieve the corresponding complexes.

*Search by Ligand category can retrieve entries with ligands that can belong to the following categories: Chemical compound, DNA, Nucleotide, Peptide, Protein, or RNA.*

8. Select a peptide length if needed.

*Selecting a peptide length automatically enters the “Peptide” choice in the “Ligand category” drop-down list.*

9. Proceed to Step 24.

#### **Selection C: Search by IMGT Entry Type Using IMGT-ONTOLOGY Concepts**

10. Locate the “IMGT entry type” radio button, select an entry type (PDB, INN, Kabat, or any) (Fig. 2).

11. Locate the three sections “IDENTIFICATION,” “DESCRIPTION,” and “CLASSIFICATION.”

*Selecting an “IMGT receptor type” (e.g., “IG”) will update the “IMGT receptor description” list, so that it only contains IMGT receptor description labels for the “IG.” In the same way, selecting an “IMGT group” (e.g., “TRAV”) will update the “IMGT subgroup” list, so that it only contains IMGT subgroup(s) from the IMGT “TRAV” group.*

12. Select the criteria as needed:

- i. In “IDENTIFICATION:”

Species

IMGT receptor type (IG, TR, MH, RPI, FPIA, or any)

*Options menu (MH1, MH2, or RPI-MH1Like) allows you to refine your research.*

- ii. In “DESCRIPTION:”

IMGT receptor description: IG, TR, MH, or RPI receptors are described with standardized IMGT labels.

*Options menu (FV, SCFV, FAB, or FC) allows you to refine your research.*

■ Search by IMGT entry type using IMGT-ONTOLOGY concepts

IMGT entry type	<input type="radio"/> PDB <input type="radio"/> INH <input type="radio"/> Kabat <input type="radio"/> any
IDENTIFICATION	Species <input type="text" value="any"/>
	IMGT receptor type <input type="radio"/> IG <input type="radio"/> TR <input type="radio"/> MH <input type="radio"/> RPI <input type="radio"/> FPA <input type="radio"/> any <small>&gt; Options: <input type="radio"/> Min <input type="radio"/> SH2 <input type="radio"/> NP-149TLike</small>
DESCRIPTION	IMGT receptor description <input type="text" value="any"/> <small>&gt; Options: <input type="radio"/> Fv <input type="radio"/> SOFV <input type="radio"/> FAB <input type="radio"/> FC</small>
	IMGT chain description <input type="text" value="any"/>
	IMGT domain description <input type="text" value="any"/>
CLASSIFICATION	IMGT group <input type="text" value="any"/>
	IMGT subgroup <input type="text" value="any"/>
	IMGT gene <input type="text" value="any"/>
	IMGT allele <input type="text"/>

■ Search by Resolution, Release date or Experimental method

Resolution   
 Release date   
 Experimental method

**FIGURE 2.** IMGT/3Dstructure-DB Query page: Search by IMGT entry type using IMGT-ONTOLOGY concepts.

IMGT chain description: IMGT IG, TR, MH, or RPI chains are described with standardized IMGT labels (Lefranc 2010c).

iii. In “CLASSIFICATION:”

IMGT group  
 IMGT subgroup  
 IMGT gene  
 IMGT allele

*Human IG (Lefranc and Lefranc 2001a) and TR (Lefranc and Lefranc 2001b) genes and alleles are from IMGT/GENE-DB (Giudicelli et al. 2005) with translation in IMGT/DomainDisplay (Ehrenmann et al. 2010b). Genes and alleles from other species and/or other proteins are added when the genome of these species is annotated and nomenclature can be set up (Lefranc 2010d), and/or when 3D structures are available in IMGT/3Dstructure-DB.*

13. Proceed to Step 24.

**Selection D: Search by Resolution, Release Date, or Experimental Method**

14. Locate the “Resolution” radio button (Fig. 2), and select resolution for which you want to retrieve structures (only for PDB entries).

*Six value ranges are available: 0–1.00 Å, 1.01–2.00 Å, 2.01–3.00 Å, 3.01–4.00 Å, 4.01–5.00 Å, and 5.01–6.00 Å.*

15. Locate the “Release date” radio button, and select PDB release date for which you want to retrieve structures (only for PDB entries).

16. Locate the “Experimental method” radio button, and select PDB experimental method for which you want to retrieve structures (only for PDB entries).

*Users can choose among nine experimental methods: Electron microscopy; NMR; NMR, 18 structures; NMR, 20 structures; Solution scattering; Solution scattering, theoretical model; Theoretical model; X-ray diffraction; or X-ray diffraction, single crystal.*

17. Proceed to Step 24.



The image shows two sections of a web interface. The top section, titled "Search by bibliographical references", has a radio button for "PDB" selected and "OR" selected between "PDB" and "PubMed". Below this are four drop-down menus for "Authors", "Journal", "Year", and "PMID". To the right is a text input field labeled "Title (part of)". At the bottom right of this section are "Search" and "Reset" buttons. The bottom section, titled "Chain alignment", has a radio button for "Align your sequence (FASTA format)" selected. Below this is a large text input field. At the bottom of this section are "E-value" (set to 0.0001) and "Number of results" (set to 10) dropdown menus, and "Align" and "Reset" buttons.

FIGURE 3. IMGT/3Dstructure-DB Query page: Search by bibliographical references, and Chain alignment.

#### Selection E: Search by Bibliographical References

18. Locate the section "Search by bibliographical references" (Fig. 3) and select the type of reference: PDB or PubMed.
19. Then, locate the drop-down lists below and select an author and/or a journal and/or a year and/or a PMID (for PubMed references).  
*Again, this kind of search uses linked select elements. For example, selecting an author will update the "Journal" drop-down list to only contain Journal(s) from the author selected.*
20. The research can also be performed on the title of publications (part of).
21. Proceed to Step 24.

#### Selection F: Chain Alignment

22. Locate the "Align your sequence" radio button (Fig. 3), click on it, and paste your sequence in FASTA format in the text area.  
*Your own sequence will be aligned using FASTA (Pearson and Lipman 1988) against the IMGT/3Dstructure-DB sequences with the "FASTA chain alignment" tool.*  
*The E-value can be modified and users can choose the numbers of results to display. For instance, if there are no results in "Chain Alignment results," you may try changing the E-value from 0.0001 to 0.001. Results can also be obtained with peptides of 11 AA (or even less) with an E-value of 0.5.*
23. Click on Align. This will give you the Chain alignment results.

#### IMGT/3Dstructure-DB Visualization Choice for the Results

24. Locate the "Display results" section at the bottom of the IMGT/3Dstructure-DB Query page (Fig. 4).  
*There are three possible results visualization (select by clicking on a radio button):*
  - "Overview" (selected by default)
  - "Domain type sequences"
  - "FR-IMGT/CDR-IMGT sequences and lengths" (only for V domains)
25. Select, if needed, the type of display and the corresponding views.
26. Click on Search.

6 Display results



FIGURE 4. IMGT/3Dstructure-DB Query page: Display results section (for choice of the results visualization).

### IMGT/3Dstructure-DB Display Results

27. You will get one of the possible visualizations: “Overview” (Fig. 5) (by default or after selection of some views), “Domain type sequences” (Fig. 6), or “FR-IMGT and/or CDR-IMGT sequences” (Fig. 7), depending on your selection.

At the top of the results page, the visualization choice and the query are recalled, and the number of results is shown. For results that correspond to a list of chains (and not to a list of entries), it is recalled, between parentheses, that the number of results corresponds to the “number of chains containing the selected domain type.” The list of results is sorted by ID (entry or chain).

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## Overview

Your query: Molecule name (receptor or ligand) HIV

Number of results: **111**

Click on IMGT entry ID (2nd column) for entry card

IMGT entry ID	IMGT molecule name	IMGT entry type	IMGT receptor description	Species	Ligand(s)
1	1aay	38.1 nAb, anti-gp120 [HIV-1]	FAS-GAMMA_1_KAPPA	Mus musculus	Envelope glycoprotein gp120 [Human immunodeficiency virus 1, HIV-1]
2	1ah	23.3 nAb, anti-gp24 [HIV-1]	FAS-GAMMA_1_KAPPA	Mus musculus	Caprip protein p24 (residues: 1-191) [Human immunodeficiency virus 1, HIV-1]
3	1ah	Cb41 nAb, anti-gp24 [HIV-1]	FAS-GAMMA_2C_KAPPA	Mus musculus	Eclipse-related peptide p24 [Human immunodeficiency virus 1, HIV-1]
4	1ah	Cb41 nAb, anti-gp24 [HIV-1]	FAS-GAMMA_2C_KAPPA	Mus musculus	Unrelated D-peptide p24 [Human immunodeficiency virus 1, HIV-1]
5	1ah	Cb41 nAb, anti-gp24 [HIV-1]	FAS-GAMMA_2C_KAPPA	Mus musculus	Unrelated D-peptide p24 [Human immunodeficiency virus 1, HIV-1]
6	1ah	1385 nAb, anti-gp120 [HIV-1]	FAS-GAMMA_1_KAPPA	Mus musculus	Caprip protein p24 (residues: 142-351) [Human immunodeficiency virus 1, HIV-1]
7	1ah	58.2 nAb, anti-gp120 [HIV-1]	FAS-GAMMA_1_KAPPA	Mus musculus	Envelope glycoprotein gp120 [Human immunodeficiency virus 1, HIV-1]
8	1gln	178 nAb, anti-gp120 [HIV-1]	FAS-GAMMA_1_KAPPA	Homio sapiens	Envelope glycoprotein gp120 [Human immunodeficiency virus 1, HIV-1] 2-Propanol, Neopentanol
9	1gln	178 nAb, anti-gp120 [HIV-1]	FAS-GAMMA_1_KAPPA	Homio sapiens	Envelope glycoprotein gp120 [Human immunodeficiency virus 1, HIV-1]
10	1gn1	178 nAb, anti-gp120 [HIV-1]	FAS-GAMMA_1_KAPPA	Homio sapiens	Envelope glycoprotein gp120 [Human immunodeficiency virus 1, HIV-1]
11	1gpp	30.1 nAb, anti-gp120 [HIV-1]	FAS-GAMMA_2A_KAPPA	Mus musculus	Envelope glycoprotein gp120 [Human immunodeficiency virus 1, HIV-1]
12	1hnd	Cb41 nAb, anti-gp24 [HIV-1]	FAS-GAMMA_2C_KAPPA	Mus musculus	Eclipse-related peptide p24 [Human immunodeficiency virus 1, HIV-1]
13	1hnd	Cb41 nAb, anti-gp24 [HIV-1]	FAS-GAMMA_2C_KAPPA	Mus musculus	Eclipse-related peptide p24 [Human immunodeficiency virus 1, HIV-1]
14	1hnd	Cb41 nAb, anti-gp24 [HIV-1]	FAS-GAMMA_2C_KAPPA	Mus musculus	Eclipse-related peptide p24 [Human immunodeficiency virus 1, HIV-1]
15	1lps	Fab-28 nAb, anti-reverse transcriptase [HIV-1]	FAS-GAMMA_1_KAPPA	Mus musculus	Reverse transcriptase (RT) EC: 2.7.7.49 [Human immunodeficiency virus 1, HIV-1]
16	1hsh	512 neutralizing nAb, anti-gp120 [HIV-1]	SI-GAMMA_1_KAPPA	Homio sapiens	
17	1af	Cb41 nAb, anti-gp24 [HIV-1]	FV HEAVY_KAPPA	Mus musculus	Derived peptide p24 [Human immunodeficiency virus 1, HIV-1]
18	12a	Fab25 nAb, anti-reverse transcriptase [HIV-1]	FAS-GAMMA_1_KAPPA	Mus musculus	Polysaccharide (Synthecel) Reverse transcriptase (RT), M184-I, EC:2.7.7.49 [Human immunodeficiency virus 1, HIV-1]
19	12a	1896 nAb, anti-protease [HIV-1]	3CFV HEAVY_KAPPA	Mus musculus	A terminus of protease [Human immunodeficiency virus 1, HIV-1]

FIGURE 5. IMGT/3Dstructure-DB “Overview” display of results. The “Overview” results provide IMGT entry ID, IMGT molecule name, IMGT entry type, IMGT receptor description, Species, Ligand(s), Experimental technique, Resolution, and release date (only for PDB).

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**V domain sequences**

Your query: Species Homo sapiens and Entry type PDB and Receptor type IG



Number of results: **939**

(number of chains containing the selected domain type).

Click on IMGT entry ID (2nd column) for entry card

IMGT chain ID	Species	V type domain												
		FR3-DNGT (1-26)	CDR1-IND1 (27-36)	FR4-DNGT (38-55)	CDR2-DNGT (56-65)	FR5-DNGT (66-104)	CDR3-DNGT (105-117)	FR6-DNGT (118-128)						
		A (1-15)	B (16-26)	BC (27-36)	C (38-48)	C' (47-55)	C'' (56-65)	66-74	D (75-84)	E (85-96)	F (97-104)	105-117	118-128	
1	hAb_H	1	10 15 26	27 28 27	38 39 41 48 47	55 56	65 66	74 75	80 84 85 86	96 97	104	105 111	112 117 118 128	
2	hAb_L	SALTTPPE AGGSL GGNTSEKTTG SDGK...GHR YKYYQQA KAPAVLYZ EV...N NPPSGP D RFGGK...SC NTKLTVSLDA GSEADYYC SYDSS...[129487765432]												SNFQ FGTGKHTL...
3	hAb_H	EVLVSSG GLVDF GRLLSLCVTS GTF...DYS HAVRQAP GGLWVYD I3M...TGT ZYADQK G NTLSPDRAK NGLYQMSLRV EDLALYYC NTROYVY...ANDYYH WGGELVTVSS...												
4	hAb_L	EVLVSSG GLVDF GRLLSLCVTS GTF...DYS HAVRQAP GGLWVYD I3M...TGT ZYADQK G NTLSPDRAK NGLYQMSLRV EDLALYYC NTROYVY...ANDYYH WGGELVTVSS...												
5	hAb_H	EVLVSSG GLVDF GRLLSLCVTS GTF...DYS HAVRQAP GGLWVYD I3M...TGT ZYADQK G NTLSPDRAK NGLYQMSLRV EDLALYYC NTROYVY...ANDYYH WGGELVTVSS...												
6	hAb_L	EVLVSSG GLVDF GRLLSLCVTS GTF...DYS HAVRQAP GGLWVYD I3M...TGT ZYADQK G NTLSPDRAK NGLYQMSLRV EDLALYYC NTROYVY...ANDYYH WGGELVTVSS...												
7	hAb_C	GGRTGPPSLSAVY GSRNTETIQS GSI...DHR LHWYQAP KAPALLEY DA...S NLDGAP S RFGGK...SS TGYFTIISLQF GELATYYC GYDQ...LPTT FGGDTRKSK...												
8	hAb_A	GGRTGPPSLSAVY GSRNTETIQS GSI...DHR LHWYQAP KAPALLEY DA...S NLDGAP S RFGGK...SS TGYFTIISLQF GELATYYC GYDQ...LPTT FGGDTRKSK...												
9	hAb_B	GGRTGPPSLSAVY GSRNTETIQS GSI...DHR LHWYQAP KAPALLEY DA...S NLDGAP S RFGGK...SS TGYFTIISLQF GELATYYC GYDQ...LPTT FGGDTRKSK...												
10	hAb_C	GGRTGPPSLSAVY GSRNTETIQS GSI...DHR LHWYQAP KAPALLEY DA...S NLDGAP S RFGGK...SS TGYFTIISLQF GELATYYC GYDQ...LPTT FGGDTRKSK...												
11	hAb_A	GGRTGPPSLSAVY GSRNTETIQS GSI...DHR LHWYQAP KAPALLEY DA...S NLDGAP S RFGGK...SS TGYFTIISLQF GELATYYC GYDQ...LPTT FGGDTRKSK...												
12	hAb_B	GGRTGPPSLSAVY GSRNTETIQS GSI...DHR LHWYQAP KAPALLEY DA...S NLDGAP S RFGGK...SS TGYFTIISLQF GELATYYC GYDQ...LPTT FGGDTRKSK...												
13	hAb_A	SALTTPPE AGGSL GGNTSEKTTG SDGK...GHR YKYYQQA KAPAVLYZ EV...N NPPSGP D RFGGK...SC NTKLTVSLDA GSEADYYC SYDSS...[129487765432]												SNFQ FGTGKHTL...
14	hAb_B	SALTTPPE AGGSL GGNTSEKTTG SDGK...GHR YKYYQQA KAPAVLYZ EV...N NPPSGP D RFGGK...SC NTKLTVSLDA GSEADYYC SYDSS...[129487765432]												SNFQ FGTGKHTL...
15	hAb_A	SALTTPPE AGGSL GGNTSEKTTG SDGK...GHR YKYYQQA KAPAVLYZ EV...N NPPSGP D RFGGK...SC NTKLTVSLDA GSEADYYC SYDSS...[129487765432]												SNFQ FGTGKHTL...

**FIGURE 6.** IMGT/3Dstructure-DB “Domain type sequences” display of results. This visualization provides the sequences of the domains of the IMGT/3Dstructure-DB chains that satisfy the query.

28. Click on the IMGT entry ID or IMGT chain ID to get access to the individual IMGT 3Dstructure-DB cards.

**IMGT/3Dstructure-DB card-Header**

29. Locate the IMGT/3Dstructure-DB card header. The header comprises the following.

i. The summary table of the entry:

- IMGT molecule name
- IMGT receptor type
- IMGT receptor description
- Ligands
- Species
- Crystallographic complex (CC)
- Chain ID

ii. Below the summary table:

- Experimental technique
- Resolution (in angstroms)
- PDB release date

iii. Seven or eight (if Paratope and epitope are present) tabs that give access to:

- Chain details (open by default; see Step 30).
- Contact analysis (if you click on the tab, proceed to Step 33)
- Paratope and epitope

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**FR-IMGT and/or CDR-IMGT sequences**

Your query: Species Homo sapiens and Entry type PDB and Receptor type IG and IMGT group IGHV



Number of results: **415**

(number of chains containing the selected domain type).

Click on IMGT entry ID (2nd column) for entry card

IMGT chain ID	Species	CDR3-IMGT sequence	CDR3-IMGT length
1	1a2k_H	AKTRSYVVMACYYHY	15
2	1a2k_H	AKTRSYVVMACYYHY	15
3	1a2k_B	AKRFPQGLVSNPFA	14
4	1a2k_D	AKRFPQFTAPNDY	14
5	1a2k_F	AKRFPQFTAPNDY	14
6	1a2k_H	VGRSYVDSGGVYTSNPSK	15
7	1a2k_B	AKRPHDTSQHYWNY	14
8	1a2k_D	AKRPHDTSQHYWNY	14
9	1a2k_F	AKRPHDTSQHYWNY	14
10	1a2k_H	AKRPHDTSQHYWNY	14
11	1a2k_B	AKRPHDTSQHYWNY	14
12	1a2k_H	AKRPHDTSQHYWNY	14
13	1g2k_H	ADVYEGAEDEGEYRNSFLAK	21
14	1g2k_H	ADVYEGAEDEGEYRNSFLAK	21
15	1g2k_H	ADVYEGAEDEGEYRNSFLAK	21
16	1a2k_B	AKRFPQFTAPNDY	14
17	1a2k_D	AKRFPQFTAPNDY	14
18	1a2k_H	AKRPHDTSQHYWNY	14
19	1a2k_H	AKRPHDTSQHYWNY	14
20	1a2k_H	AKRPHDTSQHYWNY	14
21	1a2k_H	AKRPHDTSQHYWNY	14
22	1a2k_H	AKRPHDTSQHYWNY	14
23	1a2k_H	AKRPHDTSQHYWNY	14
24	1a2k_A	CARDPYGGQKSEFDY	15
25	1a2k_B	CARDPYGGQKSEFDY	15

**FIGURE 7.** IMGT/3Dstructure-DB “FR-IMGT and/or CDR-IMGT sequences” display of results. This visualization provides the sequences and lengths of the FR1-IMGT, FR2-IMGT, FR3-IMGT, FR4-IMGT, CDR1-IMGT, CDR2-IMGT, or CDR3-IMGT of the V domains of the IMGT/3Dstructure-DB chains that satisfy the query.

3D visualization (JMOL or QuickPDF; only available for PDB entries)

Renumbered IMGT file (access to the renumbered coordinate file according to the IMGT unique numbering; the “download” link directly downloads the renumbered IMGT file [in gzip format])

INN definitions and properties (only available for INN entries)

IMGT numbering comparison (comparison of the residue numbering between the IMGT file and the PDB file)

Reference and links

Printable card

**IMGT/3Dstructure-DB Card – Chain Details**

30. Locate in the “IMGT/3Dstructure-DB card,” the section “Chain details” (open by default) (Fig. 8) that comprises information on the chain itself, then per domain.

The information for each chain includes:

- Chain ID (for example 1bey\_H)
- Chain length in AA (for example 219)
- IMGT chain description with the delimitations of the different domains (for example VH-CH1 = VH(1–121)[D1] + CH1(122–210)[CH2])
- Chain sequence with delimitations of the regions and domains, highlighting of amino acids that are different from the closest genes and alleles.

The information for each V domain, as an example, includes:

- IMGT domain description with the delimitations of the domain (for example VH(1–121)[D1])

Chain details		
For the IMGT/3Dstructure-DB card of a given residue, click on its letter to a sequence. Differences with the closest IMGT allele sequences are in orange.		
Chain details of <a href="#">IGHV4-59*01</a> (IGHV4-59*01) (Human) (IMGT/3Dstructure-DB) [View   Help]		
Chain ID	1bey_H	
Chain length	219	
IMGT chain description	VH-CH1	
	= VH (1-121) (E1) + CH1 (122-219) (E2)	
Chain sequence	<p>           [ 1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 20 21 22 23 24 25 26 27 28 29 30 31 32 33 34 35 36 37 38 39 40 41 42 43 44 45 46 47 48 49 50 51 52 53 54 55 56 57 58 59 60 61 62 63 64 65 66 67 68 69 70 71 72 73 74 75 76 77 78 79 80 81 82 83 84 85 86 87 88 89 90 91 92 93 94 95 96 97 98 99 100 101 102 103 104 105 106 107 108 109 110 111 112 113 114 115 116 117 118 119 120 121 122 123 124 125 126 127 128 129 130 131 132 133 134 135 136 137 138 139 140 141 142 143 144 145 146 147 148 149 150 151 152 153 154 155 156 157 158 159 160 161 162 163 164 165 166 167 168 169 170 171 172 173 174 175 176 177 178 179 180 181 182 183 184 185 186 187 188 189 190 191 192 193 194 195 196 197 198 199 200 201 202 203 204 205 206 207 208 209 210 211 212 213 214 215 216 217 218 219 ]         </p> <p>           DVLGDSPLKIPPSCLSLTCTYSQTTFYHNRQPPQGLNGDIDKAAITTTMPSKAVTFLKTSANPFLLSSTLA            [N AND D] J-REGION [ ] DS (122-219) (E2)            AETAVYLRISHTAAPPDNGQQLSTYDASDQSPWFLAPSKDTGATLGLMDYHPPTVWAGALTSQHTFPAQLD            [ ]            SLYSLSDVVTIPSSLLDTQYTCNWRKPTKQKYY            [ ]         </p> <p> <a href="#">Sequence in FASTA format</a>   <a href="#">Sequence in IMGT format</a> </p>	
V-DOMAIN	IMGT domain description	VH (1-121) (E1)
	IMGT gene and allele name	IGHV4-59*01 (73.00%)(Human)   <a href="#">Alignment details</a>
	IMGT gene and allele name	IGHV4*01 (92.90%)(Human)   IGHV4*02 (92.90%)(Human)   IGHV4*03 (92.90%)(Human)   <a href="#">Alignment details</a>
	2D representation	IMGT Collier de Perles on 1 layer   <a href="#">IMGT Collier de Perles on 2 layers</a>
	Contact analysis	<a href="#">Domain contacts (residues)</a>
	CDR-IMGT lengths	[ 8.10.12 ]
	Sheet composition	[ A' B' D' E ] [ A'' C'' F'' G'' ]
	<p>           DVLGDSPLKIPPSCLSLTCTYSQTTFYHNRQPPQGLNGDIDKAAITTTMPSKAVTFLKTSANPFLLSSTLA            [ 1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 20 21 22 23 24 25 26 27 28 29 30 31 32 33 34 35 36 37 38 39 40 41 42 43 44 45 46 47 48 49 50 51 52 53 54 55 56 57 58 59 60 61 62 63 64 65 66 67 68 69 70 71 72 73 74 75 76 77 78 79 80 81 82 83 84 85 86 87 88 89 90 91 92 93 94 95 96 97 98 99 100 101 102 103 104 105 106 107 108 109 110 111 112 113 114 115 116 117 118 119 120 121 122 123 124 125 126 127 128 129 130 131 132 133 134 135 136 137 138 139 140 141 142 143 144 145 146 147 148 149 150 151 152 153 154 155 156 157 158 159 160 161 162 163 164 165 166 167 168 169 170 171 172 173 174 175 176 177 178 179 180 181 182 183 184 185 186 187 188 189 190 191 192 193 194 195 196 197 198 199 200 201 202 203 204 205 206 207 208 209 210 211 212 213 214 215 216 217 218 219 ]         </p> <p>           LSEHTAADIWYTCARISHTAAPPDNGQQLSTYDASDQSPWFLAPSKDTGATLGLMDYHPPTVWAGALTSQHTFPAQLD            [ ]         </p> <p> <a href="#">IMGT/DomainGapAlign results</a> </p>	
C-DOMAIN	IMGT domain description	CH1 (122-219) (E2)
	IMGT gene and allele name	IGHG1*01 (100.00%)(Human)   IGHG1*02 (100.00%)(Human)   IGHG1*03 (99.99%)(Human)   <a href="#">Alignment details</a>
	2D representation	IMGT Collier de Perles on 1 layer   <a href="#">IMGT Collier de Perles on 2 layers</a>
	Contact analysis	<a href="#">Domain contacts (residues)</a>
	Sheet composition	[ A' B' D' E ] [ F'' G'' ]
		<p>           ...ASTKPSDFPLAPDQASTL...GATLALGLMDYHP...SPYTVWAGALTS...DWTTPWGLDS...GLYSLSQVTV            PSSLL...DTQYTCNWRKPT...SNTKQKYY            [ ]         </p> <p> <a href="#">IMGT/DomainGapAlign results</a> </p>

**FIGURE 8.** IMGT/3Dstructure-DB card. “IMGT/3Dstructure-DB card” is available for each entry of the database. “Chain details” section for the VH-CH1 chain (1bey\_H) of the alemtuzumab Fab is shown. Chains and domains are described with standardized IMGT labels. Similar result displays are provided for IMGT/2Dstructure-DB cards. However, in those cases information on experimental structural data (hydrogen bonds in IMGT Collier de Perles on two layers, Domain contacts) are only available in the corresponding IMGT/3Dstructure-DB cards, if the antibodies have been crystallized.

- IMGT gene and allele name with the percentage of identity for the V (for example IGHV4-59\*01 [73.00%][Human])
- IMGT gene and allele name with the percentage of identity for the J (for example IGHJ4\*01 [92.90%][Human]) as well as other alleles giving the same percentage of identity
- For V domain: CDR-IMGT lengths (for example [8.10.12]), Sheet composition (for example [A'BDE][A''CC''FG]), the domain AA sequence with CDR-IMGT delimitations and highlighting of AA that are different from the closest V and J genes and alleles
- 2D representation (Step 31)
- Contact analysis (Step 32)

31. For more information, click on:

- “Sequence in FASTA format” or “Sequence in IMGT format,” to access the chain sequence in FASTA or IMGT format.
- “Alignment details” to access the alignment with the closest identified gene and allele name for each domain.
- “IMGT Collier de Perles” (on one layer or two layers for V and C) to get the IMGT Collier de Perles for each domain (**IMGT/Collier de Perles: IMGT Standardized Representation of Domains (IG, TR, and IgSF Variable and Constant Domains, MH and MhSF Groove Domains)**) (Ehrenmann et al. 2011).
- “IMGT/DomainGapAlign results” to get the IMGT/DomainGapAlign results for each domain (**IMGT/DomainGapAlign: IMGT Standardized Analysis of Amino Acid Sequences of**

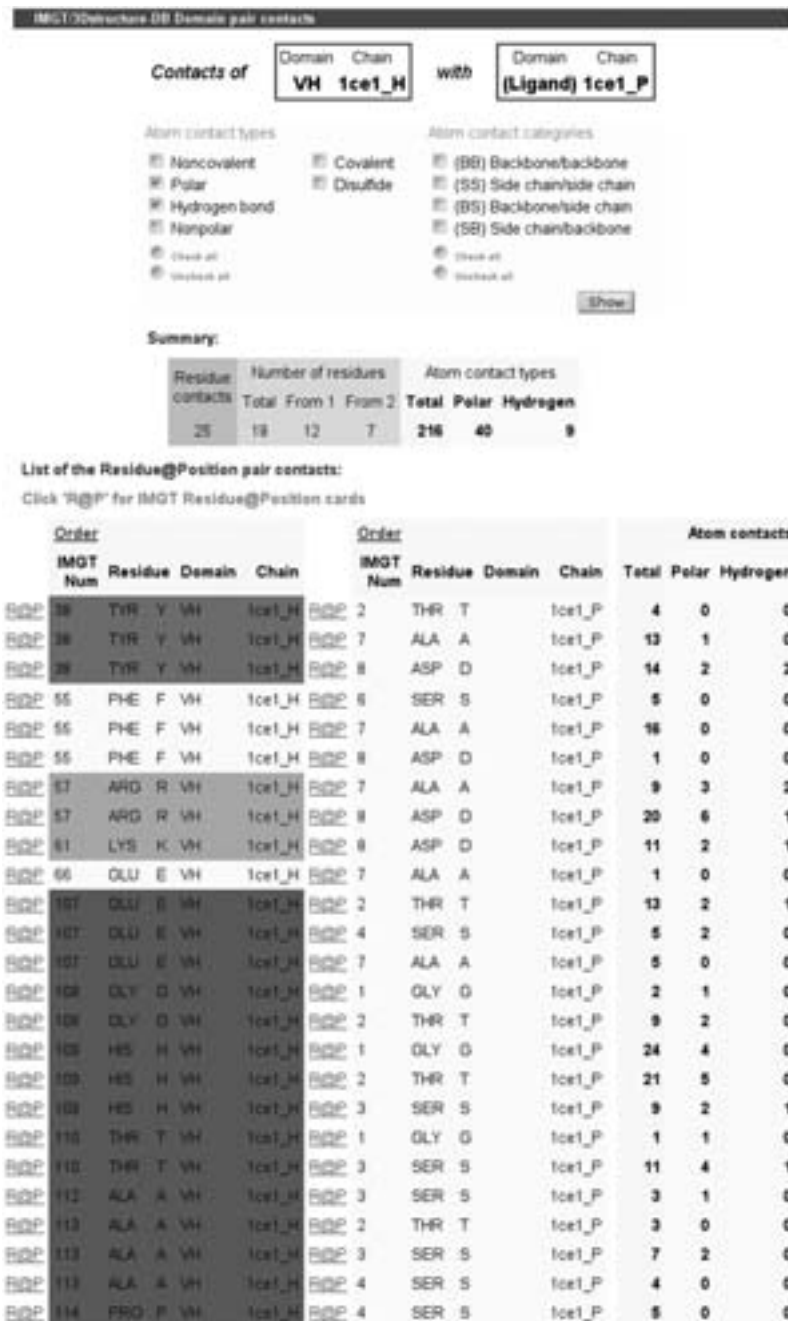


Variable, Constant, and Groove Domains (IG, TR, MH, IgSF, MhSF)) (Ehrenmann and Lefranc 2011).

IMGT/3Dstructure-DB Card-Contact Analysis

32. In the IMGT/3Dstructure-DB card, click on “Domain contacts (overview).”

*This gives access to “IMGT/3Dstructure-DB Domain pair contacts (overview).”*



**FIGURE 9.** IMGT/3Dstructure-DB Domain pair contacts between the VH domain of alemtuzumab (1ce1\_H) and the CD52 peptide ligand (1ce1\_P). “Polar” and “Hydrogen bonds” were selected prior to display, in “Atom contact types.” Amino acids belonging to the CDR1-IMGT, CDR2-IMGT, and CDR3-IMGT are colored in red, orange, and purple, respectively. Positions 55 and 66 are anchor positions. Clicking on R@P gives access to the IMGT Residue@Position cards.

33. Locate “Atom contact types” (noncovalent, polar, hydrogen bond, etc.) and “Atom contact categories” ((BB) Backbone/backbone, (SS) Side chain/side chain, etc.) and select those you want to analyze.
34. Select the domain pair for which you want to analyze contacts by clicking on the corresponding “DomPair” (for instance between VH and ligand).  
*This gives access to the “IMGT/3Dstructure-DB Domain pair contacts” (e.g., VH domain of 1ce1\_H with the ligand 1ce1\_P) (Fig. 9).*
35. Select the “IMGT Residue@Position card” that provides the contacts of a residue at a given position in a domain by clicking on the corresponding “R@P.”  
*This gives access to the IMGT Residue@Position card.*  
*A Residue@Position, for example: 28 – PHE (F) – VH – 1ce1\_H, is defined by the IMGT position numbering, the residue name, the IMGT domain description, and the IMGT chain ID. The IMGT Residue@Position card provides, for a given residue at a given position, structural information, and contacts classified by atom contact types (noncovalent, polar, hydrogen bond, etc.) and atom contact categories ([BB] Backbone/backbone, [SS] Side chain/side chain, etc).*  
*The IMGT Residue@Position cards can also be accessed directly from the amino acid sequences of the IMGT/3Dstructure-DB card (Step 29 in this article; Kaas et al. 2004) or from the IMGT Colliers de Perles (Ruiz and Lefranc 2002); IMGT/Collier de Perles: IMGT Standardized Representation of Domains (IG, TR, and IgSF Variable and Constant Domains, MH and MhSF Groove Domains) (Ehrenmann et al. 2011), by clicking on one AA.*

## DISCUSSION

IMGT/3Dstructure-DB data are particularly useful in antibody engineering and humanization design (Lefranc 2009; Ehrenmann et al. 2010a). Indeed, they allow one to precisely analyze and to easily compare amino acid sequences of the framework regions (FR-IMGT) and complementarity determining regions (CDR-IMGT), between the nonhuman (mouse, rat...) and the closest human V domains. A recent analysis performed on humanized antibodies used in oncology underlines the importance of a correct delimitation of the CDR to be grafted (Magdelaine-Beuzelin et al. 2007). IMGT/3Dstructure-DB facilitates the identification of potential immunogenic residues at given positions in chimeric or humanized antibodies, including those of the constant domains (Jefferis and Lefranc 2009). Moreover, the low potential immunogenicity of nonhuman primate antibodies and their potential use as therapeutics in humans has been shown by this approach. This is particularly important for nonhuman primate antibodies neutralizing *Bacillus anthracis* and the anthrax lethal toxin or the ricin toxin that could be used as therapeutics in ricin intoxication and against bioweapons (Pelat et al. 2008, 2009). These therapeutic applications emphasize the importance of the IMGT/3Dstructure-DB standardized approach that bridges the gap between sequences and 3D structures, whatever the species (Kaas and Lefranc 2007; Lefranc et al. 2008).

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